

(19)



Europäisches Patentamt

European Patent Office

Office européen des brevets



(11)

**EP 1 092 764 A2**

(12)

## EUROPEAN PATENT APPLICATION

(43) Date of publication:

**18.04.2001 Bulletin 2001/16**

(51) Int. Cl.<sup>7</sup>: **C12M 1/36**

(21) Application number: **00121663.9**

(22) Date of filing: **04.10.2000**

(84) Designated Contracting States:

**AT BE CH CY DE DK ES FI FR GB GR IE IT LI LU  
MC NL PT SE**

Designated Extension States:

**AL LT LV MK RO SI**

(30) Priority: **11.10.1999 EP 99120289**

**08.09.2000 EP 00119676**

(71) Applicant:

**F. HOFFMANN-LA ROCHE AG  
4070 Basel (CH)**

(72) Inventors:

- **Bartok, Attila**  
**8002 Zürich (CH)**

• **Müh, Thorsten**

**51375 Leverkusen (DE)**

• **Rückel, Markus**

**82377 Penzberg (DE)**

(74) Representative:

**Müller, Ingrid, Dr. et al  
Grenzacherstrasse 124  
CH-4070 Basel (CH)**

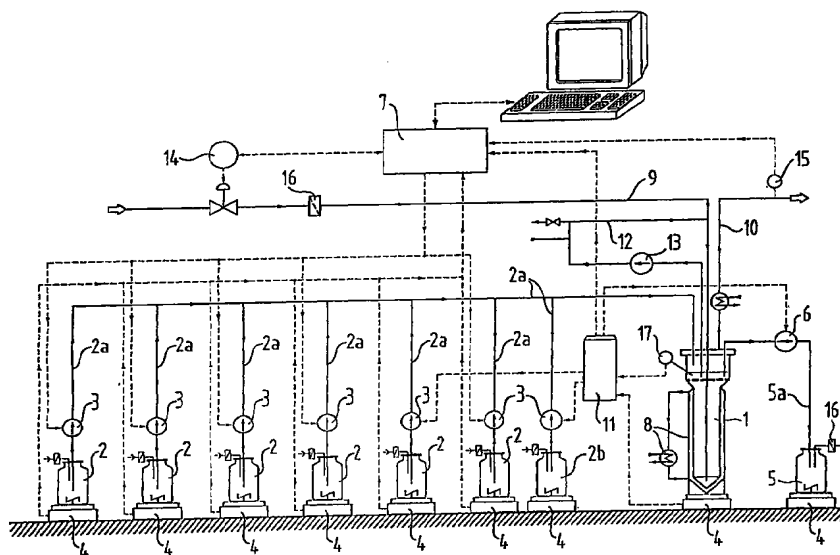
### Remarks:

The sequence listing, which is published as annex to the application documents, was filed after the date of filing. The applicant has declared that it does not include matter which goes beyond the content of the application as filed.

### (54) Continuous fermentation process

(57) The invention is concerned with a continuous process for the manufacture of proteins by means of protein-producing microorganism in which process the microorganism is optionally immobilized on a solid carrier and/or the nutrients and other agents required for the growth of the microorganism and the optimal pro-

duction of protein are fed into the reactor individually at a constant dilution rate. Furthermore, the invention is concerned with a process for the manufacture of proteins using a fermentation assembly.



EP 1 092 764 A2

## Description

[0001] The present invention relates to a continuous process for the manufacture of proteins.

[0002] In accordance with the present invention it has been found that splitting of cultivation media used in a continuous fermentation process allows to study the influence on growth and metabolite-production of microorganisms and thus to determine optimal conditions for the fermentation process. A continuously delivered fermentation medium can generally be split into as many fractions as it contains ingredients. Examples of such ingredients are carbon, nitrogen, phosphorus and sulfur sources as well as vitamins and complex substrates such as corn steep, yeast extract and other natural products. Furthermore, every required mineral, micro- or trace element can be provided separately as a solution of a water-soluble salt, such as a chloride, sulfate or nitrate. In this manner a fermentation medium of any desired composition can be obtained, provided that the desired amounts of the ingredients are (water)-soluble and no disturbing interactions (e.g., precipitation, reaction) occur in the individual feed solutions or in the fermentation medium.

[0003] In one aspect, the present invention is concerned with a continuous process for the manufacture of proteins by means of protein-producing microorganism.

[0004] More particularly, the invention is concerned with a continuous process for the manufacture of proteins by means of protein-producing microorganism in which process the microorganism is optionally immobilized on a solid carrier and/or the nutrients and other agents required for the growth of the microorganism and the optimal production of protein are fed into the reactor individually at a constant dilution rate.

[0005] In a preferred aspect, the invention is concerned with a process for the manufacture of proteins using a fermentation assembly that comprises

a vessel suitable for carrying out reactions involving living or inactivated cells;

at least two storage flasks connected to said vessel for supply of liquids and means to transport said liquids from said storage flasks to said vessel;

individual appliances monitoring the supply of the contents of said storage flasks to said vessel;

a harvest flask connected to said vessel and means to transport fermentation broth from said vessel to said harvest flask; and

a device for controlling and maintaining a constant dilution rate in said vessel with varying rates of individual supply of liquid from said storage flasks to said vessel.

[0006] Any conventional fermentation vessel can be used for the purpose of this invention. The vessel may be made of materials such as stainless steel, glass or ceramics and may have a volume of from e.g., 100 ml to 2500 m<sup>3</sup> although these figures are not critical to the invention. For continuous operation the inside of the vessel is optionally equipped with, e.g., a receptacle or sieve plate for uptake of immobilized cells. Further, the fermentation vessel is connected to a series of storage flasks that contain nutrient solutions and solutions for maintaining and controlling a desired pH and other parameters, such as foam formation, redox potential etc. in the fermentation broth. Depending on the particular needs of the fermentation, there may be separate storage flasks for individual supply of substrates that serve as carbon or nitrogen or mineral source for the living cells.

[0007] It has been found in accordance with the invention that the process is advantageously carried out at a constant dilution rate in the fermentation vessel. As used herein, the term "dilution rate" denotes the total volume of liquids supplied to the fermentation vessel per volume of the fermentation vessel per hour [h<sup>-1</sup>].

[0008] Accordingly, it is a particular feature of the present invention to carry out the fermentation process at a constant dilution rate in the fermentation vessel while varying the supply of individual nutrient components or other additives during the fermentation process. To facilitate this task a storage flask containing an inert component, e.g., water is optionally provided that allows to complement the supply of liquids thus keeping the total supply of liquid constant.

[0009] The assembly that is preferably used to carry out the process of this invention further comprises means to transport the individual components of the fermentation medium from the storage flasks to the fermentation vessel, and appliances for monitoring the amount of liquid supplied to the fermentation vessel. Every combination of measuring instruments (e.g., volumetric or mass flow rate by either gravimetric, anemometric, magnetic, ultrasonic, Venturi, J, cross-relation, thermal, Coriolis, radiometric) and transfer units (e.g., pumps or pressure difference) can be used for this purpose. Additionally, every transfer unit can be applied as a dosing unit (e.g., gear, peristaltic, piston, membrane or excenter pump). For operation on small scale the supply is suitably monitored by weighing the storage flasks that contain nutrient or additive solutions in a predetermined concentration.

[0010] The device for controlling and maintaining a constant dilution rate in the fermentation vessel is suitably a sys-

tem comprising a measuring instrument that monitors the flow from the storage flasks and a controlling unit, e.g., a computer-software control that calculates the actual mass flow rates, compares them to the desired value and adjusts the pump setting accordingly. An appropriate system is, e.g., the Process Automation System, National Instruments, Bridge View, USA, for Windows NT 4.0 (represented by National Instruments, Sonnenbergstrasse 53, 5408 Ennetbaden, Switzerland) that is connected to the various operating units (scales, pumps) through a serial-interface box (Rocket Port, Control Europe Ltd, Great Britain, represented by Technosoft AG Rothackerstrasse 13, 5702 Nied-  
 5 erlenz, Switzerland).

[0011] An assembly that can be used in the process of this invention is depicted in Figure 1.

[0012] The fermentation vessel 1 (Fermentor) is equipped with inlet tubes 2a from storage flasks 2 (suitably  
 10 equipped with a stirrer) for supply of salt solution (Salts), nutrient solution (Nutrients), particular substrates (Substrate 1 and Substrate 2) for supply of, e.g., distinct carbon sources, agent for controlling the pH (Base), water for controlling a constant dilution rate, and antifoam. Pumps 3 transport liquids from the storage flasks 2 to the fermentor 1. Scales 4 monitor the amount of liquids supplied to and discharged from the fermentor. Further, the fermentor has inlet tubes 9 for oxygen supply and outlet tubes 10 for exhaust controlled by units 14 and 15. Pump 6 discharges fermentation broth  
 15 via outlet tubes 5a to a harvest flask 5. A main controlling unit 7 monitors and steers the overall process. Controlling unit 11 monitors and steers individual control systems 17 for temperature, pH, gas pressure, fermentor content and supply of antifoam agents. Circuit 12 including pump 13 is used for taking samples from the fermentation broth and for providing a controlled gas flow for moving the fermentation broth. Inlet and outlet gas flow is controlled by flow control 14 and 15. Sterile filters 16 are provided optionally. Optionally, the fermentation vessel 1 is equipped with a thermostating unit  
 20 8.

[0013] In the process of the present invention, any protein-producing microorganism either natural, e.g. fungal origin or bacterial origin or microorganisms which have been transformed by protein encoding DNA whereby such transformed microorganisms can be bacteria or fungi or yeasts, preferably from the genus *Peniophora*, *Aspergillus*,  
 25 *Hansenula* or *Pichia*, especially *Aspergillus niger*, *Aspergillus awanari*, *Aspergillus sojae*, *Aspergillus oryzae* or *Hansenula polymorpha* or *Pichia pastoris*.

[0014] In this context, the skilled person in the art selects such a protein-producing microorganism which is known to be useful for the production of a desired protein.

[0015] In a preferred embodiment of the present invention the protein is selected from the group consisting of proteins having the activity of an enzyme such as catalase, lactase, phenoloxidase, oxidase, oxidoreductase, glucanase  
 30 cellulase, xylanase and other polysaccharide, peroxidase, lipase, hydrolase, esterase, cutinase, protease and other proteolytic enzymes, aminopeptidase, carboxypeptidase, phytase, lyase, pectinase and other pectinolytic enzymes, amylase, glucosidase, mannosidase, isomerase, invertase, transferase, ribonuclease, chitinase, and desoxyribonuclease. Furthermore, in a preferred embodiment of the present invention the protein is selected from the group of therapeutic proteins such as antibodies, vaccines, antigens, or of antibacterial and/or health-beneficial proteins such as  
 35 lactoternin, lactoperoxidase or lysozyme.

[0016] It will be understood by those skilled in the art that the term "activity" includes not only native activities referring to naturally occurring enzymes or therapeutic functions, but also those activities or functions which have been modified by amino acid substitutions, deletions, additions, or other modifications which may be made to enhance or modify the desired activity, or the thermostability, pH tolerance and/or further properties.

[0017] In a most preferred embodiment of the invention the selected protein is a protein having the activity of a phytase.  
 40

[0018] Examples of proteins having the activity of a phytase are described in EP 684 313, EP 897 010, EP 897 985 or in Examples 6 to 16 and Figures 2 - 22 of the present invention.

45 Figure 2: Design of the consensus phytase sequence. The letters represent the amino acid residues in the one-letter code. The following sequences were used for the alignment: *phyA* from *Aspergillus terreus* 9A-1 (Mitchell et al, 1997; from amino acid (aa) 27), *phyA* from *A. terreus* cbs116.46; (van Loon et al., 1998; from aa 27), *phyA* from *Aspergillus niger* var. *awamori* (Piddington et al, 1993; from aa 27), *phyA* from *A. niger* T213; Mitchell et al. 1997 from aa 27), *phyA* from *A. niger* strain NRRL3135 (van Hartingsveldt et al, 1993; from aa 27), *phyA* from *Aspergillus fumigatus* ATCC 13073 (Pasamontes et al, 1997; from aa 25), *phyA* from *A. fumigatus* ATCC 32722 (EP 897 985; Figur 1; from aa 27), *phyA* from *A. fumigatus* ATCC 58128 (EP 897 985; Figur 1; from aa 27), *phyA* from *A. fumigatus* ATCC 26906 (EP 897 985; Figur 1; from aa 27), *phyA* from *A. fumigatus* ATCC 32239 (EP 897 985; Figur 1; from aa 30), *phyA* from *Emmericella nidulans* (Pasamontes et al, 1997a; from aa 25), *phyA* from *Talaromyces thermophilus* (Pasamontes et al, 1997a; from aa 24), and *phyA* from *Myceliophthora thermophila* (Mitchell et al, 1997; from aa 19). The alignment was calculated using the program PILEUP. The location of the gaps was refined by hand. Capitalized amino acid residues in the alignment at a given position belong to the amino acid coalition that establish the consensus residue. In bold, beneath the calculated consensus sequence, the amino acid sequence of the finally constructed consensus phytase (Fcp) is shown. The gaps in the calculated consensus  
 55

sequence were filled by hand according to principals stated in Example 6.

**Figure 3:** DNA sequence of the consensus phytase-1 gene (*fcp*) and of the primers used for the gene construction. The calculated amino acid sequence (Figure 2) was converted into a DNA sequence using the program BACK-TRANSLATE (Devereux *et al.*, 1984) and the codon frequency table of highly expressed yeast genes (GCG program package, 9.0). The signal peptide of the phytase from *A. terreus* cbs.116.46 was fused to the *N*-terminus. The bold bases represent the sequences of the oligonucleotides used to generate the gene. The names of the respective oligonucleotides are alternately noted above or below the sequence. The underlined bases represent the start and stop codon of the gene. The bases written in italics show the two introduced *Eco* RI sites.

**Figure 4:** Alignment and consensus sequence of five *Basidiomycetes* phytases. The letters represent the amino acid residues in the one-letter code. The amino acid sequences of the phytases from *Paxillus involutus*, phyA1 (aa 21) and phyA2 (aa 21, WO 98/28409), *Trametes pubescens* (aa 24, WO 98/28409), *Agrocybe pediades* (aa 19, WO 98/28409), and *Peniophora lycii* (aa 21, WO 98/28409) starting with the amino acid residues mentioned in parentheses, were used for the alignment and the calculation of the corresponding consensus sequence called "Basidio" (Example 7). The alignment was performed by the program PILEUP. The location of the gaps was refined by hand. The consensus sequence was calculated by the program PRETTY. While a vote weight of 0.5 was assigned to the two *P. involutus* phytases, all other genes were used with a vote weight of 1.0 for the consensus sequence calculation. At positions, where the program was not able to determine a consensus residue, the Basidio sequence contains a dash. Capitalized amino acid residues in the alignment at a given position belong to the amino acid coalition that establish the consensus residue.

**Figure 5:** Design of consensus phytase-10 amino acid sequence. Adding the phytase sequence of *Thermomyces lanuginosus* (Berka *et al.*, 1998) and the consensus sequence of the phytases from five *Basidiomycetes* to the alignment of Figure 2, an improved consensus sequence was calculated by the program PRETTY. Additionally, the amino acid sequence of *A. niger* T213 was omitted; therefore, a vote weight of 0.5 was used for the remaining *A. niger* phytase sequences. For further information see Example 8.

**Figure 6:** DNA and amino acid sequence of consensus phytase-10. The amino acid sequence is written above the corresponding DNA sequences using the one-letter code. The sequence of the oligonucleotides which were used to assemble the gene are in bold letters. The labels of oligonucleotides and the amino acids which were changed compared to those for consensus phytase -1 are underlined. The *fcp10* gene was assembled from the following oligonucleotides: CP-1, CP-2, CP-3.10, CP-4.10, CP-5.10, CP-6, CP-7.10, CP-8.10, CP-9.10, CP-10.10, CP-11.10, CP-12.10, CP-13.10, CP-14.10, CP-15.10, CP-16.10, CP-17.10, CP18.10, CP-19.10, CP-20.10, CP-21.10, CP-22.10. The newly synthesized oligonucleotides are additionally marked by number 10. The phytase contains the following 32 exchanges relative to consensus phytase -1: Y54F, E58A, D69K, D70G, A94K, N134Q, I158V, S187A, Q188N, D197N, S204A, T214L, D220E, L234V, A238P, D246H, T251N, Y259N, E267D, E277Q, A283D, R291I, A320V, R329H, S364T, I366V, A379K, S396A, G404A, Q415E, A437G, A463E. The mutations accentuated in bold letters revealed a stabilizing effect on consensus phytase-1 when tested as single mutations in consensus phytase-1.

**Figure 7:** Alignment for the design of consensus phytase-11. In contrast to the design of consensus phytase-10, for the design of the amino acid sequence of consensus phytase-11, all *Basidiomycete* phytases were used as independent sequences using an assigned vote weight of 0.2 for each *Basidiomycete* sequence. Additionally, the amino acid sequence of *A. niger* T213 phytase was used in that alignment, again.

**Figure 8:** DNA and amino acid sequence of consensus phytase-1-thermo[8]-Q50T-K91A. The amino acid sequence is written above the corresponding DNA sequence using the one-letter code. The replaced amino acid residues are underlined. The stop codon of the gene is marked by a star (\*).

**Figure 9:** DNA and amino acid sequence of consensus phytase-10-thermo[3]-Q50T-K91A. The amino acid sequence is written above the corresponding DNA sequence using the one-letter code. The replaced amino acid residues are underlined. The stop codon of the gene is marked by a star (\*).

**Figure 10:** DNA and amino acid sequence of *A. fumigatus* ATCC 13073 phytase  $\alpha$ -mutant. The amino acid sequence is written above the corresponding DNA sequence using the one-letter code. The replaced amino acid residues are underlined. The stop codon of the gene is marked by a star (\*).

**Figure 11:** DNA and amino acid sequence of consensus phytase-7. The amino acids are written above the corresponding DNA sequence using the one-letter code. The sequences of the oligonucleotides used to assemble the gene are in bold letters. Oligonucleotides and amino acids that were exchanged are underlined and their corresponding triplets are highlighted in small cases. The *fcy7* gene was assembled from the following oligonucleotides: CP-1, CP-2, CP-3, CP-4.7, CP-5.7, CP-6, CP-7, CP-8.7, CP-9, CP-10.7, CP-11.7, CP-12.7, CP-13.7, CP-14.7, CP-15.7, CP-16, CP-17.7, CP-18.7, CP-19.7, CP-20, CP-21, CP-22. The newly synthesized oligonucleotides are additionally marked by number 7. The phytase contains the following 24 exchanges in comparison to the original consensus phytase -1: S89D, S92G, A94K, D164S, P201S, G203A, G205S, H212P, G224A, D226T, E255T, D256E, V258T, P265S, Q292H, G300K, Y305H, A314T, S364G, M365I, A397S, S398A, G404A, and A405S.

**Figure 12:** Differential scanning calorimetry (DSC) of consensus phytase-1 and consensus phytase-10. The protein samples were concentrated to ca. 50-60 mg/ml and extensively dialyzed against 10 mM sodium acetate, pH 5.0. A constant heating rate of 10 °C/min was applied up to 95 °C. DSC of consensus phytase-10 (upper graph) yielded a melting temperature of 85.4 °C, which is 7.3 °C higher than the melting point of consensus phytase-1 (78.1 °C, lower graph).

**Figure 13:** Differential scanning calorimetry (DSC) of consensus phytase-10-thermo[3]-Q50T and consensus phytase-10-thermo[3]-Q50T-K91A. The protein samples were concentrated to ca. 50-60 mg/ml and extensively dialyzed against 10 mM sodium acetate, pH 5.0. A constant heating rate of 10 °C/min was applied up to 95 °C. DSC of consensus phytase-10-thermo-[3]-Q50T (upper graph) yielded a melting temperature of 88.6 °C, while the melting point of consensus phytase-10-thermo[3]-Q50T-K91A was found at 89.3 °C.

**Figure 14:** Comparison of the temperature optimum between consensus phytase-1, consensus phytase-10 and consensus phytase-10-thermo[3]-Q50T. For the determination of the temperature optimum, the phytase standard assay was performed at a series of temperatures between 37 and 86 °C. The diluted supernatant of transformed *S. cerevisiae* strains was used for the determination. The other components of the supernatant showed no influence on the determination of the temperature optimum:  $\Delta$ , consensus phytase-1;  $\diamond$ , consensus phytase-10;  $\blacksquare$ , consensus phytase 10-thermo[3]-Q50T.

**Figure 15:** pH-dependent activity profile and substrate specificity of consensus phytase-10 and its variants thermo[3]-Q50T and thermo[3]-Q50T-K91A. Graph a) shows the pH-dependent activity profile of consensus phytase-10 ( $\square$ ), consensus phytase-10-thermo[3]-Q50T ( $\Delta$ ), and consensus phytase-10-thermo[3]-Q50T-K91A ( $\triangle$ ). The phytase activity was determined using the standard assay in appropriate buffers (see Example 15) at different pH-values. Graph b) shows the corresponding substrate specificity tested by replacement of phytate by the indicated compounds in the standard assay; open bars, consensus phytase-10 (white bars, consensus phytase-10-thermo-Q50T; dark bars, consensus phytase-10-thermo-Q50T-K91A). The numbers correspond to the following compounds: 1, phytate; 2, *p*-nitrophenyl phosphate; 3, phenyl phosphate; 4, fructose-1,6-bisphosphate; 5, fructose-6-phosphate; 6, glucose-6-phosphate; 7, ribose-5-phosphate; 8, DL-glycerol-3-phosphate; 9, glycerol-2-phosphate; 10, 3-phosphoglycerate; 11, phosphoenolpyruvate; 12, AMP; 13, ADP; 14, ATP.

**Figure 16:** pH-dependent activity profile and substrate specificity of consensus phytase-1-thermo[8]-Q50T and of consensus phytase-1-thermo[8]-Q50T-K91A. Graph a) shows the pH-dependent activity profile of the Q50T- ( $\blacksquare$ ) and the Q50T-K91A-variant ( $\triangle$ ). The phytase activity was determined using the standard assay in appropriate buffers (see Example 15) at different pH-values. Graph b) shows the corresponding substrate specificities tested by replacement of phytate by the indicated compounds in the standard assay (open bars, consensus phytase-1-thermo[8]-Q50T; filled bars, consensus phytase-1-thermo[8]-Q50T-K91A). The substrates are listed in the legend of Figure 15.

**Figure 17:** Differential scanning calorimetry (DSC) of consensus phytase-1-thermo[8]-Q50T and consensus phytase-1-thermo[8]-Q50T-K91A. The protein samples were concentrated to ca. 50-60 mg/ml and extensively dialyzed against 10 mM sodium acetate, pH 5.0. A constant heating rate of 10 °C/min was applied up to 95 °C. DSC of consensus phytase-1-thermo[8]-Q50T (upper graph) showed a melting temperature of 84.7 °C, while the melting point of consensus phytase-1-thermo[8]-Q50T-K91A was found at 85.7 °C.

**Figure 18:** Comparison of the temperature optimum between consensus phytase-1, consensus phytase-1-thermo[3] and consensus phytase-1-thermo[8]. For the determination of the temperature optimum, the phytase standard assay was performed at a series of temperatures between 37 and 86 °C. Purified protein from the supernatant of transformed *S. cerevisiae* strains was used for the determination. O, consensus phytase-1;  $\square$ , consensus

phytase-1-thermo [3];  $\Delta$ , consensus phytase-1-thermo[8].

**Figure 19:** Comparison of the pH-dependent activity profile and substrate specificity of consensus phytase-1, consensus phytase-7, and of the phytase from *A. niger* NRRL 3135. Graph a) shows the pH-dependent activity profile of consensus phytase-1 ( $\blacksquare$ ), the phytase from *A. niger* NRRL 3135 (O), and of consensus phytase-7 ( $\Delta$ ). The phytase activity was determined using the standard assay in appropriate buffers (see Example 15) at different pH-values. Graph b) shows the corresponding substrate specificity tested by replacement of phytate by the indicated compounds in the standard assay (black bars, *A. niger* NRRL 3135 phytase; grey bars, consensus phytase-1, dashed bars, consensus phytase-7). The substrates are listed in the legend of Figure 15.

**Figure 20:** Differential scanning calorimetry (DSC) of the phytase from *A. fumigatus* ATCC 13073 and of its stabilized  $\alpha$ -mutant, which contains the following amino acid exchanges: F55Y, V100I, F114Y, A243L, S265P, N294D.

The protein samples were concentrated to ca. 50-60 mg/ml and extensively dialyzed against 10 mM sodium acetate, pH 5.0. A constant heating rate of 10 °C/min was applied up to 95 °C. DSC of consensus *A. fumigatus* 13073 phytase (lower graph) revealed a melting temperature of 62.5 °C, while the melting point of the  $\alpha$ -mutant was found at 67.0 °C.

**Figure 21:** Comparison of the temperature optimum of *A. fumigatus* 13073 wild-type phytase, its  $\alpha$ -mutant, and a further stabilized  $\alpha$ -mutant (E59A-S126N-R329H-S364T-G404A). For the determination of the temperature optimum, the phytase standard assay was performed at a series of temperatures between 37 and 75 °C. The diluted supernatants of transformed *S. cerevisiae* strains were used for the determination. The other components of the supernatant showed no influence on the determination of the temperature optimum. O, *A. fumigatus* ATCC 13073 phytase;  $\Delta$ , *A. fumigatus* ATCC 13073  $\alpha$ -mutant;  $\square$ , *A. fumigatus* ATCC 13073 alpha-mutant-(E59A-S126N-R329H-S364T-G404A)-Q27T;  $\blacksquare$ , *A. fumigatus* ATCC 13073  $\alpha$ -mutant-(E59A-S126N-R329H-S364T-G404A)-Q27T-K68A. The mutations Q51T and K92A in the *A. fumigatus*  $\alpha$ -mutants correspond to -1 Q50T and K91A in consensus phytase, respectively.

**Figure 22:** Amino acid sequence of consensus phytase -12 (consphy12) which contains a number of active site residues transferred from the "basidio" consensus sequence to consensus phytase-10-thermo[3]-Q50T-K91A.

**[0019]** The culture medium used in the fermentation process in accordance with the present invention usually contains nutrients for the cells or microorganisms such as digestible nitrogen sources and inorganic substances, vitamins, micro- and trace elements and other growth-promoting factors. In addition, the culture medium contains a carbon source. Various organic or inorganic substances may be used as nitrogen sources in the fermentation process in accordance with the present invention, such as nitrates, ammonium salts, yeast extract, meat extract, peptone, casein, cornsteep liquor, amino acids and urea. Typical inorganic substances that can be used in the fermentation are calcium, iron, zinc, nickel, manganese, cobalt, copper, molybdenum, and alkali salts such as chlorides, sulphates and phosphates as well as boric acid. As a carbon source, glycerol or sugar-like mono-, di-, oligo- or polysaccharides, e.g., glucose, fructose, sucrose, maltose, starch, glycogen, cellulose or substrates containing such substances, e.g., molasses, glucose syrups and fructose syrups can be used. The concentration of glucose and / or methanol in the total feed stream may vary from about 10 to about 500 g/l for each component and is preferably from about 200 to about 300 g/l. While the fermentation medium is principally an aqueous medium such medium may contain organic solvents such as alcohols, e.g. methanol, ethanol or isopropanol. Further, the fermentation medium may also be a dispersion or suspension, in which case the fermentation is suitably carried out with stirring.

**[0020]** For continuous operation, the cells are optionally immobilized on a solid porous carrier. Any solid porous carrier with any porosity, size and geometry conventionally used in fermentation processes and exerting no toxic effects on the particular cell or microorganism which is to be immobilized can be used for the purpose of this invention. Examples of such carriers are those made from inorganic material and having a pore diameter of from about 0.5 to about 100  $\mu$ m, preferably from about 10 to about 30  $\mu$ m diameter. Examples of inorganic materials are ceramics and natural minerals such as steatite, zeolite, bentonite, silicates (glasses), aluminum silicates, aluminum oxide, magnesium aluminum silicates and magnesium aluminum oxides. Such carriers are commercially available, e.g., from Ceramtec, Marktreidwitz, Germany, Schott Engineering GmbH, Mainz, Germany and others. Preferably, the carriers are spherical with a mean diameter of from about 0,2 to about 20 mm diameter. The carriers can be loaded with the living cells in a manner known per se by contacting the carrier particles with an appropriate cell culture. If desired, the carrier particles loaded with the cells can be further processed by applying a membrane-type coating layer, such as described in German Offenlegungsschrift DE 3421049. Suitably, the carrier is present in the fermentation vessel on a fixed bed. Further, the culture medium, its components and their containments, respectively are suitably sterilized prior to use if autosterilization (e.g., by methanol, ethanol, ammonia) cannot be guaranteed. Heat sterilization with steam (e.g., at 121°C and 1 bar pressure

during 20 minutes) and filtration (0.2 µm) for sensitive components are preferred. Alternative sterilization methods may be applied. Media components need not necessarily be sterilized when running the process in continuous mode.

[0021] Depending on the particular cell or organism used the fermentation may be carried out at a pH between about 2 and about 11. In a preferred aspect of the invention, the fermentation process for the manufacture of phytase is carried out using the microorganism, *Hansenula polymorpha* transformed by a phytase encoding DNA sequence as described in EP 897 010, EP 897 985, or Example 11 of the present case. According to that particular aspect of the invention, the preferred carbon source is a mixture of glucose and methanol. Further, in accordance with that particular aspect of the invention, the fermentation may be carried out at a pH between about 4 and 5, preferably at about pH 4.6. A preferred temperature range for carrying out such fermentation process is between about 10 and 50 °C, more preferably the fermentation temperature is about 30 °C. The aeration rate is preferably adjusted to between about 0.01 and about 1.5 volume of gas per volume of liquid with a dissolved oxygen concentration (DO) of in between 0.01 and about 500 %. A DO of 100 % denotes oxygen saturation of the solution at atmospheric pressure (1 bar) and reactor temperature. The fermentation can be carried out at a pressure of from about 0.1 to about 100 bar, preferably, the fermentation is carried out at atmospheric pressure, i.e., at about 1 bar. The dilution rate can vary from about 0.001 to about 0.5 per hour.

[0022] The invention is illustrated further by the Examples given below.

#### Example 1

[0023] Storage solutions for feed medium were prepared as follows:

##### 1.1 CaCl<sub>2</sub>/H<sub>3</sub>BO<sub>3</sub> Solution

[0024]

CaCl <sub>2</sub> • 2 H <sub>2</sub> O	18.75	g/l
H <sub>3</sub> BO <sub>3</sub>	0.0125	g/l

[0025] This solution was sterilized at 121 °C for 20 minutes.

##### 1.2 Microelements Solution

[0026]

(NH <sub>4</sub> ) <sub>2</sub> Fe(SO <sub>4</sub> ) <sub>2</sub> • 6 H <sub>2</sub> O	2.5	g/l
CuSO <sub>4</sub> • 5 H <sub>2</sub> O	0.2	g/l
ZnSO <sub>4</sub> • 7 H <sub>2</sub> O	0.75	g/l
MnSO <sub>4</sub> • 5 H <sub>2</sub> O	1.0	g/l
Na-EDTA	2.5	g/l

[0027] This solution was sterilized at 121 °C for 20 minutes.

##### 1.3 Trace Elements Solution

[0028]

NiSO <sub>4</sub> • 6 H <sub>2</sub> O	0.025	g/l
--	-------	-----

## EP 1 092 764 A2

(continued)

CoCl <sub>2</sub> • 6 H <sub>2</sub> O	0.025	g/l
Na <sub>2</sub> MoO <sub>4</sub> • 2 H <sub>2</sub> O	0.025	g/l
KJ		

[0029] This solution was sterilized at 121 °C for 20 minutes.

### 1.4 Salts + Vitamin Solution

[0030]

KH <sub>2</sub> PO <sub>4</sub>	50.0	g/l
NH <sub>4</sub> H <sub>2</sub> PO <sub>4</sub>	100.0	g/l
MgSO <sub>4</sub> • 7 H <sub>2</sub> O	45.0	g/l
(NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	50.0	g/l
KCl	23.0	g/l
NaCl	5.0	g/l
vitamin solution (D-biotin, 600 mg/l thiamin • HCl 200 g/l in 50 % isopropanol/water)	5.0	ml/l

[0031] The vitamin solution was sterilized by filtration (0.2 µm) and added to the salt solution that was sterilized at 121 °C for 20 minutes.

### 1.5 Glucose Solution

[0032] 770 g of D-glucose • H<sub>2</sub>O were dissolved in 480 g of water and sterilized (121 °C, 20 min) to yield 1 l solution containing 57 % (by weight) of D-glucose.

### 1.6 Methanol

[0033] Pure methanol was assumed to be sterile and filled into a sterilized flask.

### 1.7 Antifoam

[0034] A sterilized (121 °C, 20 min) solution of 10% antifoam (Struktol J 673, Schill & Seilacher, Hamburg, Germany) was provided for supply on demand by foam-control.

### 1.8 Base

[0035] A solution of ca. 12,5 % (by weight) of ammonia in sterile water was filled into a sterilized flask.

### Example 2

[0036] A fixed bed bioreactor (1 litre) was set up following the principle illustrated in Figure 1 with individual storage flasks being provided for the solutions 1.1 to 1.8 of Example 1. The fixed bed of porous steatite spheres (4 mm diameter, pore diameter 10-30 µm, 280 pores per ml, CeramTec, Marktreidwitz, Germany) was contained by a sieve plate at the top. The reactor was sterilized (121 °C, 20 min) and thereafter filled with an inoculum culture of *Hansenula polymorpha* transformed with a phytase encoding DNA as described, e.g. in EP 897 010, EP 897 985 or Example 11. Then the connection to the storage flasks was established. The inoculum culture was grown on a medium containing glycerol as a carbon source instead of glucose. The reactor was put to batch operation until all glycerol was consumed, which was determined by a rise of the dissolved oxygen concentration. Then the feed stream was turned on and the fermentation



was run under process conditions as given below:

5	Temperature	30	°C
	pH	4.6	Diluted oxygen concentration
	P <sub>total</sub>	10 <sup>5</sup>	N/m <sup>2</sup>
10	P <sub>O2</sub>	10 <sup>5</sup>	N/m <sup>2</sup>
	Dilution rate	0.0067	h <sup>-1</sup>
	aeration rate	100	ml/min
	V <sub>fluid</sub>	1190	ml <sup>-1</sup>
15	V <sub>fixed bed</sub>	950	ml <sup>-1</sup>

[0037] Substrate composition as provided  
by storage flasks 1-8; (actual concentrations  
20 in feed stream given) :

25	D-glucose	305	g/l
	Methanol	264	g/l
	CaCl <sub>2</sub> /H <sub>3</sub> BO <sub>3</sub> Solution	12.2	g/l
	Microelement Solution	20.9	g/l
30	Trace Element Solution	17.2	g/l
	Salts + vitamin Solution	44.7	g/l

Analytics:

35 [0038] Bio-Rad Protein Assay Kit I (Bio-Rad, Glattbrugg, Switzerland) was used to determine the total protein concentration. A factor for the calculation of phytase concentration ( $c_{\text{phyt}}$ ) from total protein concentration ( $c_{\text{tp}}$ ) was determined as  $c_{\text{phyt}} = 0.76 \cdot c_{\text{tp}}$ .

[0039] To determine the biomass in the medium two samples of 1 ml were centrifuged, washed with 1 ml of water,  
40 centrifuged again, dried at 85 °C for two days and weighed.

Results:

[0040] Under the above process conditions the biomass was 59 g/l. Given a dilution rate of 0.0067 per hour the productivity was 0.078 g of phytase per litre per hour.

[0041] In a fermentation that was run fed-batch-wise the biomass was 125 g/l; the productivity, however, was calculated to 0.054 g phytase per litre per hour.

### Example 3

50 [0042] A fermentation in analogy to Example 2 but omitting the steatite spheres (i.e., without immobilisation of the microorganism) was carried out. A nutrient and a salt and vitamin solution of the following composition were pumped into the reactor separately:

55

## EP 1 092 764 A2

Nutrient Solution:

[0043]

5

10

15

20

NiSO <sub>4</sub> • 6 H <sub>2</sub> O	8.33	mg/l
CoCl <sub>2</sub> • 6 H <sub>2</sub> O	8.33	mg/l
Na <sub>2</sub> MoO <sub>4</sub> • 2 H <sub>2</sub> O	8.33	mg/l
KJ	8.33	mg/l
(NH <sub>4</sub> ) <sub>2</sub> Fe(SO <sub>4</sub> ) <sub>2</sub> • 6 H <sub>2</sub> O	833.33	mg/l
CuSO <sub>4</sub> • 5 H <sub>2</sub> O	66.67	mg/l
ZnSO <sub>4</sub> • 7 H <sub>2</sub> O	250	mg/l
MnSO <sub>4</sub> • 5 H <sub>2</sub> O	333.33	mg/l
Na-EDTA	833.33	mg/l
CaCl <sub>2</sub> • 2 H <sub>2</sub> O	6250	mg/l
H <sub>3</sub> BO <sub>3</sub>	4.17	mg/l

Salts + Vitamins Solution:

25

[0044]

30

35

40

KH <sub>2</sub> PO <sub>4</sub>	50.0	g/l
NH <sub>4</sub> H <sub>2</sub> PO <sub>4</sub>	100.0	g/l
MgSO <sub>4</sub> • 7 H <sub>2</sub> O	45.0	g/l
(NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	50.0	g/l
KCl	23.0	g/l
NaCl	5.0	g/l
vitamin solution (D-biotin, 600 mg/l thiamin • HCl 200 g/l in 50 % isopropanol/water)	5.0	ml/l

[0045] The supply of these two solutions was adjusted to provide in the feed stream a concentration of 51 g/l of Nutrient Solution and 61 g/l of Salts + Vitamins Solution. The dilution rate was adjusted to 0.009 h<sup>-1</sup>. The pH was kept at 4.6 by addition of 12.5 wt% ammonium hydroxide.

45 [0046] Furthermore, Glucose Solution as in Example 1 and methanol were fed into the reactor separately to maintain a glucose concentration of 275 g/l and a methanol concentration of 260 g/l in the feed stream.

[0047] The productivity of this fermentation was 0.088 g phytase per litre per hour. Biomass in outflow was 58 g/l.

### Example 4

50

[0048] In a fermentation process in analogy to Example 3 but adjusting glucose concentration to 290 g/l, methanol concentration to 260 g/l, and keeping the dilution rate constant at 0.009 h<sup>-1</sup>, the productivity was 0.092 g phytase per litre per hour. Biomass in outflow was 60.4 g/l.

### 55 Example 5

[0049] In a fermentation process in analogy to Example 3 but adjusting glucose concentration to 270 g/l, methanol concentration to 280 g/l, and keeping the dilution rate constant at 0.009 h<sup>-1</sup>, the productivity was 0.094 g phytase per

litre per hour. Biomass in outflow was 56.8 g/l.

#### Example 6:

#### Design of the amino acid sequence of consensus phytase-1

#### Alignment of the amino acid sequences

**[0050]** The alignment was calculated using the program PILEUP from the GCG Sequence Analysis Package Release 9.0 (Devereux *et al.*, 1984) with the standard parameters (gap creation penalty 12, gap extension penalty 4). The location of the gaps was refined using a text editor. Table 1 shows the sequences (see Figure 2), without the signal sequence, that were used for the performance of the alignment starting with the amino acid (aa) as mentioned in Table 1.

Table 1

Origin and vote weight of the phytase amino acid sequences used for the design of consensus phytase-1

- <i>phyA</i> from <i>Aspergillus terreus</i> 9A-1, aa 27, vote weight 0.5 (Mitchell <i>et al.</i> , 1997)
- <i>phyA</i> from <i>Aspergillus terreus</i> cbs116.46, aa 27, vote weight 0.5 (EP 897 985; Figure 1)
- <i>phyA</i> from <i>Aspergillus niger</i> var. <i>awamori</i> , aa 27, vote weight 0.33 (Piddington <i>et al.</i> , 1993)
- <i>phyA</i> from <i>Aspergillus niger</i> T213, aa 27, vote weight 0.33
- <i>phyA</i> from <i>Aspergillus niger</i> strain NRRL3135, aa 27, vote weight 0.33 (van Hartingsveldt <i>et al.</i> , 1993)
- <i>phyA</i> from <i>Aspergillus fumigatus</i> ATCC 13073, aa 26, vote weight 0.2 (Pasamontes <i>et al.</i> , 1997)
- <i>phyA</i> from <i>Aspergillus fumigatus</i> ATCC 32722, aa 26, vote weight 0.2 (EP 897 985; Figure 1)
- <i>phyA</i> from <i>Aspergillus fumigatus</i> ATCC 58128, aa 26, vote weight 0.2 (EP 897 985; Figure 1)
- <i>phyA</i> from <i>Aspergillus fumigatus</i> ATCC 26906, aa 26, vote weight 0.2 (EP 897 985; Figure 1)
- <i>phyA</i> from <i>Aspergillus fumigatus</i> ATCC 32239, aa 30, vote weight 0.2 (EP 897 985; Figure 1)
- <i>phyA</i> from <i>Emericella nidulans</i> , aa 25, vote weight 1.0 (Pasamontes <i>et al.</i> , 1997a)
- <i>phyA</i> from <i>Talaromyces thermophilus</i> ATCC 20186, aa 24, vote weight 1.0 (Pasamontes <i>et al.</i> , 1997a)
- <i>phyA</i> from <i>Myceliophthora thermophila</i> , aa 19, vote weight 1.0 (Mitchell <i>et al.</i> , 1997)

#### Calculation of the amino acid sequence of consensus phytase-1

**[0051]** Using the refined alignment as input, the consensus sequence was calculated by the program PRETTY from the GCG Sequence Analysis Package Release 9.0 (Devereux *et al.*, 1984). PRETTY prints sequences with their columns aligned and can display a consensus sequence for an alignment. A vote weight that pays regard to the similarity between the amino acid sequences of the aligned phytases was assigned to all sequences. The vote weight was set in such a way that the combined impact of all phytases from one sequence subgroup (same species, but from different strains), e. g. the amino acid sequences of all phytases from *A. fumigatus*, on the election was set one, that means that each sequence contributes with a value of 1 divided by the number of strain sequences (see Table 1). By this means, it was possible to prevent that very similar amino acid sequences, e. g. of the phytases from different *A. fumigatus* strains, dominate the calculated consensus sequence.

**[0052]** The program PRETTY was started with the following parameters: The plurality defining the number of votes below which there is no consensus was set on 2.0. The threshold, which determines the scoring matrix value below which an amino acid residue may not vote for a coalition of residues, was set on 2. PRETTY used the PrettyPep.Cmp consensus scoring matrix for peptides.

**[0053]** Ten positions of the alignment (position 46, 66, 82, 138, 162, 236, 276, 279, 280, 308; Figure 2), for which the program was not able to determine a consensus residue, were filled by hand according to the following rules: if a most frequent residue existed, this residue was chosen (138, 236, 280); if a prevalent group of similar equivalent residues occurred, the most frequent or, if not available, one residue of this group was selected (46, 66, 82, 162, 276, 308). If there was neither a prevalent residue nor a prevalent group, one of the occurring residues was chosen according to

common assumptions on their influence on the protein stability (279). Eight other positions (132, 170, 204, 211, 275, 317, 384, 447; Figure 2) were not filled with the amino acid residue selected by the program but normally with amino acids that occur with the same frequency as the residues that were chosen by the program. In most cases, the slight underrating of the three *A. niger* sequences (sum of the vote weights: 0.99) was eliminated by this correction.

#### Conversion of the consensus phytase-1 amino acid sequence to a DNA sequence

[0054] The first 26 amino acid residues of *A. terreus* cbs116.46 phytase were used as signal peptide and, therefore, fused to the N-terminus of all consensus phytases. For this stretch, we used a special method to calculate the corresponding DNA sequence. Purvis et al (1987) proposed that the incorporation of rare codons in a gene has an influence on the folding efficiency of the protein. The DNA sequence for the signal sequence was calculated using the approach of Purvis et al (1987) and optimized for expression in *S. cerevisiae*. For the remaining parts of the protein, we used the codon frequency table of highly expressed *S. cerevisiae* genes, obtained from the GCG program package, to translate the calculated amino acid sequence into a DNA sequence.

[0055] The resulting sequence of the *fcg* gene is shown in Figure 3.

#### Construction and cloning of the consensus phytase-1 gene

[0056] The calculated DNA sequence of consensus phrase-1 (*fcg*) was divided into oligonucleotides of 85 bp, alternately using the sequence of the sense and the anti-sense strand. Every oligonucleotide overlaps 20 bp with its previous and its following oligonucleotide of the opposite strand. The location of all primers, purchased from Microsynth, Balgach (Switzerland) and obtained in a PAGE-purified form, is indicated in Figure 3.

#### PCR-Reactions

[0057] In three PCR reactions, the synthesized oligonucleotides were composed to the entire gene. For the PCR, the High Fidelity Kit from Boehringer Mannheim (Boehringer Mannheim, Germany) and the thermo cycler The Protokol (TM) from AMS Biotechnology (Europe) Ltd. (Lugano, Switzerland) were used.

[0058] Oligonucleotides CP-1 to CP-10 (Mix 1, Figure 3) were mixed to a concentration of 0.2 pmol/μl of each oligonucleotide. A second oligonucleotide mixture (Mix 2) was prepared with CP-9 to CP-22 (0.2 pmol/μl of each oligonucleotide). Additionally, four short primers were used in the PCR reactions:

CP-a: *Eco* RI  
5'-TATATGAATTCATGGGCGTGTTTCGTC-3' (SEQ ID No. 1)

CP-b: 5'-TGAAAAGTTTCATTGAAGGTTTC-3' (SEQ ID No. 2)

CP-c: 5'-TCTTCGAAAGCAGTACAAGTAC-3' (SEQ ID No. 3)

CP-e: *Eco* RI  
5'-TATATGAATTCTTAAGCGAAAC-3' (SEQ ID No. 4)

PCR reaction a: 10 μl Mix 1 (2.0 pmol of each oligonucleotide)  
2 μl nucleotides (10 mM each nucleotide)  
2 μl primer CP-a (10 pmol/μl)  
2 μl primer CP-c (10 pmol/μl)

## EP 1 092 764 A2

10,0 µl PCR buffer  
0.75 µl polymerase mixture (2.6U)  
73.25 µl H<sub>2</sub>O

5 PCR reaction *b*:  
10 µl Mix 2 (2.0 pmol of each oligonucleotide)  
2 µl nucleotides (10 mM each nucleotide)  
2 µl primer CP-b (10 pmol/µl)  
2 µl primer CP-e (10 pmol/µl)  
10,0 µl PCR buffer  
10 0.75 µl polymerase mixture (2.6 U)  
73.25 µl H<sub>2</sub>O

Reaction conditions for PCR reactions *a* and *b*:  
15 step 1 2 min - 45°C  
step 2 30 sec - 72°C  
step 3 30 sec - 94°C  
step 4 30 sec - 52°C  
step 5 1 min - 72°C

[0059] Steps 3 to 5 were repeated 40-times.

20 [0060] The PCR products (670 and 905 bp) were purified by an agarose gel electrophoresis (0.9% agarose) and a following gel extraction (QIAEX II Gel Extraction Kit, Qiagen, Hilden, Germany). The purified DNA fragments were used for the PCR reaction *c*.

PCR reaction *c*:  
25 6 µl PCR product of reaction *a* (≈50 ng)  
6 µl PCR product of reaction *b* (≈50 ng)  
2 µl primer CP-a (10 pmol/µl)  
2 µl primer CP-e (10 pmol/µl)  
10,0 µl PCR buffer  
30 0.75 µl polymerase mixture (2.6 U)  
73.25 µl H<sub>2</sub>O

Reaction conditions for PCR reaction *c*:  
35 step 1 2 min - 94°C  
step 2 30 sec - 94°C  
step 3 30 sec - 55°C  
step 4 1 min - 72°C

[0061] Steps 2 to 4 were repeated 31-times.

40 [0062] The resulting PCR product (1.4 kb) was purified as mentioned above, digested with *Eco* RI, and ligated in an *Eco* RI-digested and dephosphorylated pBsk(-)-vector (Stratagene, La Jolla, CA, USA). 1 µl of the ligation mixture was used to transform *E. coli* XL-1 competent cells (Stratagene, La Jolla, CA, USA). All standard procedures were carried out as described by Sambrook *et al.* (1987). The DNA sequence of the constructed consensus phytase gene (*fcp*, Figure 3) was controlled by sequencing as known in the art.

### Example 7

45 Design of an improved consensus phytase (consensus phytase-10) amino acid sequence

[0063] The alignments used for the design of consensus phytase-10 were calculated using the program PILEUP from the GCG Sequence Analysis Package Release 9.0 (Devereux *et al.*, 1984) with the standard parameters (gap creation penalty 12, gap extension penalty 4). The location of the gaps was refined using a text editor.

50 [0064] The following sequences were used for the alignment of the *Basidiomycete* phytases starting with the amino acid (aa) mentioned in Table 2:

55

Table 2

Origin and vote weight of five *Basidiomycete* phytases used for the calculation of the corresponding amino acid consensus sequence (basidio)

- *phyA1* from *Paxillus involutus* NN005693, aa 21, vote weight 0.5 (WO 98/28409)
- *phyA2* from *Paxillus involutus* NN005693, aa 21, vote weight 0.5 (WO 98/28409)
- *phyA* from *Trametes pubescens* NN9343, aa 24, vote weight 1.0 (WO 98/28409)
- *phyA* from *Agrocybe pediades* NN009289, aa 19, vote weight 1.0 (WO 98/28409)
- *phyA* from *Peniophora lycii* NN006113, aa 21, vote weight 1.0 (WO 98/28409)

[0065] The alignment is shown in Figure 4.

[0066] In Table 3 the genes, which were used for the performance of the final alignment, are arranged. The first amino acid (aa) of the sequence which is used in the alignment is mentioned behind the organism's designation.

Table 3

Origin and vote weight of the phytase sequences used for the design of consensus phytase 10

- *phyA* from *Aspergillus terreus* 9A-1, aa 27, vote weight 0.5 (Mitchell *et al.*, 1997)
- *phyA* from *Aspergillus terreus* cbs116.46, aa 27, vote weight 0.5 (EP 897 985; Figur 1)
- *phyA* from *Aspergillus niger* var. *awamori*, aa 27, vote weight 0.5 (Piddington *et al.*, 1993)
- *phyA* from *Aspergillus niger* strain NRRL3135, aa 27, vote weight 0.5 (van Hartingsveldt *et al.*, 1993)
- *phyA* from *Aspergillus fumigatus* ATCC 13073, aa 26, vote weight 0.2 (Pasamontes *et al.*, 1997)
- *phyA* from *Aspergillus fumigatus* ATCC 32722, aa 26, vote weight 0.2 (EP 897 985; Figur 1)
- *phyA* from *Aspergillus fumigatus* ATCC 58128, aa 26, vote weight 0.2 (EP 897 985; Figur 1)
- *phyA* from *Aspergillus fumigatus* ATCC 26906, aa 26, vote weight 0.2 (EP 897 985; Figur 1)
- *phyA* from *Aspergillus fumigatus* ATCC 32239, aa 30, vote weight 0.2 (EP 897 985; Figur 1)
- *phyA* from *Emmericella nidulans*, aa 25, vote weight 1.0 (Pasamontes *et al.*, 1997a)
- *phyA* from *Talaromyces thermophilus* ATCC 20186, aa 24, vote weight 1.0 (Pasamontes *et al.*, 1997a)
- *phyA* from *Myceliophthora thermophila*, aa 19, vote weight 1.0 (Mitchell *et al.*, 1997)
- *phyA* from *Thermomyces lanuginosa*, aa 36, vote weight 1.0 (Berka *et al.*, 1998)
- Consensus sequence of five *Basidiomycete* phytases, vote weight 1.0 (Basidio, Figure 4)

[0067] The corresponding alignment is shown in Figure 5.

#### Calculation of the amino acid sequence of consensus phytase-10

[0068] To improve the alignment, we combined the consensus sequence of five phytases from four different *Basidiomycetes*, called Basidio, still containing the undefined sequence positions (see Figure 4), nearly all phytase sequences used for calculation of the original consensus phytase and one new phytase sequence from the *Ascomycete* *Thermomyces lanuginosus* to a larger alignment.

We set plurality on 2.0 and threshold on 3. The used vote weights are listed in Table 3. The alignment and the corresponding consensus sequence are presented in Figure 5. The new consensus phytase -10 sequence has 32 different amino acids in comparison to the original consensus phytase. Positions for which the program PRETTY was not able to calculate a consensus amino acid residue were filled according to rules mentioned in Example 6. None of the residues suggested by the program was replaced.

[0069] Furthermore, we included all *Basidiomycete* phytases as single amino acid sequences but assigning a vote weight of 0.2 in the alignment. The corresponding alignment is shown in Figure 7. The calculated consensus amino acid

sequence (consensus phytase-11) has the following differences to the sequence of consensus phytase-10: D35X, X(K)69K, X(E)100E, A101R, Q134N, X(K)153N, X(H)190H, X(A)204S, X(E)220D, E222T, V227A, X(R)271R, H287A, X(D)288D, X(K)379K, X(I)389I, E390X, X(E)415E, X(A)416A, X(R)446L, E463A, where the numbering is as in Fig. 6.

[0070] Letter X means that the program was not able to calculate a consensus amino acid; the amino acid in parenthesis corresponds to the amino acid finally included into the consensus phytase-10.

[0071] We also checked single amino acid replacements suggested by the improved consensus phytase sequences 10 and 11 on their influence on the stability of the original consensus phytase -1. The approach is described in example 8.

#### 10 Conversion of consensus phytase-10 amino acid sequence to a DNA sequence

[0072] The first 26 amino acid residues of *A. terreus* cbs116.46 phytase were used as signal peptide and, therefore, fused to the *N*-terminus of consensus phytase-10. The used procedure is further described in Example 6.

[0073] The resulting sequence of the *fcy10* gene is shown in Figure 6.

#### 15 Construction and cloning of the consensus phytase-10 gene (*fcy10*)

[0074] The calculated DNA sequence of *fcy10* was divided into oligonucleotides of 85 bp, alternately using the sequence of the sense and the anti-sense strand. Every oligonucleotide overlaps 20 bp with its previous and its following oligonucleotide of the opposite strand. The location of all primers, purchased from Microsynth, Balgach (Switzerland) and obtained in a PAGE-purified form, is indicated in Figure 6.

#### PCR-Reactions

[0075] In three PCR reactions, the synthesized oligonucleotides were composed to the entire gene. For the PCR, the High Fidelity Kit from Boehringer Mannheim (Boehringer Mannheim, Mannheim, Germany) and the thermocycler The Protokol™ from AMS Biotechnology (Europe) Ltd. (Lugano, Switzerland) were used. The following oligonucleotides were used in a concentration of 0.2 pmol/ml.

Mix 1.10: CP-1, CP-2, CP-3.10, CP-4.10, CP-5.10, CP-6, CP-7.10, CP-8.10, CP-9.10, CP-10.10

Mix 2.10: CP-9.10, CP-10.10, CP-11.10, CP-12.10, CP-13.10, CP-14.10, CP-15.10, CP-16.10, CP-17.10, CP-18.10, CP-19.10, CP-20.10, CP-21.10, CP-22.10

[0076] The newly synthesized oligonucleotides are marked by number 10. The phytase contains the following 32 exchanges, which are underlined in Figure 6, in comparison to the original consensus phytase -1: Y54F, E58A, D69K, D70G, A94K, N134Q, I158V, S187A, Q188N, D197N, S204A, T214L, D220E, L234V, A238P, D246H, T251N, Y259N, E267D, E277Q, A283D, R291I, A320V, R329H, S364T, I366V, A379K, S396A, G404A, Q415E, A437G, A463E.

[0077] Four short PCR primers were used for the assembling of the oligonucleotides:

CP-a: *Eco* RI  
5'-TATATGAATTTCATGGGCGTGTTCGTC-3' (SEQ, ID No. 1)

CP-b: 5'-TGAAAAGTTCATTGAAGGTTTC-3' (SEQ, ID No. 2)

CP-c.10: 5'-TCTTCGAAAGCAGTACACAAAC-3' (SEQ, ID No. 5)

CP-e: *Eco* RI  
5'-TATATGAATTCTTAAGCGAAAC-3' (SEQ, ID No. 4)

## EP 1 092 764 A2

PCR reaction *a*:  
10 µl Mix 1.10 (2.0 pmol of each oligonucleotide)  
2 µl nucleotides (10 mM each nucleotide)  
2 µl primer CP-a (10 pmol/ml)  
2 µl primer CP-c.10 (10 pmol/ml)  
10,0 µl PCR buffer  
0.75 µl polymerase mixture (2.6 U)  
73.25 µl H<sub>2</sub>O

PCR reaction *b*:  
10 µl Mix 2.10 (2.0 pmol of each oligonucleotide)  
2 µl nucleotides (10 mM each nucleotide)  
2 µl primer CP-b (10 pmol/ml)  
2 µl primer CP-e (10 pmol/ml)  
10,0 µl PCR buffer  
0.75 µl polymerase mixture (2.6 U)  
73.25 µl H<sub>2</sub>O

Reaction conditions for PCR reactions *a* and *b*:  
step 1 2 min - 45°C  
step 2 30 sec - 72 °C  
step 3 30 sec - 94 °C  
step 4 30 sec - 52 °C  
step 5 1 min - 72°C

[0078] Steps 3 to 5 were repeated 40-times.

[0079] The PCR products (670 and 905 bp) were purified by an agarose gel electrophoresis (0.9% agarose) and a following gel extraction (QIAEX II Gel Extraction Kit, Qiagen, Hilden, Germany). The purified DNA fragments were used for the PCR reaction *c*.

PCR reaction *c*:  
6 µl PCR product of reaction *a*( ≈50 ng)  
6 µl PCR product of reaction *b*( ≈50 ng)  
2 µl primer CP-a (10 pmol/ml)  
2 µl primer CP-e (10 pmol/ml)  
10,0 µl PCR buffer  
0.75 µl polymerase mixture (2.6 U)  
73.25 µl H<sub>2</sub>O

Reaction conditions for PCR reaction *c*:  
step 1 2 min - 94°C  
step 2 30 sec - 94 °C  
step 3 30 sec - 55 °C  
step 4 1 min - 72°C

Steps 2 to 4 were repeated 31-times.

[0080] The resulting PCR product (1.4 kb) was purified as mentioned above, digested with *Eco* RI, and ligated in an *Eco* RI-digested and dephosphorylated pBsk(-)-vector (Stratagene, La Jolla, CA, USA). 1 µl of the ligation mixture was used to transform *E. coli* XL-1 competent cells (Stratagene, La Jolla, CA, USA). All standard procedures were carried out as described by Sambrook *et al.* (1987). The DNA sequence of the constructed gene (*fcp10*) was checked by sequencing as known in the art.

### Example 8

Increasing the thermostability of consensus phytase-1 by introduction of single mutations suggested by the amino acid sequence of consensus phytase-10 and/or consensus phytase-11

[0081] In order to increase the thermostability of homologous genes, it is also possible to test the stability effect of each differing amino acid residue between the protein of interest and the calculated consensus sequence and to combine all stabilizing mutations into the protein of interest. We used the consensus phytase -1 as protein of interest and tested the effect on the protein stability of 34 amino acids, which differed between consensus phytase -1 on one hand and consensus phytases 10 and/or -11 on the other hand, by single mutation..



[0082] To construct muteins for expression in *A. niger*, *S. cerevisiae*, or *H. polymorpha*, the corresponding expression plasmid containing the consensus phytase gene was used as template for site-directed mutagenesis (see Examples 11 - 13). Mutations were introduced using the "quick exchange™ site-directed mutagenesis kit" from Stratagene (La Jolla, CA, USA) following the manufacturer's protocol and using the corresponding primers. All mutations made and their corresponding primers are summarized in Table 4. Plasmids harboring the desired mutation were identified by DNA sequence analysis as known in the art.

**Table 4: Primers used for site-directed mutagenesis of consensus phytases**

(Exchanged bases are highlighted in bold. The introduction of a restriction site is marked above the sequence. When a restriction site is written in parenthesis, the mentioned site was destroyed by introduction of the mutation.)

mutation	Primer set
	<i>Kpn</i> I
Q50T	5'-CACTTGTGGGGTACCTACTCTCCATACTTCTC-3' (SEQ ID No. 6) 5'-GAGAAGTATGGAGAGTAGGTACCCCAACAAGTG-3' (SEQ ID No. 7)
Y54F	5'-GGTCAATACTCTCCATTCTTCTCTTTGGAAG-3' (SEQ ID No. 8) 5'-CTTCCAAAGAGAAGAATGGAGAGTATTGACC-3' (SEQ ID No. 9)
E58A	5'-CATACTTCTCTTTGGCAGACGAATCTGC-3' (SEQ ID No. 10) 5'-GCAGATTCGTCTGCCAAAGAGAAGTATG-3' (SEQ ID No. 11)

5	D69K	<i>Aat</i> II	5'-CTCCAGACGTCCCAAAGGACTGTAGAGTTAC-3' (SEQ ID No. 12)
			5'-GTAAGTCTACAGTCCTTTGGGACGTCTGGAG-3' (SEQ ID No. 13)
10	D70G	<i>Aat</i> II	5'-CTCCAGACGTCCCAGACGGCTGTAGAGTTAC-3' (SEQ ID No. 14)
			5'-GTAAGTCTACAGCCGTCTGGGACGTCTGGAG-3' (SEQ ID No. 15)
15	K91A		5'-GATACCCAACTTCTTCTGCGTCTAAGGCTTACTCTG-3'
			(SEQ ID No. 16)
20	A94K		5'-CAGAGTAAGCCTTAGACGCAGAAGAAGTTGGGTATC-3'
			(SEQ ID No. 17)
25	A101R	<i>Sca</i> I	5'-CTTCTAAGTCTAAGAAGTACTCTGCTTTG-3' (SEQ ID No. 18)
			5'-CAAAGCAGAGTACTTCTTAGACTTAGAAG-3' (SEQ ID No. 19)
30	N134Q		5'-GCTTACTCTGCTTTGATTGAACGGATTCAAAAGAACGCTAC-3'
			5'-GTAGCGTTCTTTTGAATCCGTTCAATCAAAGCAGAGTAAGC-3'
35	K153N		5'-CCATTTCGGTGAACAGCAAATGGTTAACTC-3' (SEQ ID No. 22)
			5'-GAGTTAACCATTGCTGTTACCGAATGG-3' (SEQ ID No. 23)
40	I158V	<i>Nru</i> I	5'-GATACAAGGCTCTCGCGAGAAACATTGTTC-3' (SEQ ID No. 24)
			5'-GGAACAATGTTTCTCGCGAGAGCCTTGTATC-3' (SEQ ID No. 25)
45	D197N	<i>Bss</i> HI	5'-GATTGTTCCATTTCGTGCGCGCTTCTGGTTC-3' (SEQ ID No. 26)
			5'-GAACCAGAAGCGCGCACGAATGGAACAATC-3' (SEQ ID No. 27)
50	S187A	<i>Bcl</i> I	5'-CTCCAGTTATTAACGTGATCATTCCAGAAGG-3' (SEQ ID No. 28)
			5'-CCTTCTGGAATGATCACGTTAATAACTGGAG-3' (SEQ ID No. 29)
55	T214L	<i>Apa</i> I	5'-GGCTGACCCAGGGGCCCAACCACACCAAGC-3' (SEQ ID No. 30)
			5'-GCTTGGTGTGGTTGGGCCCTGGGTGAGCC-3' (SEQ ID No. 31)
60	E222T	<i>Nco</i> I	5'-CACTTTGGACCATGGTCTTTGTAAGTCTGCTTTTCG-3' (SEQ ID No. 32)
			5'-CGAAAGCAGTACAAAGACCATGGTCCAAAGTG-3' (SEQ ID No. 33)
65	34)	<i>Avr</i> II	5'-GCTTTCGAAGACTCTACCCTAGGTGACGACGTTG-3' (SEQ ID No. 34)
			5'-CAACGTCGTCACCTAGGGTAGAGTCTTCGAAAGC-3' (SEQ ID No. 35)

5	V227A	5'-GGTGACGACGCTGAAGCTAACTTCAC-3' (SEQ ID No. 36) 5'-GTGAAGTTAGCTTCAGCGTCGTCACC-3' (SEQ ID No. 37)
		<i>Sac</i> II
	L234V	5'-CTAACTTCACCGCGGTGTTTCGCTCCAG-3' (SEQ ID No. 38) 5'-CTGGAGCGAACACCGCGGTGAAGTTAG-3' (SEQ ID No. 39)
10	A238P 40)	5'-GCTTTGTTTCGCTCCACCTATTAGAGCTAGATTGG-3' (SEQ ID No. 40) 5'-CCAATCTAGCTCTAATAGGTGGAGCGAACAAAGC-3' (SEQ ID No. 41)
15		<i>Hpa</i> I
	T251N	5'-GCCAGGTGTTAACTTGACTGACGAAG-3' (SEQ ID No. 42) 5'-TTCGTCAGTCAAGTTAACACCTGGC-3' (SEQ ID No. 43)
20		<i>Aat</i> II
	Y259N	5'-GACGAAGACGTCGTTAACTTGATGGAC-3' (SEQ ID No. 44) 5'-GTCCATCAAGTTAACGACGCTTCGTC-3' (SEQ ID No. 45)
25		<i>Asp</i> I
	E267D	5'-GTCCATTTCGACACTGTCGCTAGAACTT C-3' (SEQ ID No. 46) 5'-GAAGTTCTAGCGACAGTGTGCAATGGAC-3' (SEQ ID No. 47)
30	E277Q	5'-CTGACGCTACTCAGCTGTCTCCATTTC-3' (SEQ ID No. 48) 5'-GAATGGAGACAGCTGAGTAGCGTCAG-3' (SEQ ID No. 49)
35	A283D	5'-GTCTCCATTCTGTGATTTGTTCACTCAC-3' (SEQ ID No. 50) 5'-GTGAGTGAACAAATCACAGAATGGAGAC-3' (SEQ ID No. 51)
40		<i>Ksp</i> I
	H287A	5'-GCTTTGTTACCGCGGACGAATGGAG-3' (SEQ ID No. 52) 5'-CTCCATTTCGTCGCGGTGAACAAAGC-3' (SEQ ID No. 53)
45		<i>Bam</i> HI
	R291I	5'-CACGACGAATGGATCCAATACGACTAC-3' (SEQ ID No. 54) 5'-GTAGTCGTATTGGATCCATTCGTCGTG-3' (SEQ ID No. 55)
50		<i>Bsi</i> WI
	Q292A	5'-GACGAATGGAGAGCGTACGACTACTTG-3' (SEQ ID No. 56) 5'-CAAGTAGTCGTACGCTCTCCATTCGTC-3' (SEQ ID No. 57)
55		<i>Hpa</i> I
	A320V	5'-GGTGTGTTGTTTCGTTAACGAATTGATTGC-3' (SEQ ID No. 58) 5'-GCAATCAATTTCGTTAACGAAACCAACACC-3' (SEQ ID No. 59)

(Bgl II)

R329H 5'-GCTAGATTGACTCACTCTCCAGTTCAAG-3' (SEQ ID No. 60)  
 5'-CTTGAAGTGGAGAGTGAGTCAATCTAGC-3' (SEQ ID No. 61)

*Eco* RV

S364T 5'-CTCACGACAACACTATGATATCTATTTTCTTC-3' (SEQ ID No. 62)  
 5'-GAAGAAAATAGATATCATAGTGTGTCGTGAG-3' (SEQ ID No. 63)

*Nco* I

I366V 5'-CGACAACTCCATGGTTTCTATTTTCTTCGC-3' (SEQ ID No. 64)  
 5'-GCGAAGAAAATAGAAACCATGGAGTTGTCG-3' (SEQ ID No. 65)

*Kpn* I

A379K 5'-GTACAACGGTACCAAGCCATTGTCTAC-3' (SEQ ID No. 66)  
 5'-GTAGACAATGGCTTGGTACCGTTGTAC-3' (SEQ ID No. 67)

S396A 5'-CTGACGGTTACGCTGCTTCTTGGAC-3' (SEQ ID No. 68)  
 5'-GTCCAAGAAGCAGCGTAACCGTCAG-3' (SEQ ID No. 69)

G404A 5'-CTGTTCCATTGCTGCTAGAGCTTAC-3' (SEQ ID No. 70)  
 5'-GTAAGCTCTAGCAGCGAATGGAACAG-3' (SEQ ID No. 71)

Q415E 5'-GATGCAATGTGAAGCTGAAAAGGAACC-3' (SEQ ID No. 72)  
 5'-GGTTCCTTTTCAGCTTCACATTGCATC-3' (SEQ ID No. 73)

*Sal* I

A437G 5'-CACGGTTGTGGTGTGCGACAAGTTGGG-3' (SEQ ID No. 74)  
 5'-CCCAACTTGTGCGACACCACAACCGTG-3' (SEQ ID No. 75)

*Mun* I

A463E 5'-GATCTGGTGGCAATTGGGAGGAATGTTTCG-3' (SEQ ID No. 76)  
 5'-CGAAACATTCTCCCAATTGCCACCAGATC-3' (SEQ ID No. 77)

and accordingly for other mutations.

[0083] The temperature optimum of the purified phytases, expressed in *Saccharomyces cerevisiae* (Example 14), was determined as outlined in Example 14. Table 5 shows the effect on the stability of consensus phytase -1 for each mutation introduced.

**Table 5: Stability effect of the individual amino acid replacements in consensus phytase-1**

(+ or - means a positive, respectively, negative effect on the protein stability up to 1 °C, ++ and -- means a positive, respectively, negative effect on the protein stability between 1 and

3 °C; the number 10 or 11 corresponds to the consensus phytase sequence that suggested  
the amino acid replacement.)

5

10

15

20

25

30

35

40

45

50

55

stabilizing		neutral		destabilizing	
mutation	effect	mutation	effect	mutation	effect
E58A (10)	+	D69A	±	Y54F (10)	-
D69K (11)	+	D70G (10)	±	V73I	-
D197N (10)	+	N134Q (10)	±	A94K (10)	-
T214L (10)	++	G186H	±	A101R (11)	-
E222T (11)	++	S187A (10)	±	K153N (11)	-
E267D (10)	+	T214V	±	I158V (10)	--
R291I*	+	T251N (10)	±	G203A	--
R329H (10)	+	Y259N (10)	±	G205S	-
S364T (10)	++	A283D (10)	±	A217V	-
A379K (11)	+	A320V (10)	±	V227A (11)	--
G404A (10)	++	K445T	±	L234V (10)	-
		A463E (10)	±	A238P (10)	--
				E277Q (10)	-
				H287A (11)	-
				Q292A (10)	-
				I366V (10)	-
				S396A (10)	--
				Q415E (11)	-
				A437G (10)	--
				E451R	--

\*: This amino acid replacement was found in another round of mutations.

**[0084]** We combined eight positive mutations (E58A, D197N, E267D, R291I, R329H, S364T, A379K, G404A) in consensus phytase -1 using the primers and the technique mentioned above in this example. Furthermore, the mutations Q50T and K91A were introduced which mainly influence the catalytical characteristics of the phytase (see EP 897

985 as well as Example 14). The DNA and amino acid sequence of the resulting phytase gene (consensus phytase-1-thermo[8]-Q50T-K91A) is shown in Figure 8. In this way, the temperature optimum and the melting point of the consensus phytase was increased by 7 °C (Figure 16, 17, 18).

[0085] Using the results of Table 5, we further improved the thermostability of consensus phytase 10 by the following back mutations K94A, V158I, and A396S that revealed a strong negative influence on the stability of consensus phytase -1. The resulting protein is consensus phytase-10-thermo [3]. Furthermore, we introduced the mutations Q50T and K91A which mainly influence the catalytical characteristics of consensus phytase (see EP 897 485 as well as Example 14 and Figures 15 and 16). The resulting DNA and amino acid sequence is shown in Figure 9. The optimized phytase showed a 4 °C higher temperature optimum and melting point than consensus phytase -10 (Figures 13 and 14). Furthermore, the phytase has also a strongly increased specific activity with phytate as substrate of 250 U/mg at pH 5.5 (Figure 15).

#### Example 9

Stabilization of the phytase of *A. fumigatus* ATCC 13073 by replacement of amino acid residues with the corresponding consensus phytase-1 and consensus phytase-10 residues

[0086] At six typical positions where the *A. fumigatus* 13073 phytase is the only or nearly the only phytase in the alignment of Figure 2 that does not contain the corresponding consensus phytase amino acid residue, the non-consensus amino acid residue was replaced by the consensus one. In a first round, the following amino acids were substituted in *A. fumigatus* 13073 phytase, containing the Q51T substitution and the signal sequence of *A. terreus* cbs.116.46 phytase (see Figure 10):

F55(28)Y, V100(73)I, F114(87)Y, A243(220)L, S265(242)P, N294(282)D.

[0087] The numbers in parentheses refer to the numbering of Figure 2.

[0088] In a second round, four of the seven stabilizing amino acid exchanges (E59A, R329H, S364T, G404A) found in the consensus phytase-10 sequence and, tested as single mutations in consensus phytase-1 (Table 5), were additionally introduced into the *A. fumigatus*  $\alpha$ -mutant. Furthermore, the amino acid replacement S154N, shown to reduce the protease susceptibility of the phytase, was introduced.

[0089] The mutations were introduced as described in example 8 (see Table 6) and expressed as described in example 11 to 13. The resulting *A. fumigatus* 13073 phytase variants were called  $\alpha$ -mutant and  $\alpha$ -mutant-E59A-S154N-R329H-S364T-G404A.

[0090] The temperature optimum (60 °C, Figure 21) and the melting point (67.0 °C, Figure 20) of the *A. fumigatus* 13073 phytase  $\alpha$ -mutant were increased by 5 - 7°C in comparison to the values of the wild-type (temperature optimum: 55 °C,  $T_m$ : 60 °C). The five additional amino acid replacements further increased the temperature optimum by 3 °C (Figure 21).

**Table 6: Mutagenesis primers for stabilization of *A. fumigatus* phytase ATCC 13073**

5	Mutation	Primer
	F55Y	5'-CACGTACTCGCCATACTTTTCGCTCGAG-3' (SEQ ID No. 78) 5'-CTCGAGCGAAAAGTATGGCGAGTACGTG-3' (SEQ ID No. 79)
10		( <i>Xho</i> I)
	E58A	5'-CCATACTTTTCGCTCGCGGACGAGCTGTCCGTG-3' (SEQ ID NO. 80) 5'-CACGGACAGCTCGTCCGCGAGCGAAAAGTAGG-3' (SEQ ID NO. 81)
15		
	V100I	5'-GTATAAGAAGCTTATTACGGCGATCCAGGCC-3' (SEQ ID No. 82) 5'-GGCCTGGATCGCCGTAATAAGCTTCTTATAC-3' (SEQ ID No. 83)
20		
	F114Y	5'-CTTCAAGGGCAAGTACGCCTTTTGAAGACG-3' (SEQ ID No. 84) 5'-CGTCTTCAAAAAGGCGTACTTGCCCTTGAAG-3' (SEQ ID No. 85)
25		
	A243L	5'-CATCCGAGCTCGCCTCGAGAAGCATCTTC-3' (SEQ ID No. 86) 5'-GAAGATGCTTCTCGAGGCGAGCTCGGATG-3' (SEQ ID No. 87)
30		
	S265P	5'-CTAATGGA TGTGTCCGTTTGATACGGTAG-3' (SEQ ID No. 88) 5'-CTACCGTATCAAACGGACACATGTCCATTAG-3' (SEQ ID No. 89)
35		
40		
45		
50		
55		



N294D 5'-GTGGAAGAAGTACGACTACCTTCAGTC-3' (SEQ ID No. 90)  
5'-GACTGAAGGTAGTCGTA~~CTT~~CCTCCAC-3' (SEQ ID No. 91)

(*Mlu* I)

R329H 5'-GCCCCGGTTGACGCATTGCGCCAGTGCAGG-3' (SEQ ID No. 92)  
5'-CCTGCACTGGCGAATGCGTCAACCGGGC-3' (SEQ ID No. 93)

*Nco* I

S364T 5'-CACACGACAACACCATGGTTTCCATCTTC-3' (SEQ ID No. 94)  
5'-GAAGATGGAAACCATGGTGTGTCGTGTG-3' (SEQ ID No. 95)

(*Bss* HI)

G404A 5'-GTGGTGCCTTTCGCCGCGGAGCCTACTTC-3' (SEQ ID No. 96)  
5'-GAAGTAGGCTCGCGCGGCGAAAGGCACCAC-3' (SEQ ID No. 97)

#### Example 10

Introduction of the active site amino acid residues of the *A. niger* NRRL 3135 phytase into the consensus phytase-1

**[0091]** We used the crystal structure of the *Aspergillus niger* NRRL 3135 phytase to define all active site amino acid residues (see Reference Example and EP 897 010). Using the alignment of Figure 2, we replaced the following active site residues and additionally the non-identical adjacent ones of the consensus phytase -1 by those of the *A. niger* phytase:

S89D, S92G, A94K, D164S, P201S, G203A, G205S, H212P, G224A, D226T, E255T, D256E, V258T, P265S, Q292H, G300K, Y305H, A314T, S364G, M365I, A397S, S398A, G404A, and A405S

**[0092]** The new protein sequence consensus phytase -7 was backtranslated into a DNA sequence (Figure 11) as described in Example 6. The corresponding gene (*fcp7*) was generated as described in Example 6 using the following oligonucleotide mixes:

Mix 1.7: CP-1, CP-2, CP-3, CP-4.7, CP-5.7, CP-6, CP-7, CP-8.7, CP-9, CP-10.7

Mix 2.7: CP-9, CP-10.7, CP-11.7, CP-12.7, CP-13.7, CP-14.7, CP-15.7, CP-16, CP-17.7, CP-18.7, CP-19.7, CP-20, CP-21, CP-22.

**[0093]** The DNA sequences of the oligonucleotides are indicated in Figure 11. The newly synthesized oligonucleotides are additionally marked by number 7. After assembling of the oligonucleotides using the same PCR primers as mentioned in Example 6, the gene was cloned into an expression vector as described in Examples 11 - 13.

**[0094]** The pH-profile of consensus phytase-7, purified after expression in *Hansenula polymorpha*, was very similar to that of *A. niger* NRRL 3135 phytase (see Figure 19).

#### Example 11

Expression of the consensus phytase genes in *Hansenula polymorpha*

**[0095]** The phytase expression vectors, used to transform *H. polymorpha* RB11 (Gellissen *et al.*, 1994), were constructed by inserting the *Eco* RI fragment of pBsk<sup>+</sup>*fcp* or variants thereof into the multiple cloning site of the *H.*

*polymorpha* expression vector pFPMT121, which is based on an *ura3* selection marker from *S. cerevisiae*, a formate dehydrogenase (*FMD*) promoter element and a methanol oxidase (*MO*) terminator element from *H. polymorpha*. The 5' end of the *fcp* gene is fused to the *FMD* promoter, the 3' end to the *MOX* terminator (Gellissen *et al.*, 1996; EP 0299 108 B). The resulting expression vectors were designated pFPMT*fcp*, pFPMT*fcp10*, pFPMT*fcp7*.

**[0096]** The constructed plasmids were propagated in *E. coli*. Plasmid DNA was purified using standard state of the art procedures. The expression plasmids were transformed into the *H. polymorpha* strain RP11 deficient in orotidine-5'-phosphate decarboxylase (*ura3*) using the procedure for preparation of competent cells and for transformation of yeast as described in Gellissen *et al.* (1996). Each transformation mixture was plated on YNB (0.14% w/v Difco YNB and 0.5% ammonium sulfate) containing 2% glucose and 1.8% agar and incubated at 37 °C. After 4 to 5 days individual transformant colonies were picked and grown in the liquid medium described above for 2 days at 37 °C. Subsequently, an aliquot of this culture was used to inoculate fresh vials with YNB-medium containing 2% glucose. After seven further passages in selective medium, the expression vector is integrated into the yeast genome in multimeric form. Subsequently, mitotically stable transformants were obtained by two additional cultivation steps in 3 ml non-selective liquid medium (YPD, 2% glucose, 10 g yeast extract, and 20 g peptone). In order to obtain genetically homogeneous recombinant strains an aliquot from the last stabilization culture was plated on a selective plate. Single colonies were isolated for analysis of phytase expression in YNB containing 2% glycerol instead of glucose to derepress the *fmd* promoter. Purification of the consensus phytases was done as described in Example 12.

#### Example 12

##### Expression of the consensus phytase genes in *Saccharomyces cerevisiae* and purification of the phytases from culture supernatant

**[0097]** The consensus phytase genes were isolated from the corresponding Bluescript-plasmid (pBsk<sup>-</sup>*fcp*, pBsk<sup>-</sup>*fcp10*, pBsk<sup>-</sup>*fcp7*) and ligated into the *Eco* RI sites of the expression cassette of the *Saccharomyces cerevisiae* expression vector pYES2 (Invitrogen, San Diego, CA, USA) or subcloned between the shortened GAPFL (glyceraldehyde-3-phosphate dehydrogenase) promoter and the *pho5* terminator as described by Janes *et al.* (1990). The correct orientation of the gene was checked by PCR. Transformation of *S. cerevisiae* strains. e. g. INVSc1 (Invitrogen, San Diego, CA, USA) was done according to Hinnen *et al.* (1978). Single colonies harboring the phytase gene under the control of the GAPFL promoter were picked and cultivated in 5 ml selection medium (SD-uracil, Sherman *et al.*, 1986) at 30°C under vigorous shaking (250 rpm) for one day. The preculture was then added to 500 ml YPD medium (Sherman *et al.*, 1986) and grown under the same conditions. Induction of the *gal1* promoter was done according to the manufacturer's instructions. After four days of incubation cell broth was centrifuged (7000 rpm, GS3 rotor, 15 min, 5°C) to remove the cells and the supernatant was concentrated by way of ultrafiltration in Amicon 8400 cells (PM30 membranes) and ultra-free-15 centrifugal filter devices (Biomax-30K, Millipore, Bedford, MA, USA). The concentrate (10 ml) was desalted on a 40 ml Sephadex G25 Superfine column (Pharmacia Biotech, Freiburg, Germany), with 10 mM sodium acetate, pH 5.0, serving as elution buffer. The desalted sample was brought to 2 M (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> and directly loaded onto a 1 ml Butyl Sepharose 4 Fast Flow hydrophobic interaction chromatography column (Pharmacia Biotech, Freiburg, Germany) which was eluted with a linear gradient from 2 M to 0 M (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> in 10 mM sodium acetate, pH 5.0. Phytase was eluted in the break-through, concentrated and loaded on a 120 ml Sephacryl S-300 gel permeation chromatography column (Pharmacia Biotech, Freiburg, Germany). Consensus phytase -1 and consensus phytase -7 eluted as a homogeneous symmetrical peak and was shown by SDS-PAGE to be approx. 95% pure.

#### Example 13

##### Expression of the consensus phytase genes in *Aspergillus niger*

**[0098]** The Bluescript-plasmids pBsk<sup>-</sup>*fcp*, pBsk<sup>-</sup>*fcp10*, and pBsk<sup>-</sup>*fcp7* were used as template for the introduction of a *Bsp* HI-site upstream of the start codon of the genes and an *Eco* RV-site downstream of the stop codon. The Expand™ High Fidelity PCR Kit (Boehringer Mannheim, Mannheim, Germany) was used with the following primers:

**Primer Asp-1:***Bsp* HI

5'-TATATCATGAGCGTGTTCGTCGTGCTACTGTTC-3' (SEQ ID No. 98)

**Primer Asp-2 used for cloning of *fcp* and *fcp7*:***Eco* RV

3'-ACCCGACTTACAAAGCGAATTCTATAGATATAT-5' (SEQ ID No. 99)

**Primer Asp-3 used for cloning of *fcp10*:***Eco* RV

3'-ACCCTTCTTACAAAGCGAATTCTATAGATATAT-5' (SEQ ID No. 100)

[0099] The reaction was performed as described by the supplier. The PCR-amplified *fcp*-genes had a new *Bsp* HI site at the start codon, introduced by primer Asp-1, which resulted in a replacement of the second amino acid residue glycine by serine. Subsequently, the DNA-fragment was digested with *Bsp* HI and *Eco* RV and ligated into the *Nco* I site downstream of the glucoamylase promoter of *Aspergillus niger* (*glaA*) and the *Eco* RV site upstream of the *Aspergillus nidulans* tryptophan C terminator (*trpC*) (Mullaney *et al.*, 1985). After this cloning step, the genes were sequenced to detect possible failures introduced by PCR. The resulting expression plasmids which basically correspond to the pGLAC vector as described in Example 9 of EP 684 313 contained the orotidine-5'-phosphate decarboxylase gene (*pyr4*) of *Neurospora crassa* as a selection marker. Transformation of *Aspergillus niger* and expression of the consensus phytase genes was done as described in EP 684 313. The consensus phytases were purified as described in Example 12.

Example 14Determination of phytase activity and of temperature optimum

[0100] Phytase activity was determined basically as described by Mitchell *et al* (1997). The activity was measured in an assay mixture containing 0.5% phytic acid ( $\approx 5$  mM) in 200 mM sodium acetate, pH 5.0. After 15 min of incubation at 37 °C, the reaction was stopped by addition of an equal volume of 15% trichloroacetic acid. The liberated phosphate was quantified by mixing 100  $\mu$ l of the assay mixture with 900  $\mu$ l H<sub>2</sub>O and 1 ml of 0.6 M H<sub>2</sub>SO<sub>4</sub>, 2% ascorbic acid and 0.5% ammonium molybdate. Standard solutions of potassium phosphate were used as reference. One unit of enzyme activity was defined as the amount of enzyme that releases 1  $\mu$ mol phosphate per minute at 37 °C. The protein concentration was determined using the enzyme extinction coefficient at 280 nm calculated according to Pace *et al* (1995): consensus phytase -1.101; consensus phytase -7, 1.068; consensus phytase -1 10, 1.039.

[0101] In case of pH-optimum curves, purified enzymes were diluted in 10 mM sodium acetate, pH 5.0. Incubations were started by mixing aliquots of the diluted protein with an equal volume of 1% phytic acid ( $\approx 10$  mM) in a series of different buffers: 0.4 M glycine/HCl, pH 2.5; 0.4 M acetate/NaOH, pH 3.0, 3.5, 4.0, 4.5, 5.0, 5.5; 0.4 M imidazole/HCl, pH 6.0, 6.5; 0.4 M Tris/HCl pH 7.0, 7.5, 8.0, 8.5, 9.0. Control experiments showed that pH was only slightly affected by the mixing step. Incubations were performed for 15 min at 37 °C as described above.

[0102] For determinations of the substrate specificities of the phytases, phytic acid in the assay mixture was replaced by 5 mM concentrations of the respective phosphate compounds. The activity tests were performed as described above.

[0103] For determination of the temperature optimum, enzyme (100  $\mu$ l) and substrate solution (100  $\mu$ l) were pre-incubated for 5 min at the given temperature. The reaction was started by addition of the substrate solution to the enzyme. After 15 min incubation, the reaction was stopped with trichloroacetic acid and the amount of phosphate released was determined.

[0104] The pH-optimum of the original consensus phytase was around pH 6.0-6.5 (80 U/mg). By introduction of the

Q50T mutation, the pH-optimum shifted to pH 6.0 (130 U/mg). After introduction of K91A, the pH optimum shifted one pH-unit into the acidic pH-range showing a higher specific activity between pH 2.5 and pH 6.0. That was shown for the stabilized mutants and for consensus phytase-10, too (Figure 15 and 16).

**[0105]** Consensus phytase-7, which was constructed to transfer the catalytic characteristics of the *A. niger* NRRL 3135 phytase into consensus phytase-1, had a pH-profile very similar to that of *A. niger* NRRL 3135 phytase (see Figure 19). The substrate specificity of consensus phytase-7 also resembled more to that of *A. niger* NRRL 3135 phytase than to that of consensus phytase-1.

**[0106]** The temperature optimum of consensus phytase-1 (71 °C) was 16-26 °C higher than the temperature optimum of the wild-type phytases (45-55 °C, Table 7) which were used to calculate the consensus sequence. The improved consensus phytase-10 showed a further increase of its temperature optimum to 80 °C (Figure 12). The temperature optimum of the consensus phytase-1-thermo[8] phytase was found in the same range (78 °C) when using the supernatant of an overproducing *S. cerevisiae* strain. The highest temperature optimum reached of 82 °C was determined for consensus phytase-10-thermo[3]-Q50T-K91A.

Table 7

Temperature optimum and $T_m$ -value of consensus phytase and of the phytases from <i>A. fumigatus</i> , <i>A. niger</i> , <i>E. nidulans</i> and <i>M. thermophila</i> . The determination of the temperature optimum was performed as described in Example 14. The $T_m$ -values were determined by differential scanning calorimetry as described in Example 15.		
phytase	temperature optimum [°C]	$T_m$ [°C]
Consensus phytase-10-thermo[3]-Q50T-K91A	82	89.3
Consensus phytase-10-thermo[3]-Q50T	82	88.6
Consensus phytase-10	80	85.4
Consensus phytase-1-thermo[8]-Q50T	78	84.7
Consensus phytase-1-thermo[8]-Q50T-K91A	78	85.7
Consensus phytase-1	71	78.1
<i>A. niger</i> NRRL3135	55	63.3
<i>A. fumigatus</i> 13073	55	62.5
<i>A. fumigatus</i> 13073 $\alpha$ -mutant	60	67.0
<i>A. fumigatus</i> 13073 $\alpha$ -mutant (optimized)	63	-
<i>A. terreus</i> 9A-1	49	57.5
<i>A. terreus</i> cbs.116.46	45	58.5
<i>E. nidulans</i>	45	55.7
<i>M. thermophila</i>	55	n. d.
<i>T. thermophilus</i>	45	n. d.

#### Example 15

##### Determination of the melting point by differential scanning calorimetry (DSC)

**[0107]** In order to determine the unfolding temperature of the phytases, differential scanning calorimetry was applied as previously published by Lehmann et al (2000). Solutions of 50-60 mg/ml homogeneous phytase were used for the tests. A constant heating rate of 10 °C/min was applied up to 90-95 °C.

**[0108]** The determined melting points reflect the results obtained for the temperature optima (Table 7). The most stable consensus phytase designed is consensus phytase-10-thermo[3]-Q50T-K91A showing a melting temperature

under the chosen conditions of 89.3 °C. This is 26 to 33.6 °C higher than the melting points of the wild-type phytases used.

#### Example 16

##### Transfer of basidiomycete phytase active site into consensus phytase-10-thermo[3]-Q50T-K91A

**[0109]** As described previously (Example 8), mutations derived from the basidiomycete phytase active site were introduced into the consensus phytase -10. The following five constructs a) to e) were prepared:

a) This construct is called consensus phytase -12, and it comprises a selected number of active site residues of the basidio consensus sequence. Its amino acid sequence (consphy12) is shown in Fig. 22 (the first 26 amino acids forms the signal peptide, amended positions are underlined);

b) a cluster of mutations (Cluster II) was transferred to the consensus phytase 10 sequence, viz.: S80Q, Y86F, S90G, K91A, S92A, K93T, A94R, Y95I;

c) analogously, another cluster of mutations (Cluster III) was transferred, viz.: T129V, E133A, Q143N, M136S, V137S, N138Q, S139A;

d) analogously, a further cluster of mutations (Cluster IV) was transferred, viz.: A168D, E171T, K172N, F173W;

e) and finally, a further cluster of mutations (Cluster V) was transferred, viz.: Q297G, S298D, G300D, Y305T.

**[0110]** These constructs were expressed as described in Examples 11 - 13.

#### References:

##### **[0111]**

Akanuma, S., Yamagishi, A., Tanaka, N. & Oshima, T. (1998). Serial increase in the thermal stability of 3-isopropyl-malate dehydrogenase from *Bacillus subtilis* by experimental evolution. *Prot. Sci.* 7, 698-705.

Arase, A., Yomo, T., Urabe, I., Hata, Y., Katsube, Y. & Okada, H. (1993). Stabilization of xylanase by random mutagenesis. *FEBS Lett.* 316, 123-127.

Berka, R. M., Rey M. W., Brown, K. M., Byun, T. & Klotz, A. V. (1998). Molecular characterization and expression of a phytase gene from the thermophilic fungus *Thermomyces lanuginosus*. *Appl. Environ. Microbiol.* 64, 4423-4427.

Blaber, M., Lindstrom, J. D., Gassner, N., Xu, J., Heinz, D. W. & Matthews, B. W. (1993). Energetic cost and structural consequences of burying a hydroxyl group within the core of a protein determined from Ala/Ser and Val/Thr substitutions in T4 lysozyme. *Biochemistry* 32, 11363-11373.

Cosgrove, D.J. (1980) Inositol phosphates - their chemistry, biochemistry and physiology: studies in organic chemistry, chapter 4. Elsevier Scientific Publishing Company, Amsterdam, Oxford, New York.

Devereux, J., Haeberli, P. & Smithies, O. (1984) A comprehensive set of sequence analysis programs for the VAX. *Nucleic Acids Res.* 12, 387-395.

Gellissen, G., Hollenberg, C. P., Janowicz, Z. A. (1994) Gene expression in methylotrophic yeasts. In: Smith, A. (ed.) Gene expression in recombinant microorganisms. Dekker, New York, 395-439.

Gellissen, G., Piontek, M., Dahlems, U., Jenzelewski, V., Gavagan, J. E., DiCosimo, R., Anton, D. I. & Janowicz, Z. A. (1996) Recombinant *Hansenula polymorpha* as a biocatalyst: coexpression of the spinach glycolate oxidase (GO) and the *S. cerevisiae* catalase T (CTT1) gene. *Appl. Microbiol. Biotechnol.* 46, 46-54.

Gerber, P. and Müller, K. (1995) Moloc molecular modeling software. *J. Comput. Aided Mol. Des.* 9, 251-268

Hinnen, A., Hicks, J. B. & Fink, G. R. (1978) Transformation of yeast. *Proc. Natl. Acad. Sci. USA* 75, 1929-1933.

Imanaka, T., Shibasaki, M. & Takagi, M. (1986). A new way of enhancing the thermostability of proteases. *Nature* 324, 695-697.

Janes, M., Meyhack, B., Zimmermann, W. & Hinnen, A. (1990) The influence of GAP promoter variants on hirudine production, average plasmid copy number and cell growth in *Saccharomyces cerevisiae*. *Curr. Genet.* 18, 97-103.

Karpusas, M., Baase, W. A., Matsumura, M. & Matthews, B. W. (1989). Hydrophobic packing in T4 lysozyme probed by cavity-filling mutants. *Proc. Natl. Acad. Sci.(USA)* 86, 8237-8241.

Lehmann, L., Kostrewa, D., Wyss, M., Brugger, R., D'Arcy, A., Pasamontes, L., van Loon, A. (2000), From DNA

sequence to improved functionality: using protein sequence comparisons to rapidly design a thermostable consensus phytase, *Protein Engineering* 13, 49-57.

Margarit, I., Campagnoli, S., Frigerio, F., Grandi, G., Fillipis, V. D. & Fontana, A. (1992). Cumulative stabilizing effects of glycine to alanine substitutions in *Bacillus subtilis* neutral protease. *Prot. Eng.* 5, 543-550.

Matthews, B. W. (1987a). Genetic and structural analysis of the protein stability problem. *Biochemistry* 26, 6885-6888.

Matthews, B. W. (1993). Structural and genetic analysis of protein stability. *Annu. Rev. Biochem.* 62, 139-160.

Matthews, B. W., Nicholson, H. & Becktel, W. (1987). Enhanced protein thermostability from site-directed mutations that decrease the entropy of unfolding. *Proc. Natl. Acad. Sci. (USA)* 84, 6663-6667.

Mitchell, D. B., Vogel, K., Weimann, B. J., Pasamontes, L. & van Loon, A. P. G. M. (1997) The phytase subfamily of histidine acid phosphatases: isolation of genes for two novel phytases from the fungi *Aspergillus terreus* and *Myceliophthora thermophila*, *Microbiology* 143, 245-252.

Mullaney, E. J., Hamer, J. E., Roberti, K. A., Yelton, M. M. & Timberlake, W. E. (1985) Primary structure of the *trpC* gene from *Aspergillus nidulans*. *Mol. Gen. Genet.* 199, 37-46.

Munoz, V. & Serrano, L. (1995). Helix design, prediction and stability. *Curr. Opin. Biotechnol.* 6, 382-386.

Pace, N. C., Vajdos, F., Fee, L., Grimsley, G. & Gray, T. (1995). How to measure and predict the molar absorption coefficient of a protein. *Prot. Sci.* 4, 2411-2423.

Pantoliano, M. W., Landner, R. C., Brian, P. N., Rollence, M. L., Wood, J. F. & Poulos, T. L. (1987). Protein engineering of subtilisin BPN': enhanced stabilization through the introduction of two cysteines to form a disulfide bond. *Biochemistry* 26, 2077-2082.

Pasamontes, L., Haiker, M., Henriquez-Huecas, M., Mitchell, D. B. & van Loon, A. P. G. M. (1997a). Cloning of the phytases from *Emmericella nidulans* and the thermophilic fungus *Talaromyces thermophilus*. *Biochim. Biophys. Acta* 1353, 217-223.

Pasamontes, L., Haiker, M., Wyss, M., Tessier, M. & van Loon, A. P. G. M. (1997) Cloning, purification and characterization of a heat stable phytase from the fungus *Aspergillus fumigatus*, *Appl. Environ. Microbiol.* 63, 1696-1700.

Piddington, C. S., Houston, C. S., Paloheimo, M., Cantrell, M., Miettinen-Oinonen, A., Nevalainen, H., & Rambosek, J. (1993) The cloning and sequencing of the genes encoding phytase (*phy*) and pH 2.5-optimum acid phosphatase (*aph*) from *Aspergillus niger* var. *awamori*. *Gene* 133, 55-62.

Purvis, I. J., Bettany, A. J. E., Santiago, T. C., Coggins, J. R., Duncan, K., Eason, R. & Brown, A. J. P. (1987). The efficiency of folding of some proteins is increased by controlled rates of translation *in vivo*. *J. Mol. Biol.* 193, 413-417.

Risse, B., Stempfer, G., Rudolph, R., Schumacher, G. & Jaenicke, R. (1992). Characterization of the stability effect of point mutations of pyruvate oxidase from *Lactobacillus plantarum*: protection of the native state by modulating coenzyme binding and subunit interaction. *Prot. Sci.* 1, 1710-1718.

Sambrook, J., Fritsch, E. F. & Maniatis, T. (1989) *Molecular Cloning: A Laboratory Manual*, 2nd Ed., Cold Spring Harbor Laboratory, Cold Spring Harbor, NY.

Sauer, R., Hehir, K., Stearman, R., Weiss, M., Jeitler-Nilsson, A., Suchanek, E. & Pabo, C. (1986). An engineered intersubunit disulfide enhances the stability and DNA binding of the N-terminal domain of 1-repressor. *Biochemistry* 25, 5992-5999.

Serrano, L., Day, A. G. & Fersht, A. R. (1993). Step-wise mutation of barnase to binase. A procedure for engineering increased stability of proteins and an experimental analysis of the evolution of protein stability. *J. Mol. Biol.* 233, 305-312.

Sheman, J. P., Finck, G. R. & Hicks, J. B. (1986) Laboratory course manual for methods in yeast genetics. Cold Spring Harbor University.

Steipe, B., Schiller, B., Plueckthun, A. & Steinbach, S. (1994). Sequence statistics reliably predict stabilizing mutations in a protein domain. *J. Mol. Biol.* 240, 188-192.

van den Burg, B., Vriend, G., Veltman, O. R., Venema & G., Eijssink, V. G. H. (1998). Engineering an enzyme to resist boiling. *Proc. Natl. Acad. Sci. (USA)* 95, 2056-2060.

Van Etten, R.L. (1982) Human prostatic acid phosphatase: a histidine phosphatase. *Ann. NY Acad. Sci.* 390, 27-50.

van Hartingsveldt, W., van Zeijl, C. M. F., Hartevelde, G. M., Gouka, R. J., Suykerbuyk, M. E. G., Luiten, R. G. M., van Paridon, P. A., Selden, G. C. M., Veenstra, A. E., van Gorcom, R. F. M., & van den Hondel, C. A. M. J. J. (1993) Cloning, characterization and overexpression of the phytase-encoding gene (*phyA*) of *Aspergillus niger*. *Gene* 127, 87-94.

Annex to the application documents - subsequently filed sequence listing

[0112]

```

5                               SEQUENCE LISTING.txt
    SEQUENCE LISTING

10    <110> F. Hoffmann-La Roche AG
    <120> Continuous fermentation process
    <130> Case 20486 EP2

15    <140> EP 00121663.9
    <141> 2000-10-04

    <150> EP 99120289.6
    <151> 1999-10-11

20    <150> EP 00119676.5
    <151> 2000-09-08

    <160> 113

25    <170> PADAT Sequenzmodul, Version 1.0

    <210> 1
    <211> 26
    <212> DNA
30    <213> Artificial Sequence

    <220>
    <223> Primer

35    <400> 1

    tatatgaatt catgggcgtg ttcgtc                                     26

40    <210> 2
    <211> 22
    <212> DNA
    <213> Artificial Sequence

    <220>
45    <223> Primer

    <400> 2

50    tgaaaagttc attgaaggtt tc                                       22

    <210> 3
    <211> 22
    <212> DNA
55    <213> Artificial Sequence

```

55



33

34

35

55

37

SEQUENCE LISTING.txt

5 <220>  
<223> Primer  
  
<400> 27

10 gaaccagaag cgcgcacgaa tggaacaatc 30

<210> 28  
<211> 31  
<212> DNA  
15 <213> Artificial Sequence

<220>  
<223> Primer  
  
20 <400> 28

ctccagttat taacgtgatc attccagaag g 31

<210> 29  
<211> 31  
<212> DNA  
25 <213> Artificial Sequence

<220>  
<223> Primer  
30 <400> 29

ccttctggaa tgatcacgtt aataactgga g 31

<210> 30  
<211> 30  
<212> DNA  
40 <213> Artificial Sequence

<220>  
<223> Primer  
  
45 <400> 30

ggctgacca ggggccaac cacaccaagc 30

<210> 31  
<211> 30  
<212> DNA  
50 <213> Artificial Sequence

55

39

## SEQUENCE LISTING.txt

5       <220>  
       <223> Primer  
       <400> 35  
  
 10       caacgtcgtc acctagggtg gagtcttcga aagc                               34  
  
       <210> 36  
       <211> 26  
 15       <212> DNA  
       <213> Artificial Sequence  
  
       <220>  
       <223> Primer  
 20       <400> 36  
  
       ggtgacgacg ctgaagctaa cttcac                               26  
  
 25       <210> 37  
       <211> 26  
       <212> DNA  
       <213> Artificial Sequence  
  
 30       <220>  
       <223> Primer  
       <400> 37  
  
 35       gtgaagttag cttcagcgtc gtcacc                               26  
  
       <210> 38  
       <211> 27  
 40       <212> DNA  
       <213> Artificial Sequence  
  
       <220>  
       <223> Primer  
 45       <400> 38  
  
       ctaacttcac cgcggtgttc gtcacc                               27  
  
 50       <210> 39  
       <211> 27  
       <212> DNA  
       <213> Artificial Sequence  
  
 55



41



43



45

46

55

48



55

## SEQUENCE LISTING.txt

	<220>	
5	<223> Primer	
	<400> 75	
10	cccaacttgt cgacaccaca accgtg	26
	<210> 76	
	<211> 30	
	<212> DNA	
15	<213> Artificial Sequence	
	<220>	
	<223> Primer	
20	<400> 76	
	gatctggtgg caattgggag gaatgtttcg	30
	<210> 77	
25	<211> 30	
	<212> DNA	
	<213> Artificial Sequence	
	<220>	
30	<223> Primer	
	<400> 77	
	cgaaacattc ctcccaattg ccaccagatc	30
35		
	<210> 78	
	<211> 28	
	<212> DNA	
40	<213> Artificial Sequence	
	<220>	
	<223> Primer	
45	<400> 78	
	cacgtactcg ccatactttt cgctcgag	28
50		
	<210> 79	
	<211> 28	
	<212> DNA	
	<213> Artificial Sequence	

51

55

## SEQUENCE LISTING.txt

5 <220>  
<223> Primer  
  
<400> 87

10 gaagatgctt ctcgaggcga gctcggatg 29

<210> 88  
<211> 29  
<212> DNA  
15 <213> Artificial Sequence

<220>  
<223> Primer  
  
<400> 88

20 ctaatggatg tgtccgtttg atacggtag 29

<210> 89  
25 <211> 31  
<212> DNA  
<213> Artificial Sequence

<220>  
30 <223> Primer  
  
<400> 89

35 ctaccgtatc aaacggacac atgtccatta g 31

<210> 90  
<211> 27  
<212> DNA  
40 <213> Artificial Sequence

<220>  
<223> Primer  
  
<400> 90

45 gtggaagaag tacgactacc ttcagtc 27

<210> 91  
50 <211> 27  
<212> DNA  
<213> Artificial Sequence

55

## SEQUENCE LISTING.txt

5 <220>  
<223> Primer  
  
<400> 91  
  
10 gactgaagggt agtcgtactt cttccac 27  
  
<210> 92  
<211> 28  
<212> DNA  
15 <213> Artificial Sequence  
  
<220>  
<223> Primer  
  
20 <400> 92  
  
gcccggttga cgcattcgcc agtgcagg 28  
  
<210> 93  
25 <211> 28  
<212> DNA  
<213> Artificial Sequence  
  
<220>  
30 <223> Primer  
  
<400> 93  
  
cctgcactgg cgaatgcgtc aaccgggc 28  
35  
  
<210> 94  
<211> 29  
<212> DNA  
40 <213> Artificial Sequence  
  
<220>  
<223> Primer  
  
45 <400> 94  
  
cacacgacaa caccatggtt tccatcttc 29  
  
<210> 95  
50 <211> 29  
<212> DNA  
<213> Artificial Sequence  
  
55

## SEQUENCE LISTING.txt

5 <220>  
<223> Primer  
  
<400> 95  
  
10 gaagatggaa accatggtgt tgcgtgtg 29  
  
<210> 96  
<211> 30  
<212> DNA  
15 <213> Artificial Sequence  
  
<220>  
<223> Primer  
  
20 <400> 96  
  
gtggtgcctt tcgccgcgcg agcctacttc 30  
  
<210> 97  
25 <211> 30  
<212> DNA  
<213> Artificial Sequence  
  
<220>  
30 <223> Primer  
  
<400> 97  
  
35 gaagtaggct cgcgcggcga aaggcaccac 30  
  
<210> 98  
<211> 33  
<212> DNA  
40 <213> Artificial Sequence  
  
<220>  
<223> Primer  
  
<400> 98  
45  
  
tatatcatga gcgtgttcgt cgtgctactg ttc 33  
  
<210> 99  
50 <211> 33  
<212> DNA  
<213> Artificial Sequence  
  
55

## SEQUENCE LISTING.txt

5 <220>  
 <223> Primer  
  
 <400> 99  
  
 acccgactta caaagcgaat tctatagata tat 33  
  
 10 <210> 100  
 <211> 33  
 <212> DNA  
 <213> Artificial Sequence  
  
 15 <220>  
 <223> Primer  
  
 <400> 100  
  
 20 acccttctta caaagcgaat tctatagata tat 33  
  
 <210> 101  
 <211> 467  
 <212> PRT  
 <213> Artificial Sequence  
  
 25 <220>  
 <223> consensus phytase-1  
  
 <400> 101  
  
 30 Met Gly Val Phe Val Val Leu Leu Ser Ile Ala Thr Leu Phe Gly Ser  
       1                  5                  10  
 Thr Ser Gly Thr Ala Leu Gly Pro Arg Gly Asn Ser His Ser Cys Asp  
                   20                  25                  30  
 35 Thr Val Asp Gly Gly Tyr Gln Cys Phe Pro Glu Ile Ser His Leu Trp  
                   35                  40                  45  
 Gly Gln Tyr Ser Pro Tyr Phe Ser Leu Glu Asp Glu Ser Ala Ile Ser  
           50                  55                  60  
 Pro Asp Val Pro Asp Asp Cys Arg Val Thr Phe Val Gln Val Leu Ser  
       65                  70                  75                  80  
 40 Arg His Gly Ala Arg Tyr Pro Thr Ser Ser Lys Ser Lys Ala Tyr Ser  
                   85                  90                  95  
 Ala Leu Ile Glu Ala Ile Gln Lys Asn Ala Thr Ala Phe Lys Gly Lys  
           100                  105                  110  
 Tyr Ala Phe Leu Lys Thr Tyr Asn Tyr Thr Leu Gly Ala Asp Asp Leu  
           115                  120                  125  
 45 Thr Pro Phe Gly Glu Asn Gln Met Val Asn Ser Gly Ile Lys Phe Tyr  
       130                  135                  140  
 Arg Arg Tyr Lys Ala Leu Ala Arg Lys Ile Val Pro Phe Ile Arg Ala  
 145                  150                  155                  160  
 50 Ser Gly Ser Asp Arg Val Ile Ala Ser Ala Glu Lys Phe Ile Glu Gly  
                   165                  170                  175  
 Phe Gln Ser Ala Lys Leu Ala Asp Pro Gly Ser Gln Pro His Gln Ala  
           180                  185                  190

55



## SEQUENCE LISTING.txt

Ser Pro Val Ile Asp Val Ile Ile Pro Glu Gly Ser Gly Tyr Asn Asn  
 195 200 205  
 5 Thr Leu Asp His Gly Thr Cys Thr Ala Phe Glu Asp Ser Glu Leu Gly  
 210 215 220  
 Asp Asp Val Glu Ala Asn Phe Thr Ala Leu Phe Ala Pro Ala Ile Arg  
 225 230 235 240  
 Ala Arg Leu Glu Ala Asp Leu Pro Gly Val Thr Leu Thr Asp Glu Asp  
 245 250 255  
 10 Val Val Tyr Leu Met Asp Met Cys Pro Phe Glu Thr Val Ala Arg Thr  
 260 265 270  
 Ser Asp Ala Thr Glu Leu Ser Pro Phe Cys Ala Leu Phe Thr His Asp  
 275 280 285  
 15 Glu Trp Arg Gln Tyr Asp Tyr Leu Gln Ser Leu Gly Lys Tyr Tyr Gly  
 290 295 300  
 Tyr Gly Ala Gly Asn Pro Leu Gly Pro Ala Gln Gly Val Gly Phe Ala  
 305 310 315 320  
 Asn Glu Leu Ile Ala Arg Leu Thr Arg Ser Pro Val Gln Asp His Thr  
 325 330 335  
 20 Ser Thr Asn His Thr Leu Asp Ser Asn Pro Ala Thr Phe Pro Leu Asn  
 340 345 350  
 Ala Thr Leu Tyr Ala Asp Phe Ser His Asp Asn Ser Met Ile Ser Ile  
 355 360 365  
 Phe Phe Ala Leu Gly Leu Tyr Asn Gly Thr Ala Pro Leu Ser Thr Thr  
 370 375 380  
 25 Ser Val Glu Ser Ile Glu Glu Thr Asp Gly Tyr Ser Ala Ser Trp Thr  
 385 390 395 400  
 Val Pro Phe Gly Ala Arg Ala Tyr Val Glu Met Met Gln Cys Gln Ala  
 405 410 415  
 Glu Lys Glu Pro Leu Val Arg Val Leu Val Asn Asp Arg Val Val Pro  
 420 425 430  
 30 Leu His Gly Cys Ala Val Asp Lys Leu Gly Arg Cys Lys Arg Asp Asp  
 435 440 445  
 Phe Val Glu Gly Leu Ser Phe Ala Arg Ser Gly Gly Asn Trp Ala Glu  
 450 455 460  
 35 Cys Phe Ala  
 465  
  
 <210> 102  
 <211> 1426  
 40 <212> DNA  
 <213> Artificial Sequence  
  
 <220>  
 <223> consensus phytase-1  
  
 45 <400> 102  
  
 tatatgaatt catgggctgt ttcgtcgtgc tactgtccat tgccaccttg ttcggttcca  
 60  
 50 catccggtac cgccttgggt cctcgtggta attctcactc ttgtgacact gttgacggtg  
 120  
 gttaccaatg tttcccagaa atttctcact tgtggggtca atactctcca tacttctctt  
 180  
  
 55

## SEQUENCE LISTING.txt

5      tggaagacga atctgetatt tctccagacg ttccagacga ctgtagagtt actttcgttc  
 240  
 aagttttgtc tagacacggt gctagatacc caacttcttc taagtctaag gcttactctg  
 300  
 ctttgattga agctattcaa aagaacgcta ctgctttcaa gggtaagtac gctttcttga  
 360  
 10      agacttaca ctacactttg ggtgctgacg acttgactcc attcggtgaa aaccaaattg  
 420  
 ttaactctgg tattaagtcc tacagaagat acaaggcttt ggctagaaag attgttccat  
 480  
 tcattagagc ttctggttct gacagagtta ttgcttctgc tgaaaagtcc attgaagggt  
 540  
 15      tccaatctgc taagttggct gacccagggt ctcaaccaca ccaagcttct ccagttattg  
 600  
 acgttattat tccagaagga tccgggtaca acaacacttt ggaccacggt acttgtactg  
 660  
 ctttcgaaga ctctgaattg ggtgacgacg ttgaagctaa cttcactgct ttgttcgctc  
 720  
 20      cagctattag agctagattg gaagctgact tgccagggtg tactttgact gacgaagacg  
 780  
 ttgtttactt gatggacatg tgtccattcg aaactgttgc tagaacttct gacgctactg  
 840  
 aattgtctcc attctgtgct ttgttctctc acgacgaatg gagacaatac gactacttgc  
 900  
 25      aatctttggg taagtactac gggtacggtg ctggtaaccc attgggtcca gctcaagggtg  
 960  
 ttggtttcgc taacgaattg attgctagat tgactagatc tccagttcaa gaccacactt  
 1020  
 30      ctactaacca cactttggac tctaaccag ctactttccc attgaacgct actttgtacg  
 1080  
 ctgacttctc tcacgacaac tctatgattt ctattttctt cgctttgggt ttgtacaacg  
 1140  
 gtactgctcc attgtctact acttctgttg aatctattga agaaactgac ggttactctg  
 1200  
 35      cttcttggac tgttccattc ggtgctagag cttacgttga aatgatgcaa tgtcaagctg  
 1260  
 aaaaggaacc attgggttaga gttttgggta acgacagagt tgttccattg cacgggttg  
 1320  
 ctgttgacaa gttgggtaga tgtaagagag acgacttcgt tgaagggttg tctttcgtc  
 1380  
 40      gatctggtgg taactgggct gaatgtttcg cttagaatt catata  
 1426

45      <210> 103  
 <211> 467  
 <212> PRT  
 <213> Artificial Sequence

50      <220>  
 <223> consensus phytase-10

<400> 103

55

## SEQUENCE LISTING.txt

Met Gly Val Phe Val Val Leu Leu Ser Ile Ala Thr Leu Phe Gly Ser  
 1 5 10 15  
 5 Thr Ser Gly Thr Ala Leu Gly Pro Arg Gly Asn Ser His Ser Cys Asp  
 20 25 30  
 Thr Val Asp Gly Gly Tyr Gln Cys Phe Pro Glu Ile Ser His Leu Trp  
 35 40 45  
 Gly Gln Tyr Ser Pro Phe Phe Ser Leu Ala Asp Glu Ser Ala Ile Ser  
 50 55 60  
 10 Pro Asp Val Pro Lys Gly Cys Arg Val Thr Phe Val Gln Val Leu Ser  
 65 70 75 80  
 Arg His Gly Ala Arg Tyr Pro Thr Ser Ser Lys Ser Lys Lys Tyr Ser  
 85 90 95  
 15 Ala Leu Ile Glu Ala Ile Gln Lys Asn Ala Thr Ala Phe Lys Gly Lys  
 100 105 110  
 Tyr Ala Phe Leu Lys Thr Tyr Asn Tyr Thr Leu Gly Ala Asp Asp Leu  
 115 120 125  
 Thr Pro Phe Gly Glu Gln Gln Met Val Asn Ser Gly Ile Lys Phe Tyr  
 130 135 140  
 20 Arg Arg Tyr Lys Ala Leu Ala Arg Lys Ile Val Pro Phe Val Arg Ala  
 145 150 155 160  
 Ser Gly Ser Asp Arg Val Ile Ala Ser Ala Glu Lys Phe Ile Glu Gly  
 165 170 175  
 Phe Gln Ser Ala Lys Leu Ala Asp Pro Gly Ala Asn Pro His Gln Ala  
 180 185 190  
 25 Ser Pro Val Ile Asn Val Ile Ile Pro Glu Gly Ala Gly Tyr Asn Asn  
 195 200 205  
 Thr Leu Asp His Gly Leu Cys Thr Ala Phe Glu Glu Ser Glu Leu Gly  
 210 215 220  
 Asp Asp Val Glu Ala Asn Phe Thr Ala Val Phe Ala Pro Pro Ile Arg  
 225 230 235 240  
 30 Ala Arg Leu Glu Ala His Leu Pro Gly Val Asn Leu Thr Asp Glu Asp  
 245 250 255  
 Val Val Asn Leu Met Asp Met Cys Pro Phe Asp Thr Val Ala Arg Thr  
 260 265 270  
 Ser Asp Ala Thr Gln Leu Ser Pro Phe Cys Asp Leu Phe Thr His Asp  
 275 280 285  
 35 Glu Trp Ile Gln Tyr Asp Tyr Leu Gln Ser Leu Gly Lys Tyr Tyr Gly  
 290 295 300  
 Tyr Gly Ala Gly Asn Pro Leu Gly Pro Ala Gln Gly Val Gly Phe Val  
 305 310 315 320  
 40 Asn Glu Leu Ile Ala Arg Leu Thr His Ser Pro Val Gln Asp His Thr  
 325 330 335  
 Ser Thr Asn His Thr Leu Asp Ser Asn Pro Ala Thr Phe Pro Leu Asn  
 340 345 350  
 Ala Thr Leu Tyr Ala Asp Phe Ser His Asp Asn Thr Met Val Ser Ile  
 355 360 365  
 45 Phe Phe Ala Leu Gly Leu Tyr Asn Gly Thr Lys Pro Leu Ser Thr Thr  
 370 375 380  
 Ser Val Glu Ser Ile Glu Glu Thr Asp Gly Tyr Ala Ala Ser Trp Thr  
 385 390 395 400  
 Val Pro Phe Ala Ala Arg Ala Tyr Val Glu Met Met Gln Cys Glu Ala  
 405 410 415  
 50 Glu Lys Glu Pro Leu Val Arg Val Leu Val Asn Asp Arg Val Val Pro  
 420 425 430  
 Leu His Gly Cys Gly Val Asp Lys Leu Gly Arg Cys Lys Arg Asp Asp

## SEQUENCE LISTING.txt

435 440 445  
 Phe Val Glu Gly Leu Ser Phe Ala Arg Ser Gly Gly Asn Trp Glu Glu  
 5 450 455 460  
 Cys Phe Ala  
 465

10 <210> 104  
 <211> 1426  
 <212> DNA  
 <213> Artificial Sequence

15 <220>  
 <223> consensus phytase-10  
 <400> 104

20 tatatgaatt catgggcgtg ttcgtcgtgc tactgtccat tgccaccttg ttcggttcca  
 60  
 catccggtac cgccttgggt cctcgtggta attctcactc ttgtgacact gttgacgggtg  
 120  
 gttaccaatg tttcccagaa atttctcact tgtgggggtca atactctcca ttcttctctt  
 180  
 25 tggctgacga atctgctatt tctccagacg ttccaaaggg ttgtagagtt actttcgttc  
 240  
 aagttttgtc tagacacggg gctagatacc caacttcttc taagtctaag aagtactctg  
 300  
 ctttgattga agctattcaa aagaacgcta ctgctttcaa gggtaagtac gctttcttga  
 360  
 30 agacttacia ctacactttg ggtgctgacg acttgactcc attcggtgaa caacaaatgg  
 420  
 ttaactctgg tattaagttc tacagaagat acaaggcttt ggctagaaag attgttccat  
 480  
 35 tcgttagagc ttctgggttct gacagagtta ttgcttctgc tgaaaagtcc attgaagggtt  
 540  
 tccaatctgc taagttgggt gaccaggtg ctaaccacaca ccaagcttct ccagttatta  
 600  
 acgttattat tccagaaggt gctgggttaca acaacacttt ggaccacggg ttgtgtactg  
 660  
 40 ctttcgaaga atctgaattg ggtgacgacg ttgaagctaa cttcactgct gttttcgttc  
 720  
 cacctattag agctagattg gaagctcact tgccaggtgt taacttgact gacgaagacg  
 780  
 ttgttaactt gatggacatg tgtccattcg aactgttgc tagaacttct gacgctactc  
 840  
 45 aattgtctcc attctgtgac ttgttctc acgacgaatg gattcaatac gactacttgc  
 900  
 aatctttggg taagtactac gggttacgggt ctggtaaccc attgggtcca gctcaagggtg  
 960  
 ttggtttcgt taacgaattg attgctagat tgactcactc tccagttcaa gaccacactt  
 1020  
 50 ctactaacca cactttggac tctaaccag ctactttccc attgaacgct actttgtacg  
 1080  
 ctgacttctc tcacgacaac actatggttt ctattttctt cgctttgggt ttgtacaacg

55

## SEQUENCE LISTING.txt

1140  
 gtactaagcc attgtctact acttctgttg aatctattga agaaactgac ggttacgctg  
 5 1200  
 cttcttggac tgttccattc gctgctagag cttacgttga aatgatgcaa tgtgaagctg  
 1260  
 aaaaggaacc attgggttaga gttttgggta acgacagagt tgttccattg cacgggttg  
 1320  
 gtgttgacaa gttgggtaga tgtaagagag acgacttcgt tgaagggttg tctttcgtc  
 10 1380  
 gatctgggtg taactgggaa gaatgtttcg cttaagaatt catata  
 1426

15 <210> 105  
 <211> 467  
 <212> PRT  
 <213> Artificial Sequence

20 <220>  
 <223> consensus phytase-1-thermo[8]-Q50T-K91A  
 <400> 105

25 Met Gly Val Phe Val Val Leu Leu Ser Ile Ala Thr Leu Phe Gly Ser  
 1 5 10 15  
 Thr Ser Gly Thr Ala Leu Gly Pro Arg Gly Asn Ser His Ser Cys Asp  
 20 25 30  
 Thr Val Asp Gly Gly Tyr Gln Cys Phe Pro Glu Ile Ser His Leu Trp  
 35 35 40 45  
 Gly Thr Tyr Ser Pro Tyr Phe Ser Leu Ala Asp Glu Ser Ala Ile Ser  
 50 55 60  
 Pro Asp Val Pro Asp Asp Cys Arg Val Thr Phe Val Gln Val Leu Ser  
 65 70 75 80  
 Arg His Gly Ala Arg Tyr Pro Thr Ser Ser Ala Ser Lys Ala Tyr Ser  
 35 85 90 95  
 Ala Leu Ile Glu Ala Ile Gln Lys Asn Ala Thr Ala Phe Lys Gly Lys  
 100 105 110  
 Tyr Ala Phe Leu Lys Thr Tyr Asn Tyr Thr Leu Gly Ala Asp Asp Leu  
 115 120 125  
 Thr Pro Phe Gly Glu Asn Gln Met Val Asn Ser Gly Ile Lys Phe Tyr  
 40 130 135 140  
 Arg Arg Tyr Lys Ala Leu Ala Arg Lys Ile Val Pro Phe Ile Arg Ala  
 145 150 155 160  
 Ser Gly Ser Asp Arg Val Ile Ala Ser Ala Glu Lys Phe Ile Glu Gly  
 165 170 175  
 45 Phe Gln Ser Ala Lys Leu Ala Asp Pro Gly Ser Gln Pro His Gln Ala  
 180 185 190  
 Ser Pro Val Ile Asn Val Ile Ile Pro Glu Gly Ser Gly Tyr Asn Asn  
 195 200 205  
 Thr Leu Asp His Gly Thr Cys Thr Ala Phe Glu Asp Ser Glu Leu Gly  
 210 215 220  
 50 Asp Asp Val Glu Ala Asn Phe Thr Ala Leu Phe Ala Pro Ala Ile Arg  
 225 230 235 240  
 Ala Arg Leu Glu Ala Asp Leu Pro Gly Val Thr Leu Thr Asp Glu Asp

55

## SEQUENCE LISTING.txt

5 Val Val Tyr Leu Met Asp Met Cys Pro Phe Asp Thr Val Ala Arg Thr  
 245 250 255  
 260 265 270  
 Ser Asp Ala Thr Glu Leu Ser Pro Phe Cys Ala Leu Phe Thr His Asp  
 275 280 285  
 Glu Trp Ile Gln Tyr Asp Tyr Leu Gln Ser Leu Gly Lys Tyr Tyr Gly  
 290 295 300  
 10 Tyr Gly Ala Gly Asn Pro Leu Gly Pro Ala Gln Gly Val Gly Phe Ala  
 305 310 315 320  
 Asn Glu Leu Ile Ala Arg Leu Thr His Ser Pro Val Gln Asp His Thr  
 325 330 335  
 Ser Thr Asn His Thr Leu Asp Ser Asn Pro Ala Thr Phe Pro Leu Asn  
 340 345 350  
 15 Ala Thr Leu Tyr Ala Asp Phe Ser His Asp Asn Thr Met Ile Ser Ile  
 355 360 365  
 Phe Phe Ala Leu Gly Leu Tyr Asn Gly Thr Lys Pro Leu Ser Thr Thr  
 370 375 380  
 Ser Val Glu Ser Ile Glu Gly Thr Asp Gly Tyr Ser Ala Ser Trp Thr  
 385 390 395 400  
 20 Val Pro Phe Ala Ala Arg Ala Tyr Val Glu Met Met Gln Cys Gln Ala  
 405 410 415  
 Glu Lys Glu Pro Leu Val Arg Val Leu Val Asn Asp Arg Val Val Pro  
 420 425 430  
 Leu His Gly Cys Ala Val Asp Lys Leu Gly Arg Cys Lys Arg Asp Asp  
 435 440 445  
 25 Phe Val Glu Gly Leu Ser Phe Ala Arg Ser Gly Gly Asn Trp Ala Glu  
 450 455 460  
 Cys Phe Ala  
 465  
 30  
 <210> 106  
 <211> 1404  
 <212> DNA  
 <213> Artificial Sequence  
 35  
 <220>  
 <223> consensus phytase-1-thermo[8]-Q50T-K91A  
 <400> 106  
 40  
 atgggcgtgt tcgtcgtgct actgtccatt gccaccttgt tcggttccac atccggtacc  
 60  
 gccttggggc ctcgtggtaa ttctcactct tgtgacactg ttgacgggtgg ttaccaatgt  
 120  
 45 ttcccagaaa tttctcactt gtgggggtacc tactctccat acttctcttt ggcagacgaa  
 180  
 tctgctatct ctccagacgt tccagacgac tgtagagtta ctttcgttca agttttgtct  
 240  
 agacacggtg ctagataccc aacttcttct gcggtctaagg cttactctgc tttgattgaa  
 300  
 50 gctattcaaa agaacgctac tgctttcaag ggtaagtacg ctttcttgaa gacttacaac  
 360  
 tacactttgg gtgctgacga cttgactcca ttcggtgaaa accaaatggt taactctggt  
 55

## SEQUENCE LISTING.txt

420  
 5   attaagttct acagaagata caaggctttg gctagaaaga ttgttccatt cattagagct  
 480  
 tctggttctg acagagttat tgcttctgct gaaaagttca ttgaaggttt ccaatctgct  
 540  
 aagttggctg acccaggttc tcaaccacac caagcttctc cagttattaa cgtgatcatt  
 600  
 10   ccagaaggat ccggttacaa caacactttg gaccacggta cttgtactgc tttcgaagac  
 660  
 tctgaattag gtgacgacgt tgaagctaac ttcactgctt tggtcgctcc agctattaga  
 720  
 gctagattgg aagctgactt gccagggtgt actttgactg acgaagacgt tgtttacttg  
 780  
 15   atggacatgt gtccattcga cactgtcgct agaacttctg acgctactga attgtctcca  
 840  
 ttctgtgctt tgttcactca cgacgaatgg atccaatacg actacttgca aagcttgggt  
 900  
 aagtactacg gttacgggtgc tggtaaccca ttgggtccag ctcaagggtg tggtttcgct  
 960  
 20   aacgaattga ttgctagatt gactcactct ccagttcaag accacacttc tactaaccac  
 1020  
 actttggact ctaaccacgc tactttccca ttgaacgcta ctttgtacgc tgacttctct  
 1080  
 25   cacgacaaca ctatgatata tattttcttc gctttgggtt tgtacaacgg taccaagcca  
 1140  
 ttgtctacta cttctgttga atctattgaa gaaactgacg gttactctgc ttcttggact  
 1200  
 gttccattcg ctgctagagc ttacgttgaa atgatgcaat gtcaagctga aaaggaacca  
 1260  
 30   ttggtttagag ttttgggttaa cgacagagtt gttccattgc acggttgtgc tgttgacaag  
 1320  
 ttgggtagat gtaagagaga cgacttcgtt gaaggtttgt ctttcgctag atctggtggt  
 1380  
 aactgggctg aatgtttcgc ttaa  
 1404  
 35  
  
 <210> 107  
 <211> 467  
 <212> PRT  
 40   <213> Artificial Sequence  
  
 <220>  
 <223> consensus phytase-10-thermo[3]-Q50T-K91A  
  
 45   <400> 107  
  
 Met Gly Val Phe Val Val Leu Leu Ser Ile Ala Thr Leu Phe Gly Ser  
 1                   5                   10                   15  
 Thr Ser Gly Thr Ala Leu Gly Pro Arg Gly Asn Ser His Ser Cys Asp  
 20                   25                   30  
 50   Thr Val Asp Gly Gly Tyr Gln Cys Phe Pro Glu Ile Ser His Leu Trp  
 35                   40                   45  
 Gly Thr Tyr Ser Pro Phe Phe Ser Leu Ala Asp Glu Ser Ala Ile Ser

55

## SEQUENCE LISTING.txt

5           50                   55                   60  
 Pro Asp Val Pro Lys Gly Cys Arg Val Thr Phe Val Gln Val Leu Ser  
   65                   70                   75                   80  
 Arg His Gly Ala Arg Tyr Pro Thr Ser Ser Ala Ser Lys Ala Tyr Ser  
                   85                   90                   95  
 10   Ala Leu Ile Glu Ala Ile Gln Lys Asn Ala Thr Ala Phe Lys Gly Lys  
                   100                   105                   110  
 Tyr Ala Phe Leu Lys Thr Tyr Asn Tyr Thr Leu Gly Ala Asp Asp Leu  
                   115                   120                   125  
 Thr Pro Phe Gly Glu Gln Gln Met Val Asn Ser Gly Ile Lys Phe Tyr  
                   130                   135                   140  
 15   Arg Arg Tyr Lys Ala Leu Ala Arg Lys Ile Val Pro Phe Ile Arg Ala  
   145                   150                   155                   160  
 Ser Gly Ser Asp Arg Val Ile Ala Ser Ala Glu Lys Phe Ile Glu Gly  
                   165                   170                   175  
 Phe Gln Ser Ala Lys Leu Ala Asp Pro Gly Ala Asn Pro His Gln Ala  
                   180                   185                   190  
 20   Ser Pro Val Ile Asn Val Ile Ile Pro Glu Gly Ala Gly Tyr Asn Asn  
                   195                   200                   205  
 Thr Leu Asp His Gly Leu Cys Thr Ala Phe Glu Glu Ser Glu Leu Gly  
                   210                   215                   220  
 25   Asp Asp Val Glu Ala Asn Phe Thr Ala Val Phe Ala Pro Pro Ile Arg  
   225                   230                   235                   240  
 Ala Arg Leu Glu Ala His Leu Pro Gly Val Asn Leu Thr Asp Glu Asp  
                   245                   250                   255  
 Val Val Asn Leu Met Asp Met Cys Pro Phe Asp Thr Val Ala Arg Thr  
                   260                   265                   270  
 30   Ser Asp Ala Thr Gln Leu Ser Pro Phe Cys Asp Leu Phe Thr His Asp  
                   275                   280                   285  
 Glu Trp Ile Gln Tyr Asp Tyr Leu Gln Ser Leu Gly Lys Tyr Tyr Gly  
                   290                   295                   300  
 Tyr Gly Ala Gly Asn Pro Leu Gly Pro Ala Gln Gly Val Gly Phe Val  
                   305                   310                   315                   320  
 35   Asn Glu Leu Ile Ala Arg Leu Thr His Ser Pro Val Gln Asp His Thr  
                   325                   330                   335  
 Ser Thr Asn His Thr Leu Asp Ser Asn Pro Ala Thr Phe Pro Leu Asn  
                   340                   345                   350  
 Ala Thr Leu Tyr Ala Asp Phe Ser His Asp Asn Thr Met Val Ser Ile  
                   355                   360                   365  
 40   Phe Phe Ala Leu Gly Leu Tyr Asn Gly Thr Lys Pro Leu Ser Thr Thr  
                   370                   375                   380  
 Ser Val Glu Ser Ile Glu Glu Thr Asp Gly Tyr Ser Ala Ser Trp Thr  
                   385                   390                   395                   400  
 45   Val Pro Phe Ala Ala Arg Ala Tyr Val Glu Met Met Gln Cys Glu Ala  
                   405                   410                   415  
 Glu Lys Glu Pro Leu Val Arg Val Leu Val Asn Asp Arg Val Val Pro  
                   420                   425                   430  
 Leu His Gly Cys Gly Val Asp Lys Leu Gly Arg Cys Lys Arg Asp Asp  
                   435                   440                   445  
 50   Phe Val Glu Gly Leu Ser Phe Ala Arg Ser Gly Gly Asn Trp Glu Glu  
                   450                   455                   460  
 Cys Phe Ala  
   465



## SEQUENCE LISTING.txt

<210> 108  
 <211> 1404  
 <212> DNA  
 <213> Artificial Sequence  
  
 <220>  
 <223> consensus phytase-10-thermo-[3]-Q50T-K91A  
  
 <400> 108

```

atgggcgtgt tgcgtcgtgct actgtccatt gccaccttgt tcggttccac atccggtacc
60
gccttggggtc ctcgtggtaa ctctcactct tgtgacactg ttgacgggtg ttaccaatgt
120
ttcccagaaa tttctcactt gtgggggtaca tactctccat tcttctcttt ggctgacgaa
180
tctgctatct ctccagacgt tccaaagggg tgtagagtta ctttcggttca agttttgtct
240
agacacgggtg ctagataccc aacttcttct gcgtctaagg cgtactctgc tttgattgaa
300
gctattcaaaa agaacgctac tgcttttcaag ggtaagtacg ctttcttgaa gacttacaac
360
tacacttttg gtgctgacga cttgactcca ttcgggtgaac aacaaatggt taactctggt
420
attaagttct acagaagata caaggctttg gctagaaaga ttgttccatt cattagagct
480
tctgggtctg acagagttat tgcttctgct gaaaagtcca ttgaagggtt ccaatctgct
540
aagttggctg acccaggtgc taaccacac caagcttctc cagttattaa cgttattatt
600
ccagaagggtg ctggttacaa caacactttg gaccacgggt tgtgtactgc tttcgaagaa
660
tctgaattgg gtgacgacgt tgaagctaac ttcactgctg ttttcgctcc accaattaga
720
gctagattgg aagctcactt gccaggtggt aacttgactg acgaagacgt tgttaacttg
780
atggacatgt gtccattcga cactgttgct agaacttctg acgctactca attgtctcca
840
ttctgtgact tgttcactca cgacgaatgg attcaatacg actacttgca atctttgggt
900
aagtactacg gttacgggtg tggtaaccca ttgggtccag ctcaagggtg tggtttcggt
960
aacgaattga ttgctagatt gactcactct ccagttcaag accacacttc tactaaccac
1020
actttggact ctaaccacgc tactttccca ttgaacgcta ctttgtacgc tgacttctct
1080
cacgacaaca ctatggtttc tattttcttc gctttgggtt tgtacaacgg tactaagcca
1140
ttgtctacta cttctgttga atctattgaa gaaactgacg gttactctgc ttcttggtgact
1200
gttccattcg ctgctagagc ttacgttgaa atgatgcaat gtgaagctga aaaggaacca
1260
ttgggttagag ttttggttaa cgacagagtt gttccattgc acgggttggtg tgttgacaag
1320

```

## SEQUENCE LISTING.txt

ttgggtagat gtaagagaga cgacttcggt gaagggttgt ctttcgctag atctggtggg  
1380

aactgggaag aatgtttcgc ttaa  
1404

<210> 109

<211> 467

<212> PRT

<213> Artificial Sequence

<220>

<223> A.fumigatus ATCC 13073 phytase alpha-mutant

<400> 109

Met	Gly	Val	Phe	Val	Val	Leu	Leu	Ser	Ile	Ala	Thr	Leu	Phe	Gly	Ser
1				5					10					15	
Thr	Ser	Gly	Thr	Ala	Leu	Gly	Pro	Arg	Gly	Asn	His	Ser	Lys	Ser	Cys
			20					25					30		
Asp	Thr	Val	Asp	Leu	Gly	Tyr	Gln	Cys	Ser	Pro	Ala	Thr	Ser	His	Leu
		35				40					45				
Trp	Gly	Thr	Tyr	Ser	Pro	Tyr	Phe	Ser	Leu	Glu	Asp	Glu	Leu	Ser	Val
	50					55				60					
Ser	Ser	Lys	Leu	Pro	Lys	Asp	Cys	Arg	Ile	Thr	Leu	Val	Gln	Val	Leu
	65				70					75					80
Ser	Arg	His	Gly	Ala	Arg	Tyr	Pro	Thr	Ser	Ser	Lys	Ser	Lys	Lys	Tyr
			85					90					95		
Lys	Lys	Leu	Ile	Thr	Ala	Ile	Gln	Ala	Asn	Ala	Thr	Asp	Phe	Lys	Gly
		100					105						110		
Lys	Tyr	Ala	Phe	Leu	Lys	Thr	Tyr	Asn	Tyr	Thr	Leu	Gly	Ala	Asp	Asp
	115					120					125				
Leu	Thr	Pro	Phe	Gly	Glu	Gln	Gln	Leu	Val	Asn	Ser	Gly	Ile	Lys	Phe
	130				135					140					
Tyr	Gln	Arg	Tyr	Lys	Ala	Leu	Ala	Arg	Ser	Val	Val	Pro	Phe	Ile	Arg
	145			150					155						160
Ala	Ser	Gly	Ser	Asp	Arg	Val	Ile	Ala	Ser	Gly	Glu	Lys	Phe	Ile	Glu
			165				170							175	
Gly	Phe	Gln	Gln	Ala	Lys	Leu	Ala	Asp	Pro	Gly	Ala	Thr	Asn	Arg	Ala
		180					185						190		
Ala	Pro	Ala	Ile	Ser	Val	Ile	Ile	Pro	Glu	Ser	Glu	Thr	Phe	Asn	Asn
	195					200					205				
Thr	Leu	Asp	His	Gly	Val	Cys	Thr	Lys	Phe	Glu	Ala	Ser	Gln	Leu	Gly
	210				215					220					
Asp	Glu	Val	Ala	Ala	Asn	Phe	Thr	Ala	Leu	Phe	Ala	Pro	Asp	Ile	Arg
	225				230				235					240	
Ala	Arg	Leu	Glu	Lys	His	Leu	Pro	Gly	Val	Thr	Leu	Thr	Asp	Glu	Asp
			245				250						255		
Val	Val	Ser	Leu	Met	Asp	Met	Cys	Pro	Phe	Asp	Thr	Val	Ala	Arg	Thr
		260					265					270			
Ser	Asp	Ala	Ser	Gln	Leu	Ser	Pro	Phe	Cys	Gln	Leu	Phe	Thr	His	Asn
		275				280					285				
Glu	Trp	Lys	Lys	Tyr	Asp	Tyr	Leu	Gln	Ser	Leu	Gly	Lys	Tyr	Tyr	Gly
	290					295					300				

## SEQUENCE LISTING.txt

5 Tyr Gly Ala Gly Asn Pro Leu Gly Pro Ala Gln Gly Ile Gly Phe Thr  
 305 310 315 320  
 Asn Glu Leu Ile Ala Arg Leu Thr Arg Ser Pro Val Gln Asp His Thr  
 325 330 335  
 Ser Thr Asn Ser Thr Leu Val Ser Asn Pro Ala Thr Phe Pro Leu Asn  
 340 345 350  
 10 Ala Thr Met Tyr Val Asp Phe Ser His Asp Asn Ser Met Val Ser Ile  
 355 360 365  
 Phe Phe Ala Leu Gly Leu Tyr Asn Gly Thr Glu Pro Leu Ser Arg Thr  
 370 375 380  
 Ser Val Glu Ser Ala Lys Glu Leu Asp Gly Tyr Ser Ala Ser Trp Val  
 385 390 395 400  
 15 Val Pro Phe Gly Ala Arg Ala Tyr Phe Glu Thr Met Gln Cys Lys Ser  
 405 410 415  
 Glu Lys Glu Pro Leu Val Arg Ala Leu Ile Asn Asp Arg Val Val Pro  
 420 425 430  
 Leu His Gly Cys Asp Val Asp Lys Leu Gly Arg Cys Lys Leu Asn Asp  
 435 440 445  
 20 Phe Val Lys Gly Leu Ser Trp Ala Arg Ser Gly Gly Asn Trp Gly Glu  
 450 455 460  
 Cys Phe Ser  
 465  
 25  
 <210> 110  
 <211> 1404  
 <212> DNA  
 <213> Artificial Sequence  
 30  
 <220>  
 <223> A.fumigatus ATCC 13073 phytase alpha-mutant  
 <400> 110  
 35  
 atggggggttt tcgtcgttct attatctatc gcgactctgt tcggcagcac atcgggcact  
 60  
 gcgctggggcc cccgtggaaa tcaactccaag tcctgcgata cggtagacct aggggtaccag  
 120  
 tgctcccctg cgactttctca tctatggggc acgtactcgc catacttttc gctcgaggac  
 180  
 40 gagctgtccg tgtcgagtaa gtttcccaag gattgccgga tcaccttggg acaggtgcta  
 240  
 tcgcgccatg gagegcggta cccaaccagc tccaagagca aaaagtataa gaagcttatt  
 300  
 45 acggcgatcc aggccaatgc caccgacttc aagggaagt acgccttttt gaagacgtac  
 360  
 aactatactc tgggtgcgga tgacctcact ccctttgggg agcagcagct ggtgaactcg  
 420  
 ggcatcaagt tctaccagag gtacaaggct ctggcgcgca gtgtggtgcc gtttatcg  
 480  
 50 gcctcaggct cggaccgggt tattgcttcg ggagagaagt tcatcgaggg gttccagcag  
 540  
 gcgaagctgg ctgatcctgg cgcgaagaac cgcgccgctc cggcgattag tgtgattatt  
 600

55

## SEQUENCE LISTING.txt

cccgagagcg agacgttcaa caatacgtcg gaccacggcg tgtgcaacgaa gtttgaggcg  
 660  
 5 agtcagctgg gagatgaggt tgcggccaat ttcactgcgc tctttgcacc cgacatccga  
 720  
 gctcgctcg agaagcatct tcttgccgtg acgctgacag acgaggacgt tgtcagtcta  
 780  
 atggacatgt gtccgtttga tacggtagcg cgcaccagcg acgcaagtca gctgtcaccg  
 840  
 10 tttctgtcaac tcttcactca caatgagtgg aagaagtacg actaccttca gtccttgggc  
 900  
 aagtactacg gctacggcgc aggcaaccct ctgggaccgg ctcaggggat agggttcacc  
 960  
 15 aacgagctga ttgcccgggt gacgcgttcg ccagtgcagg accacaccag cactaactcg  
 1020  
 actctagtct ccaacccggc caccttcccg ttgaacgcta ccatgtacgt cgacttttca  
 1080  
 caccacaaca gcatggtttc catcttcttt gcattggggc tgtacaacgg cactgaaccc  
 1140  
 20 ttgtcccga cctcggtgga aagcgccaag gaattggatg ggtattctgc atcctgggtg  
 1200  
 gtgcctttcg gcgcgcgagc ctacttcgag acgatgcaat gcaagtcgga aaaggagcct  
 1260  
 cttgttcgcg ctttgattaa tgaccgggtt gtgccactgc atggctgcga tgtggacaag  
 1320  
 25 ctggggcgat gcaagctgaa tgactttgtc aagggttga gttggggccag atctgggggc  
 1380  
 aactggggag agtgcttttag ttga  
 1404  
 30  
 <210> 111  
 <211> 467  
 <212> PRT  
 <213> Artificial Sequence  
 35  
 <220>  
 <223> consensus phytase-7  
 <400> 111  
 40  
 Met Gly Val Phe Val Val Leu Leu Ser Ile Ala Thr Leu Phe Gly Ser  
 1 5 10 15  
 Thr Ser Gly Thr Ala Leu Gly Pro Arg Gly Asn Ser His Ser Cys Asp  
 20 25 30  
 Thr Val Asp Gly Gly Tyr Gln Cys Phe Pro Glu Ile Ser His Leu Trp  
 35 40 45  
 45 Gly Gln Tyr Ser Pro Tyr Phe Ser Leu Glu Asp Glu Ser Ala Ile Ser  
 50 55 60  
 Pro Asp Val Pro Asp Asp Cys Arg Val Thr Phe Val Gln Val Leu Ser  
 65 70 75 80  
 50 Arg His Gly Ala Arg Tyr Pro Thr Asp Ser Lys Gly Lys Lys Tyr Ser  
 85 90 95  
 Ala Leu Ile Glu Ala Ile Gln Lys Asn Ala Thr Ala Phe Lys Gly Lys  
 100 105 110  
 55

## SEQUENCE LISTING.txt

Tyr Ala Phe Leu Lys Thr Tyr Asn Tyr Thr Leu Gly Ala Asp Asp Leu  
 115 120 125  
 5 Thr Pro Phe Gly Glu Asn Gln Met Val Asn Ser Gly Ile Lys Phe Tyr  
 130 135 140  
 Arg Arg Tyr Lys Ala Leu Ala Arg Lys Ile Val Pro Phe Ile Arg Ala  
 145 150 155 160  
 Ser Gly Ser Ser Arg Val Ile Ala Ser Ala Glu Lys Phe Ile Glu Gly  
 165 170 175  
 10 Phe Gln Ser Ala Lys Leu Ala Asp Pro Gly Ser Gln Pro His Gln Ala  
 180 185 190  
 Ser Pro Val Ile Asp Val Ile Ile Ser Glu Ala Ser Ser Tyr Asn Asn  
 195 200 205  
 Thr Leu Asp Pro Gly Thr Cys Thr Ala Phe Glu Asp Ser Glu Leu Ala  
 210 215 220  
 15 Asp Thr Val Glu Ala Asn Phe Thr Ala Leu Phe Ala Pro Ala Ile Arg  
 225 230 235 240  
 Ala Arg Leu Glu Ala Asp Leu Pro Gly Val Thr Leu Thr Asp Thr Glu  
 245 250 255  
 20 Val Thr Tyr Leu Met Asp Met Cys Ser Phe Glu Thr Val Ala Arg Thr  
 260 265 270  
 Ser Asp Ala Thr Glu Leu Ser Pro Phe Cys Ala Leu Phe Thr His Asp  
 275 280 285  
 Glu Trp Arg His Tyr Asp Tyr Leu Gln Ser Leu Lys Lys Tyr Tyr Gly  
 290 295 300  
 25 His Gly Ala Gly Asn Pro Leu Gly Pro Thr Gln Gly Val Gly Phe Ala  
 305 310 315 320  
 Asn Glu Leu Ile Ala Arg Leu Thr Arg Ser Pro Val Gln Asp His Thr  
 325 330 335  
 Ser Thr Asn His Thr Leu Asp Ser Asn Pro Ala Thr Phe Pro Leu Asn  
 340 345 350  
 30 Ala Thr Leu Tyr Ala Asp Phe Ser His Asp Asn Gly Ile Ile Ser Ile  
 355 360 365  
 Phe Phe Ala Leu Gly Leu Tyr Asn Gly Thr Ala Pro Leu Ser Thr Thr  
 370 375 380  
 35 Ser Val Glu Ser Ile Glu Glu Thr Asp Gly Tyr Ser Ser Ala Trp Thr  
 385 390 395 400  
 Val Pro Phe Ala Ser Arg Ala Tyr Val Glu Met Met Gln Cys Gln Ala  
 405 410 415  
 Glu Lys Glu Pro Leu Val Arg Val Leu Val Asn Asp Arg Val Val Pro  
 420 425 430  
 40 Leu His Gly Cys Ala Val Asp Lys Leu Gly Arg Cys Lys Arg Asp Asp  
 435 440 445  
 Phe Val Glu Gly Leu Ser Phe Ala Arg Ser Gly Gly Asn Trp Ala Glu  
 450 455 460  
 Cys Phe Ala  
 465  
 45  
 <210> 112  
 <211> 1426  
 <212> DNA  
 50 <213> Artificial Sequence  
 <220>  
 <223> consensus phytase-7  
 55

## SEQUENCE LISTING.txt

&lt;400&gt; 112

5

tatatgaatt catgggcgtg ttcgtcgtgc tactgtccat tgccaccttg ttcgggtcca  
 60  
 catccgggtac cgccttgggt cctcgtggta attctcactc ttgtgacact gttgacggtg  
 120  
 gttaccaatg tttcccagaa atttctcact tgtgggggtca atactctcca tacttctctt  
 180  
 tggaagacga atctgctatt tctccagacg ttccagacga ctgtagagtt actttcgttc  
 240  
 aagttttgtc tagacacggg gctagatacc caactgactc taagggttaag aagtactctg  
 300  
 ctttgattga agctattcaa aagaacgcta ctgctttcaa gggtaagtac gctttcttga  
 360  
 agacttacia ctacactttg ggtgctgacg acttgactcc attcgggtgaa aaccaaattg  
 420  
 ttaactctgg tattaagttc tacagaagat acaaggcttt ggctagaaag attgttccat  
 480  
 tcattagagc ttctgggttct tctagagtta ttgcttctgc tgaaaagtgc attgaagggt  
 540  
 tccaatctgc taagttgggt gaccaggtt ctcaaccaca ccaagcttct ccagttattg  
 600  
 acgttattat ttctgacgct tcttcttaca acaacacttt ggaccaggt acttgactg  
 660  
 ctttcgaaga ctctgaattg gctgacactg ttgaagctaa cttcactgct ttgttcgctc  
 720  
 cagctattag agctagattg gaagctgact tgccaggtgt tactttgact gacactgaag  
 780  
 ttacttactt gatggacatg tgttctttcg aaactgttgc tagaacttct gacgctactg  
 840  
 aattgtctcc attctgtgct ttgttctcctc acgacgaatg gagacactac gactacttgc  
 900  
 aatctttgaa gaagtactac ggtcacgggtg ctggtaacct attgggtcca actcaagggtg  
 960  
 ttggtttcgc taacgaattg attgctagat tgactagatc tccagttcaa gaccacactt  
 1020  
 ctactaacca cactttggac tctaaccag ctactttccc attgaacgct actttgtacg  
 1080  
 ctgacttctc tcacgacaac ggtattatct ctattttctt cgctttgggt ttgtacaacg  
 1140  
 gtactgctcc attgtctact acttctgttg aatctattga agaaactgac ggttactctt  
 1200  
 ctgcttggac tgttccattc gcttctagag cttacgttga aatgatgcaa tgtcaagctg  
 1260  
 aaaaggaacc attgggttaga gttttgggtta acgacagagt tgttccattg cacgggtgtg  
 1320  
 ctgttgacaa gttgggttaga tgtaagagag acgacttcgt tgaagggttg tctttcgcta  
 1380  
 gatctgggtg taactgggct gaatgtttcg cttaagaatt catata  
 1426

50

&lt;210&gt; 113

&lt;211&gt; 467

55

## SEQUENCE LISTING.txt

&lt;212&gt; PRT

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;223&gt; consensus phytase-12

&lt;400&gt; 113

```

5
10 Met Gly Val Phe Val Val Leu Leu Ser Ile Ala Thr Leu Phe Gly Ser
    1          5          10          15
    Thr Ser Gly Thr Ala Leu Gly Pro Arg Gly Asn Ser His Ser Cys Asp
    20          25          30
15 Thr Val Asp Gly Gly Tyr Gln Cys Phe Pro Glu Ile Ser Ser Asn Trp
    35          40          45
    Ser Pro Tyr Ser Pro Tyr Phe Ser Leu Ala Asp Glu Ser Ala Ile Ser
    50          55          60
20 Pro Asp Val Pro Lys Gly Cys Arg Val Thr Phe Val Gln Val Leu Gln
    65          70          75          80
    Arg His Gly Ala Arg Phe Pro Thr Ser Gly Ala Ala Thr Arg Ile Ser
    85          90          95
25 Ala Leu Ile Glu Ala Ile Gln Lys Asn Ala Thr Ala Phe Lys Gly Lys
    100          105          110
    Tyr Ala Phe Leu Lys Thr Tyr Asn Tyr Thr Leu Gly Ala Asp Asp Leu
    115          120          125
30 Val Pro Phe Gly Ala Asn Gln Ser Ser Gln Ala Gly Ile Lys Phe Tyr
    130          135          140
35 Arg Arg Tyr Lys Ala Leu Ala Arg Lys Ile Val Pro Phe Ile Arg Ala
    145          150          155          160
    Ser Gly Ser Asp Arg Val Ile Asp Ser Ala Thr Asn Trp Ile Glu Gly
    165          170          175
40 Phe Gln Ser Ala Lys Leu Ala Asp Pro Gly Ala Asn Pro His Gln Ala
    180          185          190
    Ser Pro Val Ile Asn Val Ile Ile Pro Glu Gly Ala Gly Tyr Asn Asn
    195          200          205
45 Thr Leu Asp His Gly Leu Cys Thr Ala Phe Glu Glu Ser Glu Leu Gly
    210          215          220
    Asp Asp Val Glu Ala Asn Phe Thr Ala Val Phe Ala Pro Pro Ile Arg
    225          230          235          240
50 Ala Arg Leu Glu Ala His Leu Pro Gly Val Asn Leu Thr Asp Glu Asp
    245          250          255
55

```

## SEQUENCE LISTING.txt

[illegible]

## Claims

1. A fermentation assembly comprising

a vessel suitable for carrying out reactions involving living cells;

at least two storage flasks connected to said vessel for supply of liquids and means to transport said liquids from said storage flasks to said vessel;

individual appliances monitoring the supply of the contents of said storage flasks to said vessel;



a harvest flask connected to said vessel and means to transport fermentation broth from said vessel to said harvest flask; and

a device for controlling and maintaining a constant dilution rate in said vessel with varying rates of individual supply of liquid from said storage flasks to said vessel.

2. An assembly as in claim 1 and in accordance with Figure 1 comprising

a fermentor 1 equipped with inlet tubes 2a from storage flasks 2 for supply of liquids; pumps 3 for transporting liquids from the storage flasks 2 to fermentor 1; scales 4 for monitoring the amount of liquids supplies to and discharged from the fermentor; gas inlet tubes 9 and outlet tubes 10; pump 6 for discharging fermentation broth via outlet tubes 5a to a harvest flask 5; main controlling unit 7 for overall process monitoring and steering; controlling unit 11 for monitoring and steering individual control systems 17 for temperature, pH, gas pressure, fermentor content and antifoam agents; circuit 12 including pump 13 for gas supply and taking samples; gas inlet and outlet flow control 14 and 15; and, optionally, sterile filters 16 and thermostating unit 8.

3. An assembly as in claims 1 or 2, wherein said storage flasks comprise individual flasks for solutions of carbon, nitrogen, and mineral sources required for the growth of said cells and optimal formation of the desired reaction product.

4. An assembly as in any one of claims 1 to 3, wherein said storage flasks comprise at least one individual flask containing a controlling agent.

5. An assembly as in any one of claims 1 to 4, wherein said storage flasks comprise an individual flask containing water.

6. An assembly as in any one of claims 1 to 5, wherein said vessel contains a fixed bed and/or an expanded bed and/or a moving bed of immobilized living cells.

7. An assembly as in claim 6 wherein the living cells are immobilized on a porous carrier.

8. A continuous process for the manufacture of proteins from cultures of living cells in which process the nutrients and other agents required for the growth of the cells and the optimal production of the desired protein are fed into the reactor individually at a constant dilution rate.

9. A continuous process according to claim 8 wherein the protein is selected from the group consisting of catalase, lactase, phenoloxidase, oxidase, oxidoreductase, glucanase cellulase, xylanase and other polysaccharide, peroxidase, lipase, hydrolase, esterase, cutinase, protease and other proteolytic enzymes, aminopeptidase, carboxypeptidase, phytase, lyase, pectinase and other pectinolytic enzymes, amylase, glucosidase, mannosidase, isomerase, invertase, transferase, ribonuclease, chitinase, and desoxyribonuclease or the protein is selected from the group of therapeutic proteins such as antibodies, vaccines, antigens, or of antibacterial and/or health-beneficial proteins such as lactoternin, lactoperoxidase or lysozyme.

10. A continuous process according to claim 8 wherein the protein is selected from the group consisting of proteins having the activity of a therapeutic protein such as antibodies, vaccines, antigens.

11. A process as in any one of claims 8 to 10 wherein the cells are immobilized.

12. A process as in any one of claims 8 to 11 wherein the cell is a phytase-producing microorganism.

13. A process as in claim 12, wherein the phytase-producing microorganism is *Hansenula polymorpha*.

14. A process as in claim 13, wherein the phytase-producing microorganism is *Hansenula polymorpha* transformed by a DNA encoding a phytase of fungal or consensus origin.

15. A process as in any one of claims 8 to 14, wherein the cell or microorganism is in a fixed bed and/or an expanded bed and/or a moving bed on a porous carrier.

**EP 1 092 764 A2**

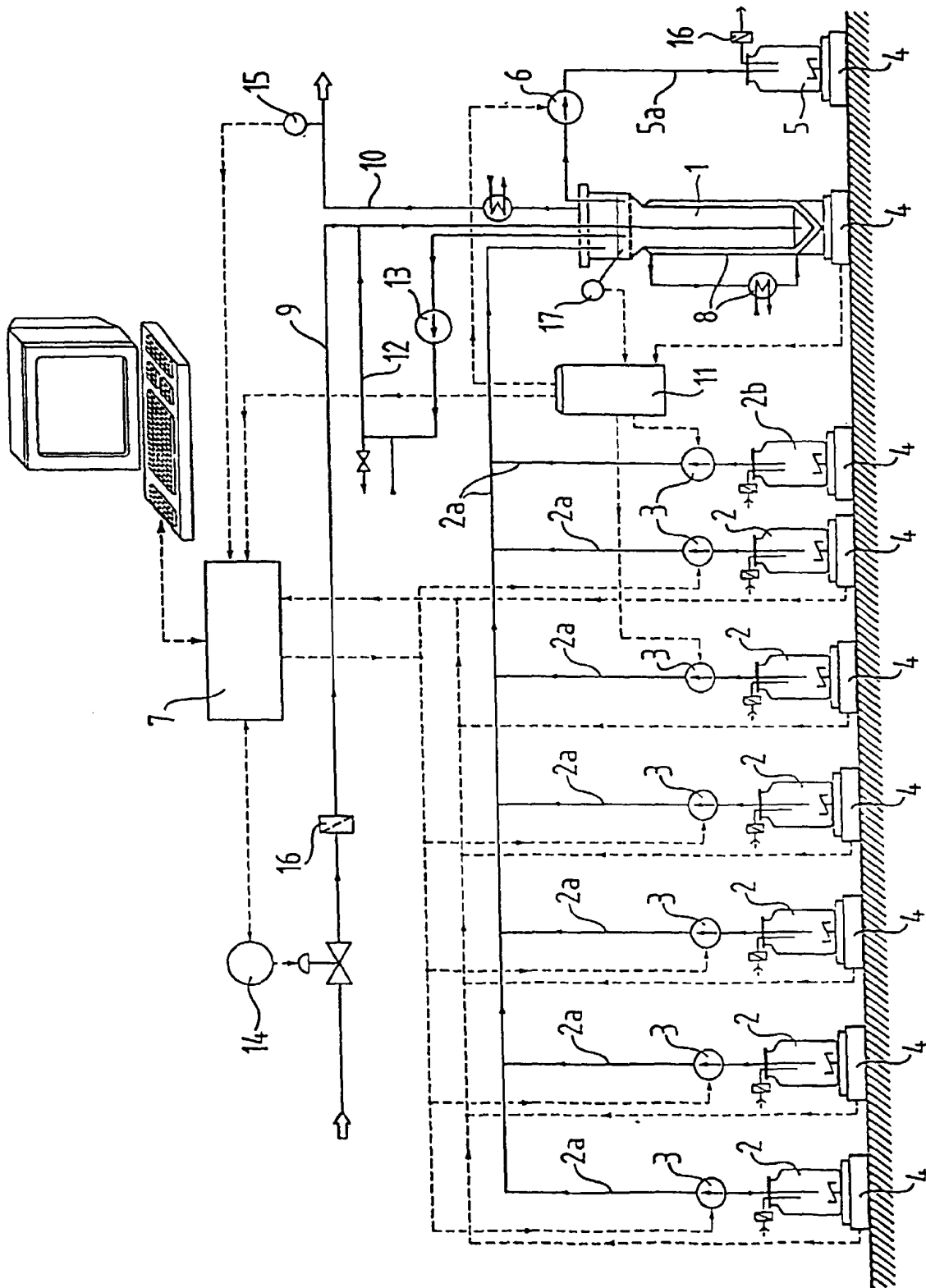
**16.** A process as in any one of claims 8 to 15, wherein the carbon source is glycerol or a sugar like a mono-, di- or polysaccharide.

**17.** A process as in claim 16, wherein the carbon source is glucose.

**18.** A process as in any one of claims 8 to 15, wherein the carbon source is methanol.

**19.** A process as in any one of claims 8 to 15, wherein the carbon source is glucose and methanol.

**20.** A process as in 19, wherein the total amount of methanol and glucose is from about 10 to about 500 g/l each.



**Figure 2**

50	1
<i>A. terreus</i> 9A-1	KhsDCNSVDh GYQCFPELSH kWGLYAPYFS
LQDESPFP1D VPEDChITFV	
<i>A. terreus</i> cbs	NhsDCTSVDr GYQCFPELSH kWGLYAPYFS
LQDESPFP1D VPDDChITFV	
<i>A. niger</i> var. <i>awamori</i>	NqsTCDTVDQ GYQCFSETSH LWGQYAPFFS
LANESAISPD VPAGCrVTFA	
<i>A. niger</i> T213	NqsSCDTVDQ GYQCFSETSH LWGQYAPFFS
LANESVISPD VPAGCrVTFA	
<i>A. niger</i> NRRL3135	NqsSCDTVDQ GYQCFSETSH LWGQYAPFFS
LANESVISPE VPAGCrVTFA	
<i>A. fumigatus</i> 13073	GSkSCDTVD1 GYQCsPATSH LWGQYSPFFS
LEDElSVSSK LPKDCrITLV	
<i>A. fumigatus</i> 32722	GSkSCDTVD1 GYQCsPATSH LWGQYSPFFS
LEDElSVSSK LPKDCrITLV	
<i>A. fumigatus</i> 58128	GSkSCDTVD1 GYQCsPATSH LWGQYSPFFS
LEDElSVSSK LPKDCrITLV	
<i>A. fumigatus</i> 26906	GSkSCDTVD1 GYQCsPATSH LWGQYSPFFS
LEDElSVSSK LPKDCrITLV	
<i>A. fumigatus</i> 32239	GSkACDTVE1 GYQCsPGTSH LWGQYSPFFS
LEDElSVSSD LPKDCrVTFV	
<i>E. nidulans</i>	QNHSCNTADG GYQCFPNVSH VWGQYSPYFS
IEQESAISed VPHGCEvTFV	
<i>T. thermophilus</i>	DSHSCNTVEG GYQCrPEISH sWGQYSPFFS
LADQSEISPD VPQNCKITFV	
<i>M. thermophila</i>	ESRPCDTpD1 GFQCgTAISH FWGQYSPYFS
VpSElDaS.. IPDDCeVTFA	
Consensus	NSHSCDTVDG GYQCFPEISH LWGQYSPYFS
LEDESAISPD VPDDC-VTFV	
<b>Consensus phytase</b>	<b>NSHSCDTVDG GYQCFPEISH LWGQYSPYFS</b>
<b>LEDESAISPD VPDDCrVTFV</b>	
100	51
<i>A. terreus</i> 9A-1	QVLARHGARs PThSKtKAYA AtIAAIQKSA
TaFpGKYAFL QSYNYSLDSE	
<i>A. terreus</i> cbs	QVLARHGARs PTDSKtKAYA AtIAAIQKNA
TaLpGKYAFL KSYNYSMGSE	
<i>A. niger</i> var. <i>awamori</i>	QVLSRHGARY PTESKgKkYS ALIEEIQQNV
TtFDGKYAFL KTYNYSLGAD	
<i>A. niger</i> T213	QVLSRHGARY PTESKgKkYS ALIEEIQQNV
TtFDGKYAFL KTYNYSLGAD	
<i>A. niger</i> NRRL3135	QVLSRHGARY PTDSKgKkYS ALIEEIQQNA
TtFDGKYAFL KTYNYSLGAD	
<i>A. fumigatus</i> 13073	QVLSRHGARY PTSSKsKkYK kLVTAIQaNA
TdFKGKFAFL KTYNYTLGAD	
<i>A. fumigatus</i> 32722	QVLSRHGARY PTSSKsKkYK kLVTAIQaNA
TdFKGKFAFL KTYNYTLGAD	
<i>A. fumigatus</i> 58128	QVLSRHGARY PTSSKsKkYK kLVTAIQaNA
TdFKGKFAFL KTYNYTLGAD	
<i>A. fumigatus</i> 26906	QVLSRHGARY PTSSKsKkYK kLVTAIQaNA
TdFKGKFAFL KTYNYTLGAD	
<i>A. fumigatus</i> 32239	QVLSRHGARY PTASKsKkYK kLVTAIQKNA
TeFKGKFAFL ETYNYTLGAD	

<i>E. nidulans</i>	QVLSRHGARY PTESKsKAYS GLIEAIQKNA
TsFwGQYAFL ESYNYTLGAD	
<i>T. thermophilus</i>	QLLSRHGARY PTSSKtELYS QLISrIQKTA
TaYKGyYAFL KDYrYqLGAN	
<i>M. thermophila</i>	QVLSRHGARa PtlKRaaSYv DLIDrIHhGA
IsYgPgYEFL RTYDYLTLGAD	
Consensus	QVLSRHGARY PTSSK-KAYS ALIEAIQKNA T-
FKGKYAFL KTYNYTLGAD	
<b>Consensus phytase</b>	<b>QVLSRHGARY PTSSKSKAYS ALIEAIQKNA</b>
<b>TAFKGKYAFL KTYNYTLGAD</b>	

101

150	
<i>A. terreus</i> 9A-1	ELTPFGrNQL rDlGaQFYeR YNALTRhInP
FVRATDASRV hESAekFVEG	
<i>A. terreus</i> cbs	NLTPFGrNQL qDlGaQFYRR YDTLTRhInP
FVRAADSSRV hESAekFVEG	
<i>A. niger</i> var. <i>awamori</i>	DLTPFGEQEL VNSGIKFYQR YESLTRNIIP
FIRSSGSSRV IASGEKFIEG	
<i>A. niger</i> T213	DLTPFGEQEL VNSGIKFYQR YESLTRNIIP
FIRSSGSSRV IASGEKFIEG	
<i>A. niger</i> NRRL3135	DLTPFGEQEL VNSGIKFYQR YESLTRNIVP
FIRSSGSSRV IASGKKFIEG	
<i>A. fumigatus</i> 13073	DLTPFGEQQL VNSGIKFYQR YKALARSVVP
FIRASGSDRV IASGEKFIEG	
<i>A. fumigatus</i> 32722	DLTPFGEQQL VNSGIKFYQR YKALARSVVP
FIRASGSDRV IASGEKFIEG	
<i>A. fumigatus</i> 58128	DLTPFGEQQL VNSGIKFYQR YKALARSVVP
FIRASGSDRV IASGEKFIEG	
<i>A. fumigatus</i> 26906	DLTAFGEQQL VNSGIKFYQR YKALARSVVP
FIRASGSDRV IASGEKFIEG	
<i>A. fumigatus</i> 32239	DLTPFGEQQM VNSGIKFYQK YKALAgSVVP
FIRSSGSDRV IASGEKFIEG	
<i>E. nidulans</i>	DLTiFGENQM VD SGaKFYRR YKNLARKnTP
FIRASGSDRV VASAekFING	
<i>T. thermophilus</i>	DLTPFGENQM IqlGIKFYnH YKSLARNaVP
FVRCSGSDRV IASGrIFIEG	
<i>M. thermophila</i>	ELTRtGQQQM VNSGIKFYRR YRALARKsIP
FVRTAGqDRV VhSAENFTQG	
Consensus	DLTPFGENQM VNSGIKFYRR YKALARK-VP
FVRASGSDRV IASAEKFIEG	
<b>Consensus phytase</b>	<b>DLTPFGENQM VNSGIKFYRR YKALARKIVP</b>
<b>FIRASGSDRV IASAEKFIEG</b>	

200

151

*A. terreus* 9A-1 FQTARqDDHh ANpHQPSPrV DVaIPEGSAY  
 NNTLEHSIcT AFES...STV  
*A. terreus* cbs FQNARqGDPh ANpHQPSPrV DVVIPEGTAY  
 NNTLEHSIcT AFEA...STV  
*A. niger* var. *awamori* FQSTKLkDPr AqpgQSSPki DVVISEASSs  
 NNTLDPGTCT VFED...SEL  
*A. niger* T213 FQSTKLkDPr AqpgQSSPki DVVISEASSs  
 NNTLDPGTCT VFED...SEL  
*A. niger* NRRL3135 FQSTKLkDPr AqpgQSSPki DVVISEASSs  
 NNTLDPGTCT VFED...SEL  
*A. fumigatus* 13073 FQqAKLADPG A.TNRAAPAI SVIIPeSETF  
 NNTLDHGVCT kFEA...SQL  
*A. fumigatus* 32722 FQqAKLADPG A.TNRAAPAI SVIIPeSETF  
 NNTLDHGVCT kFEA...SQL  
*A. fumigatus* 58128 FQqAKLADPG A.TNRAAPAI SVIIPeSETF  
 NNTLDHGVCT kFEA...SQL  
*A. fumigatus* 26906 FQqAKLADPG A.TNRAAPAI SVIIPeSETF  
 NNTLDHGVCT kFEA...SQL  
*A. fumigatus* 32239 FQqANVADPG A.TNRAAPVI SVIIPeSETY  
 NNTLDHSVCT NFEA...SEL  
*E. nidulans* FRKAQLhDHG S..gQATPVV NVIIPeIdGF  
 NNTLDHSTCV SFEN...DER  
*T. thermophilus* FQSAKvIdPh SDkHDAPPTI NVIIEEGPSY  
 NNTLDtGSCP VFED...SSg  
*M. thermophila* FHSAlLADRG STvRPTlPyd mVVIPETAGa  
 NNTLHNDlCT AFEEgpySTI

Consensus FQSAKLADPG S-PHQASpVI NVIIPeGSGY  
 NNTLDHGTCT AFED---SEL  
 Consensus phytase FQSAKLADPG SQPHQASpVI DVIIPeGSGY  
 NNTLDHGTCT AFED...SEL

250

201

*A. terreus* 9A-1 GDDAvANFTA VFAPAIaQRL EADLPgVqLS  
 TDDVvNLmAM CPfETVSlTD  
*A. terreus* cbs GDAAADNFTA VFAPAIakRL EADLPgVqLS  
 ADDVvNLmAM CPfETVSlTD  
*A. niger* var. *awamori* ADTVEANFTA TFAPsIRQRL ENDLsgVtLT  
 DTEVTyLMDM CSFDtISTSt  
*A. niger* T213 ADTVEANFTA TFAPsIRQRL ENDLsgVtLT  
 DTEVTyLMDM CSFDtISTSt  
*A. niger* NRRL3135 ADTVEANFTA TFVPSIRQRL ENDLsgVtLT  
 DTEVTyLMDM CSFDtISTSt  
*A. fumigatus* 13073 GDEVAANFTA lFAPDIRARa EkHLPgVtLT  
 DEDVVsLMDM CSFDtVARTS  
*A. fumigatus* 32722 GDEVAANFTA lFAPDIRARa EkHLPgVtLT  
 DEDVVsLMDM CSFDtVARTS  
*A. fumigatus* 58128 GDEVAANFTA lFAPDIRARa EkHLPgVtLT  
 DEDVVsLMDM CSFDtVARTS  
*A. fumigatus* 26906 GDEVAANFTA lFAPDIRARa KkHLPgVtLT  
 DEDVVsLMDM CSFDtVARTS  
*A. fumigatus* 32239 GDEVEANFTA lFAPAIRARI EkHLPgVqLT  
 DDDVVsLMDM CSFDtVARTa  
*E. nidulans* ADEiEANFTA IMGPPIrkRL ENDLPgIKLT  
 NENViYlMDM CSFDtMARTa  
*T. thermophilus* GHDAQEKfAk qFAPAIleKI KDHLPGVDLA  
 vSDVpyLMDL CPfETLARNh  
*M. thermophila* GDDAQDTYlS TFAGPItARV NANLPgANLT  
 DADTVaLMDL CPfETVAsSS

Consensus GDDAEANFTA TFAPAIRARL EADLPGVTLT DEDVV-  
 LMDM CPFETVARTS  
 Consensus phytase GDDVEANFTA LFAPAIRARL EADLPGVTLT  
 DEDVVYLMDM CPFETVARTS

251

300

*A. terreus* 9A-1 ..... DAhTLSPFC DLFTAtEWtq  
 YNYLlSLDKY YGYGGGNPLG  
*A. terreus* cbs ..... DAhTLSPFC DLFTAaEWtq  
 YNYLlSLDKY YGYGGGNPLG  
*A. niger* var. *awamori* ..... vDTKLSPFC DLFTHdEWih  
 YDYLQSLkKY YGHGAGNPLG  
*A. niger* T213 ..... vDTKLSPFC DLFTHdEWih  
 YDYLRSLkKY YGHGAGNPLG  
*A. niger* NRRL3135 ..... vDTKLSPFC DLFTHdEWin  
 YDYLQSLkKY YGHGAGNPLG  
*A. fumigatus* 13073 ..... DASQLSPFC QLFTHnEWkk  
 YNYLQSLGKY YGYGAGNPLG  
*A. fumigatus* 32722 ..... DASQLSPFC QLFTHnEWkk  
 YNYLQSLGKY YGYGAGNPLG  
*A. fumigatus* 58128 ..... DASQLSPFC QLFTHnEWkk  
 YNYLQSLGKY YGYGAGNPLG  
*A. fumigatus* 26906 ..... DASQLSPFC QLFTHnEWkk  
 YNYLQSLGKY YGYGAGNPLG  
*A. fumigatus* 32239 ..... DASELSPFC AIFTHnEWkk  
 YDYLQSLGKY YGYGAGNPLG  
*E. nidulans* ..... HGTELSPFC AIFTEkEWlq  
 YDYLQSLSKY YGYGAGSPLG  
*T. thermophilus* ..... TDT.LSPFC ALsTQeEWqa  
 YDYYQSLGKY YGnGGGNPLG  
*M. thermophila* sdpatadagg gNGrplSPFC rLFSEsEWra  
 YDYLQSVGKW YGYGPGNPLG

Consensus ----- -DATELSPFC ALFTE-EW--  
 YDYLQSLGKY YGYGAGNPLG  
 Consensus phytase ..... .DATELSPFC ALFTHDEWRQ  
 YDYLQSLGKY YGYGAGNPLG

350  
*A. terreus* 9A-1 PVQGVGWaNE LMARLTRAPV HDHTCVNNTL  
 DASPATFPLN ATLYADFSHD  
*A. terreus* cbs PVQGVGWaNE LIARLTRSPV HDHTCVNNTL  
 DANPATFPLN ATLYADFSHD  
*A. niger* var. *awamori* PTQGVGYaNE LIARLTHSPV HDDTSSNHTL  
 DSNPATFPLN STLYADFSHD  
*A. niger* T213 PTQGVGYaNE LIARLTHSPV HDDTSSNHTL  
 DSNPATFPLN STLYADFSHD  
*A. niger* NRRL3135 PTQGVGYaNE LIARLTHSPV HDDTSSNHTL  
 DSSPATFPLN STLYADFSHD  
*A. fumigatus* 13073 PAQGIGFtNE LIARLTRSPV QDHTSTNsTL  
 vSNPATFPLN ATMYVDFSHD  
*A. fumigatus* 32722 PAQGIGFtNE LIARLTRSPV QDHTSTNsTL  
 vSNPATFPLN ATMYVDFSHD  
*A. fumigatus* 58128 PAQGIGFtNE LIARLTRSPV QDHTSTNsTL  
 vSNPATFPLN ATMYVDFSHD  
*A. fumigatus* 26906 PAQGIGFtNE LIARLTRSPV QDHTSTNsTL  
 vSNPATFPLN ATMYVDFSHD  
*A. fumigatus* 32239 PAQGIGFtNE LIARLTNSPV QDHTSTNsTL  
 DSDPATFPLN ATYVDFSHD  
*E. nidulans* PAQGIGFtNE LIARLTQSPV QDNTSTNHTL  
 DSNPATFPLD rKLYADFSHD  
*T. thermophilus* PAQGVGFvNE LIARMTSPV QDYTTVNHTL  
 DSNPATFPLN ATLYADFSHD  
*M. thermophila* PTQGVGFvNE LLARLAGvPV RDgTSTNRTL  
 DGDPrTFPLG rPLYADFSHD  
 Consensus PAQGVGF-NE LIARLTHSPV QDHTSTNHTL  
 DSNPATFPLN ATLYADFSHD  
 Consensus phytase PAQGVGFANE LIARLTRSPV QDHTSTNHTL  
 DSNPATFPLN ATLYADFSHD

351  
 400  
*A. terreus* 9A-1 SNLVSIFWAL GLYNGTAPLS qTSVESVSQT  
 DGYAAAWTVP FAARAYVEMM  
*A. terreus* cbs SNLVSIFWAL GLYNGTkPLS qTTVEDITrT  
 DGYAAAWTVP FAARAYIEMM  
*A. niger* var. *awamori* NGIISILFAL GLYNGTkPLS TTTVENITQT  
 DGFSSAWTVP FASrLYVEMM  
*A. niger* T213 NGIISILFAL GLYNGTkPLS TTTVENITQT  
 DGFSSAWTVP FASrLYVEMM  
*A. niger* NRRL3135 NGIISILFAL GLYNGTkPLS TTTVENITQT  
 DGFSSAWTVP FASrLYVEMM  
*A. fumigatus* 13073 NSMVSIFFAL GLYNGTEPLS rTSVESaKEl  
 DGYSASWVVP FGARAYFetM  
*A. fumigatus* 32722 NSMVSIFFAL GLYNGTGPLS rTSVESaKEl  
 DGYSASWVVP FGARAYFetM  
*A. fumigatus* 58128 NSMVSIFFAL GLYNGTEPLS rTSVESaKEl  
 DGYSASWVVP FGARAYFetM  
*A. fumigatus* 26906 NSMVSIFFAL GLYNGTEPLS rTSVESaKEl  
 DGYSASWVVP FGARAYFetM  
*A. fumigatus* 32239 NGMIPiFFAM GLYNGTEPLS qTSeESTKES  
 NGYSASWAVP FGARAYFetM  
*E. nidulans* NSMISiFFAM GLYNGTQPLS mDSVESIQEm  
 DGYAASWTVP FGARAYFELM  
*T. thermophilus* NTMTSiFaAL GLYNGTAKLS TTEIKSiEET  
 DGYSAAWTVP FGGRAYIEMM  
*M. thermophila* NDMMGVLgAL GaYDGVpPLD KTArrDpEEl  
 GGYAASWAVP FAARiYVEKM



Consensus NSMISIFFAL GLYNGTAPLS TTSVESIEET  
 DGYAASWTVP FGARAYVEMM  
 Consensus phytase NSMISIFFAL GLYNGTAPLS TTSVESIEET  
 DGYSASWTVP FGARAYVEMM

401

450  
*A. terreus* 9A-1 QC..... RAEKE PLVRVLVNDR  
 VMPLHGCTPD KLGRCKrDAF  
*A. terreus* cbs QC..... RAEKQ PLVRVLVNDR  
 VMPLHGCAVD NLGRCKrDDF  
*A. niger* var. *awamori* QC..... QAEQE PLVRVLVNDR  
 VVPLHGCPID aLGRCTrDSF  
*A. niger* T213 QC..... QAEQE PLVRVLVNDR  
 VVPLHGCPID aLGRCTrDSF  
*A. niger* NRRL3135 QC..... QAEQE PLVRVLVNDR  
 VVPLHGCPVD aLGRCTrDSF  
*A. fumigatus* 13073 QC..... KSEKE PLVRALINDR  
 VVPLHGCDVD KLGRCKLNDF  
*A. fumigatus* 32722 QC..... KSEKE PLVRALINDR  
 VVPLHGCDVD KLGRCKLNDF  
*A. fumigatus* 58128 QC..... KSEKE SLVRALINDR  
 VVPLHGCDVD KLGRCKLNDF  
*A. fumigatus* 26906 QC..... KSEKE PLVRALINDR  
 VVPLHGCDVD KLGRCKLNDF  
*A. fumigatus* 32239 QC..... KSEKE PLVRALINDR  
 VVPLHGCAVD KLGRCKLKDF  
*E. nidulans* QC..... E.KKE PLVRVLVNDR  
 VVPLHGCAVD KFGRCTLDDW  
*T. thermophilus* QC..... DDSDE PVVRVLVNDR  
 VVPLHGCEVD SLGRCKrDDF  
*M. thermophila* RCsggggggg ggegrQEKDE eMVRVLVNDR  
 VMTLkGCGAD ErGMCTLErF  
 Consensus QC----- ----QAEKE PLVRVLVNDR  
 VVPLHGCAVD KLGRCKLDDF  
 Consensus phytase QC..... QAEKE PLVRVLVNDR  
 VVPLHGCAVD KLGRCKRDDF

451

471  
*A. terreus* 9A-1 VAGLSFAQAG GNWADCF~~~ ~  
*A. terreus* cbs VEGLSFARAG  
 GNWAECF~~~ ~  
*A. niger* var. *awamori* VrGLSFARSG GDWAECsA~~ ~  
*A. niger* T213 VrGLSFARSG GDWAECFA~~ ~  
*A. niger* NRRL3135 VrGLSFARSG  
 GDWAECFA~~ ~  
*A. fumigatus* 13073 VKGLSWARSG GNWGECSFS~~ ~  
*A. fumigatus* 32722 VKGLSWARSG GNWGECSFS~~ ~  
*A. fumigatus* 58128 VKGLSWARSG GNWGECSFS~~ ~  
*A. fumigatus* 26906 VKGLSWARSG GNWGECSFS~~ ~  
*A. fumigatus* 32239 VKGLSWARSG  
 GNSEQSFS~~ ~  
*E. nidulans* VEGLNFARSG GNWkTCFTl~ ~  
*T. thermophilus* VrGLSFARqG GNWEGCYAas e  
*M. thermophila* IESMAFARGN GKWDlCFA~~ ~  
 Consensus VEGLSFARSG GNWAECFA-- ~  
 Consensus phytase VEGLSFARSG GNWAECFA.. .

Figure 3

CP-1

Eco RI M G V F V V L L S I A T L F G S T  
**TATATGAATTCATGGGCGTGTTTCGTGCTACTGTCCATTGCCACCTTGTTTCGGTTCCA**

1 -----+-----+-----+-----+-----+-----+ 60

ATATACTTAAGTACCCGCACAAGCAGCACGATGACAGGTAACGGTGGAACAAGCCAAGGT

S G T A L G P R G N S H S C D T V D G G  
**CATCCGGTACCGCCTTGGGTCCTCGTGGTAATTCTCACTCTTGTGACACTGTTGACGGTG**

61 -----+-----+-----+-----+-----+-----+ 120

GTAGGCCATGGCGGAACCCAGGAGCACCATTAAAGAGTGAGAACACTGTGACAACTGCCAC

CP-2

CP-3

Y Q C F P E I S H L W G Q Y S P Y F S L  
**GTTACCAATGTTTCCCAGAAATTTCTCACTTGTGGGGTCAATACTCTCCATACTTCTCTT**

121 -----+-----+-----+-----+-----+-----+ 180

**CAATGGTTACAAAGGCTCTTTAAAGAGTGAACACCCCAGTTATGAGAGGTATGAAGAGAA**

E D E S A I S P D V P D D C R V T F V Q  
**TGGAAGACGAATCTGCTATTTCTCCAGACGTTCCAGACGACTGTAGAGTTACTTTTCGTTT**

181 -----+-----+-----+-----+-----+-----+ 240

ACCTTCTGCTTAGACGATAAAGAGGTCTGCAAGGTCTGCTGACATCTCAATGAAAGCAAG

CP-4

CP-5

V L S R H G A R Y P T S S K S K A Y S A  
**AAGTTTTGTCTAGACACGGTGCTAGATACCCAACCTTCTTCTAAGTCTAAGGCTTACTCTG**

241 -----+-----+-----+-----+-----+-----+ 300

**TTCAAAACAGATCTGTGCCACGATCTATGGGTTGAAGAAGATTCAGATTC CGAATGAGAC**

**L I E A I Q K N A T A F K G K Y A F L K**

**CTTTGATTGAAGCTATTCAAAGAACGCTACTGCTTTCAAGGGTAAGTACGCTTTCTTGA**

301 -----+-----+-----+-----+-----+-----+ 360

**GAAACTAACTTCGATAAGTTTTCTTGCGATGACGAAAGTTCCCATTCATGCGAAAGAACT**

CP-6

CP-7

**T Y N Y T L G A D D L T P F G E N Q M V**

**AGACTTACAAC TACACTTTGGGTGCTGACGACTTGACTCCATTCGGTGAAAACCAAATGG**

361 -----+-----+-----+-----+-----+-----+ 420

**TCTGAATGTTGATGTGAAACCCACGACTGCTGAACTGAGGTAAGCCACTTTTGGTTTACC**

**N S G I K F Y R R Y K A L A R K I V P F**

**TTAACTCTGGTATTAAGTTCTACAGAAGATACAAGGCTTTGGCTAGAAAGATTGTTCCAT**

421 -----+-----+-----+-----+-----+-----+ 480

**AATTGAGACCATAATTCAAGATGTCTTCTATGTTCCGAAACCGATCTTTCTAACAAGGTA**

CP-8

CP-9

**I R A S G S D R V I A S A E K F I E G F**

**TCATTAGAGCTTCTGGTTCTGACAGAGTTATTGCTTCTGCTGAAAAGTTCATTGAAGGTT**

481 -----+-----+-----+-----+-----+-----+ 540

**AGTAATCTCGAAGACCAAGACTGTCTCAATAACGAAGACGACTTTTCAAGTAACCTCCAA**

Q S A K L A D P G S Q P H Q A S P V I D  
TCCAATCTGCTAAGTTGGCTGACCCAGGTTCTCAACCACACCAAGCTTCTCCAGTTATTG

541 -----+-----+-----+-----+-----+-----+ 600

AGGTTAGACGATTCAACCGACTGGGTCCAAGAGTTGGTGTGGTTCGAAGAGGTCAATAAC

CP-10

CP-11

V I I P E G S G Y N N T L D H G T C T A  
ACGTTATTATTCCAGAAGGATCcGGTTACAACAACACTTTGGACCACGGTACTTGTTACTG

601 -----+-----+-----+-----+-----+-----+ 660

TGCAATAATAAGGTCTTCCTAGgCCAATGTTGTTGTGAAACCTGGTGCCATGAACATGAC

F E D S E L G D D V E A N F T A L F A P  
CTTTCGAAGACTCTGAATTGGGTGACGACGTTGAAGCTAACTTCACTGCTTTGTTGCTC

661 -----+-----+-----+-----+-----+-----+ 720

GAAAGCTTCTGAGACTTAACCCACTGCTGCAACTTCGATTGAAGTGACGAAACAAGCGAG

CP-12

A I R A R L E A D L P G V T L T D E D V  
CAGCTATTAGAGCTAGATTGGAAGCTGACTTGCCAGGTGTTACTTTGACTGACGAAGACG

721 -----+-----+-----+-----+-----+-----+ 780

GTCGATAATCTCGATCTAACCTTCGACTGAACGGTCCACAATGAACTGACTGCTTCTGC

CP-13

V Y L M D M C P F E T V A R T S D A T E  
TTGTTTACTTGATGGACATGTGTCCATTGGAAGCTGTTGCTAGAACTTCTGACGCTACTG

781 -----+-----+-----+-----+-----+-----+ 840

**AACAAATGAACTACCTGTACACAGGTAAGCTTTGACAACGATCTTGAAGACTGCGATGAC**

**L S P F C A L F T H D E W R Q Y D Y . L Q**

**AATTGTCTCCATTCTGTGCTTTGTTCACTCACGACGAATGGAGACAATACGACTACTTGC**

841 -----+-----+-----+-----+-----+-----+ 900

**TTAACAGAGGTAAGACACGAAACAAGTGAGTGCTGCTTACCTCTGTTATGCTGATGAACG**

CP-14

CP-15

**S L G K Y Y G Y G A G N P L G P A Q G V**

**AATCTTTGGGTAAGTACTACGGTTACGGTGCTGGTAACCCATTGGGTCCAGCTCAAGGTG**

901 -----+-----+-----+-----+-----+-----+ 960

**TTAGAAACCCATTTCATGATGCCAATGCCACGACCATTGGGTAACCCAGGTGCGAGTTCCAC .**

**G F A N E L I A R L T R S P V Q D H T S**

**TTGGTTTCGCTAACGAATTGATTGCTAGATTGACTAGATCTCCAGTTCAAGACCACACTT**

961 -----+-----+-----+-----+-----+-----+ 1020

**AACCAAAGCGATTGCTTAACTAACGATCTAACTGATCTAGAGGTCAAGTTCTGGTGTGAA**

CP-16

CP-17

**T N H T L D S N P A T F P L N A T L Y A**

**CTACTAACCCACACTTTGGACTCTAACCCAGCTACTTTCCCATTTGAACGCTACTTTGTACG**

1021 -----+-----+-----+-----+-----+-----+ 1080

**GATGATTGGTGTGAAACCTGAGATTGGGTGCGATGAAAGGGTAACTTGCGATGAAACATGC**

D F S H D N S M I S I F F A L G L Y N G

CTGACTTCTCTCAGACAACCTCTATGATTTCTATTTTCTTCGCTTTGGGTTTGTACAACG

1081 -----+-----+-----+-----+-----+-----+

1140

GACTGAAGAGAGTGCTGTTGAGATACTAAAGATAAAAAGAAGCGAAACCCAAACATGTTGC

CP-18

CP-19

T A P L S T T S V E S I E E T D G Y S A

GTACTGCTCCATTGTCTACTACTTCTGTTGAATCTATTGAAGAACTGACGGTTACTCTG

1141 -----+-----+-----+-----+-----+-----+

1200

CATGACGAGGTAACAGATGATGAAGACAACCTTAGATAACTTCTTTGACTGCCAATGAGAC

S W T V P F G A R A Y V E M M Q C Q A E

CTTCTTGGAAGTGTTCATTTCGGTGCTAGAGCTTACGTTGAAATGATGCAATGTCAAGCTG

1201 -----+-----+-----+-----+-----+-----+

1260

GAAGAACCTGACAAGGTAAGCCACGATCTCGAATGCAACTTTACTACGTTACAGTTTCGAC

CP-20

CP-21

K E P L V R V L V N D R V V P L H G C A

AAAAGGAACCATTTGGTTAGAGTTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTG

1261 -----+-----+-----+-----+-----+-----+

1320

TTTTCCTTGGTAACCAATCTCAAAACCAATTGCTGTCTCAACAAGGTAACGTGCCAACAC

V D K L G R C K R D D F V E G L S F A R

CTGTTGACAAGTTGGGTAGATGTAAGAGAGACGACTTCGTTGAAGGTTTGTCTTTCGCTA

1321 -----+-----+-----+-----+-----+-----+

1380

GACAACTGTTCAACCCATCTACATTCTCTCTGCTGAAGCAACTTCCAAACAGAAAGCGAT

CP-22

S G G N W A E C F A \* Eco RI

GATCTGGTGGTAACTGGGCTGAATGTTTCGCTTAAGAATTCATATA

1381 -----+-----+-----+-----+----- 1426

CTAGACCACCATTGACCCGACTTACAAAGCGAATTCTTAAGTATAT

Figure 4

1

50

*P. involutus* (phyA1) SvP.KnTAPt FPIPeseQrn WSPYSPYFPL AeYkAPPAGC  
QInQVNIQR

*P. involutus* (phyA2) SvP.RniAPK FSIPeseQrn WSPYSPYFPL AeYkAPPAGC  
EInQVNIQR

*T. pubescens* hiPlRdTSAc LdVTrDvQqs WSmYSPYFPa AtYvAPPASC  
QInQVHIIQR

*A. pediades* GgvvQaTfvQ pfFFPpQiQds WAAYTPYYPV qaYtPPPkDC  
KItQVNIQR

*P. lycii* StQfsfvAAQ LPIPaQntsn WGPYdPFFPV EpYaAPPEGC  
tVtQVNIQR

**Basidio** S-P-R-TAAQ LPIP-Q-Q-- WSPYSPYFPV A-Y-APPAGC QI-  
QVNIIQR

51

100

*P. involutus* (phyA1) HGARFPTSGA TTRIKAGLTK LQGvqnftDA KFNFIkSfky  
dLGnsDLVPF

*P. involutus* (phyA2) HGARFPTSGA ATRIKAGLSK LQSvqnftDP KFDFIkSfTY  
dLGtsDLVPF

*T. pubescens* HGARFPTSGA AkRIQTAVAK LKAAsnyTDP lLAFVtNyTY  
sLGqDsLVeL

*A. pediades* HGARFPTSGA GTRIQA AVkK LQSAktyTDP RLDFLtNyTY  
tLGhDDLVPF

*P. lycii* HGARWPTSGA rSRqvAAVAK IQmArpfTDP KYEFLnDfvY  
kFGvADLLPF

**Basidio** HGARFPTSGA ATRIQA AVAK LQSA---TDP KLDFL-N-TY -LG-  
DDLVPF

101

150

*P. involutus* (phyA1) GAaQSfdAGQ EAFARYSkLV SKNNL PFIRA dGSDRVVDSA  
TNWTAGFAsA

*P. involutus* (phyA2) GAaQSfdAGl EvFARYSkLV SsDNLPFIRS dGSDRVVDTA  
TNWTAGFAsA

*T. pubescens* GAtQSSEAGQ EAFTRYSSLV SaDEL PFVRA SGSDRVVATA  
nNWTAGFA1A



*A. pediades* GALQSSQAGE ETFqRYSfLV SkENLPFVRA SSSNRVVDSA  
TNWTEGFSaA

*P. lycii* GAnQShQTGt DmYTRYStLf egGDVPFVRA AGdQRVVDSS  
TNWTAGFGdA

**Basidio** GA-QSSQAGQ EAFTRYs-LV S-DNLPPFVRA SGSDRVVDSA  
**TNWTAGFA-A**

151

200

*P. involutus* (phyA1) ShNTvqPkLn LILPQtGNDT LEDNMCPaAG DSDPQvNaWL  
AVafPSITAR

*P. involutus* (phyA2) SrNAiqPkLd LILPQtGNDT LEDNMCPaAG ESDPQvDaWL  
AsafPSVTAQ

*T. pubescens* SsNSitPvLs VIISEaGNDT LDDNMCPaAG DSDPQvNqWL  
AqFAPPMTAR

*A. pediades* ShHvlnPiLf VILSEslNDT LDDaMCPnAG sSDPQtGiWt  
SIYGTPIAnR

*P. lycii* SgETvlPtLq VVLqEeGNcT LcNNMCPnEv DGDest.tWL  
GVFAPnITAR

**Basidio** S-NT--P-L- VILSE-GNDT LDDNMCP-AG DSDPQ-N-WL  
**AVFAPPITAR**

201

250

*P. involutus* (phyA1) LNAAAPSVNL TDtDAfNLvs LCAFlTVSke kkSdFcTLFE  
giPGsFeAFa

*P. involutus* (phyA2) LNAAAPGANL TDaDAfNLvs LCPFmTVSke qkSdFcLFE  
giPGsFeAFa

*T. pubescens* LNAGAPGANL TDtDTyNLlt LCPFETVatE rrSeFCDIYE  
elQAE.dAFa

*A. pediades* LNqqAPGANI TAaDvsNLip LCAFETIvKE tpSpFCNLF.  
.tPEEFaqFe

*P. lycii* LNAAAPSANL SDsDAItLmd MCPFDTLsG naSpFCDLF.  
.tAEEYvSYe

**Basidio** LNAAAPGANL TD-DA-NL-- LCPFETVS-E --S-FCDLFE --PEEF-  
**AF-**

251

300

*P. involutus* (phyA1) YgGDLDKfYG TGYGQeLGPV QGVGYVNELI ARLTnsAVRD  
NTQTNRTLDA

*P. involutus* (phyA2) YaGDLDKfYG TGYGQALGPV QGVGYINELL ARLTnsAVnD  
NTQTNRTLDA

*T. pubescens* YnADLDKfYG TGYGQPLGPV QGVGYINELI ARLTaQnVsD  
HTQTNsTLDS

*A. pediades* YfGDLDKfYG TGYGQPLGPV QGVGYINELL ARLTemPVRD  
NTQTNRTLDS

*P. lycii* YyyDLdkYyG TGpGNALGPV QGVGYVNELL ARLTgQAVRD  
ETQTNRTLDS

**Basidio** Y-GDLDKfYG TGYGQPLGPV QGVGYINELL ARLT-QAVRD  
**NTQTNRTLDS**

301

350

*P. involutus* (phyA1) SPvTFPLNKT FYADFSHDNl MVAVFSAMGL FrQPAPLsTS  
vPNPwRTWrT

*P. involutus* (phyA2) APdTFPLNKT MYADFSHDNl MVAVFSAMGL FrQSAPLsTS  
tPDPNRTWLT

*T. pubescens* SPeTFPLNRT LYADFSHDNQ MVAIFSAMGL FNQSAPLDPT  
tPDPaRTFLV

*A. pediades* SP1TFPLDRS IYADLSHDNQ MIAIFSAMGL FNQSSPLDPS  
fPNPKRTWVT

*P. lycii* dPaTFPLNRT FYADFSHDNt MVPIFAALGL FNAtA.LDP1  
kPDENRlWVd

**Basidio** SP-TFPLNRT FYADFSHDNQ MVAIFSAMGL FNQSAPLDPS -  
**PDPNRTWVT**

351

400

*P. involutus* (phyA1) SsLVPFSGRM VVERLsC..f GT.....tkV  
RVLVQDqVQP

*P. involutus* (phyA2) SsVVPFSARM aVERLsC..a GT.....tkV  
RVLVQDqVQP

*T. pubescens* kKIVPFSARM VVERLdC..g GA.....qsV  
RLLVNDVQVP

*A. pediades* SRLtPFSARM VtERLlCqrd GTgsggpsri mrngnvqtfV  
RILVNDALQP

*P. lycii* SKLVPFSGHM tVEKLaC...sgkeaV  
RVLVNDVQVP

Basidio SKLVPPFSARM VVERL-C--- GT-----V  
 RVLVNDAVQP

401

441

*P. involutus* (phyA1) LEFCGGDrNG lCTLAKFVES QtFARsDGaG DFEKCFATSa ~

*P. involutus* (phyA2) LEFCGGDqDG lCALDkFVES QaYARsGGaG DFEKCLATTv ~

*T. pubescens* LAFCGADtsG vCTLDaFVES QaYARNDGEG DFEKCFAT~~ ~

*A. pediades* LKFCGGDmDS lCTLEAFVES QkYAREdGQG DFEKCFD~~~ ~

*P. lycii* LEFCGG.vDG vCeLsAFVES QtYARENGQG DfAKCgfvPs e

Basidio LEFCGGD-DG -CTLDaFVES Q-YAREdGQG DFEKCFATP- -

Figure 5

1

50

<i>A. terreus</i> 9a1	KhSDCNSVDh GYQCfPELSH kWGLYAPYFS LqDESPFP1D
VPeDCHITFV	
<i>A. terreus</i> cbs	NhsdCtSVDr GYQCfPELSH kWGLYAPYFS LqDESPFP1D
VPdDCHITFV	
<i>A. niger</i> var. <i>awamori</i>	NqsTCDTVDq GYQCfSEtSH LWGQYAPFFS LANESAISPD
VPaGCRVTFa	
<i>A. niger</i> NRRL3135	NqsSCDTVDq GYQCfSEtSH LWGQYAPFFS LANESvISPE
VPaGCRVTFa	
<i>A. fumigatus</i> 13073	GskSCDTVD1 GYQCSPAtSH LWGQYSPFFS LEDE1SVSSK
LPkDCRITLV	
<i>A. fumigatus</i> 32722	GskSCDTVD1 GYQCSPAtSH LWGQYSPFFS LEDE1SVSSK
LPkDCRITLV	
<i>A. fumigatus</i> 58128	GskSCDTVD1 GYQCSPAtSH LWGQYSPFFS LEDE1SVSSK
LPkDCRITLV	
<i>A. fumigatus</i> 26906	GskSCDTVD1 GYQCSPAtSH LWGQYSPFFS LEDE1SVSSK
LPkDCRITLV	
<i>A. fumigatus</i> 32239	GskACDTVE1 GYQCSPGtSH LWGQYSPFFS LEDE1SVSSD
LPkDCRVTFV	
<i>E. nidulans</i>	QNHSCNTaDG GYQCfPNVSH VWGQYSPYFS IEQESAISeD
VPhGceVTFV	
<i>T. thermophilus</i>	DSHSCNTVEG GYQCrPEISH sWGQYSPFFS LADQSEISPD
VPqNCKITFV	
<i>T. lanuginosa</i>	~~~~~ ----nvDIAR hWGQYSPFFS LAEvSEISPA
VPkGCRVeFV	

*M. thermophila*  
IPdDCeVTFa

ESRPCDTpDl GFQCgTAISH FWGQYSPYFS VPsElDaS..

Basidio  
pPaGCQIxqV

xSxPxrxTAA qLPipxQxqx xWSPYSPYFP VxxyxA....

Consensus NSHSCD TVDG GYQC-PEISH LWGQYSPFFS LADESAISPD VP-  
GCRVTFV

**Fcp10 NSHSCD TVDG GYQCFPEISH LWGQYSPFFS LADESAISPD  
VPKGCRVTFV**

51

100

*A. terreus* 9a1  
QSYNYSLDSE

QVLARHGARs PThSKTKaYA AtIaAIQKSA TaFpGKYAFL

*A. terreus* cbs  
KSYNYSMGSE

QVLARHGARs PTdSKTKaYA AtIaAIQKNA TaLpGKYAFL

*A. niger* var. *awamori* QVLSRHGARY PTesKGKKYS ALIeEIQQnv TtFDGKYAFL  
KTYNYSLGAD

*A. niger* NRRL3135 QVLSRHGARY PTdSKGKKYS ALIeEIQQNA TtFDGKYAFL  
KTYNYSLGAD

*A. fumigatus* 13073 QVLSRHGARY PTSSKSKKYk kLVtAIQaNA TdFKGKFAFL  
KTYNYTLGAD

*A. fumigatus* 32722 QVLSRHGARY PTSSKSKKYk kLVtAIQaNA TdFKGKFAFL  
KTYNYTLGAD

*A. fumigatus* 58128 QVLSRHGARY PTSSKSKKYk kLVtAIQaNA TdFKGKFAFL  
KTYNYTLGAD

*A. fumigatus* 26906 QVLSRHGARY PTSSKSKKYk kLVtAIQaNA TdFKGKFAFL  
KTYNYTLGAD

*A. fumigatus* 32239 QVLSRHGARY PTASKSKKYk kLVtAIQKNA TeFKGKFAFL  
ETYNITLGAD

*E. nidulans* QVLSRHGARY PTeSKSKaYS GLIeAIQKNA TsFwGQYAFL  
ESYNITLGAD

*T. thermophilus* QLLSRHGARY PTSSKTELYS qLIsrIQKtA TaYKGyYAFL  
KdYrYqLGAN

*T. lanuginosa* QVLSRHGARY PTAhKSEvYA ELLqrIQDtA TeFKGDFAFL  
RdYaYhLGAD

*M. thermophila* QVLSRHGARA PtlkRAasYv DLIdrIHhGA isYgPgYEFL  
RTYDYTLGAD

Basidio NIIqRHGARF PTSGaAtRiq AaVakLQsax xxtDPKLDFL  
xnxtYxLGxD

Consensus QVLSRHGARY PTSSKSKKYS ALI-AIQKNA T-FKGKYAFL  
KTYNITLGAD

Fcp10 QVLSRHGARY PTSSKSKKYS ALIEAIQKNA TAFKGKYAFL  
KTYNITLGAD

101

150

*A. terreus* 9a1 ELTPFGrNQL rDlGaQFYeR YNAL.TRhIn PFVRATDAsR  
VhESAeKFVE

*A. terreus* cbs NLTPFGrNQL qDlGaQFYRR YDTL.TRhIn PFVRAADSSr  
VhESAeKFVE

*A. niger* var. *awamori* DLTPFGEQEL VNSGIKFYQR YESL.TRnII PFIRSSGSsR  
VIASGEKFIE

*A. niger* NRRL3135 DLTPFGEQEL VNSGIKFYQR YESL.TRnIV PFIRSSGSsR  
VIASGKKFIE

*A. fumigatus* 13073 DLTPFGEQQL VNSGIKFYQR YKAL.ARsVV PFIRASGSDR  
VIASGEKFIE

*A. fumigatus* 32722 DLTPFGEQQL VNSGIKFYQR YKAL.ARsVV PFIRASGSDR  
VIASGEKFIE

*A. fumigatus* 58128 DLTPFGEQQL VNSGIKFYQR YKAL.ARsVV PFIRASGSDR  
VIASGEKFIE

*A. fumigatus* 26906 DLTAfGEQQL VNSGIKFYQR YKAL.ARsVV PFIRASGSDR  
VIASGEKFIE

*A. fumigatus* 32239 DLTPFGEQQM VNSGIKFYQK YKAL.AgsVV PFIRSSGSDR  
VIASGEKFIE

*E. nidulans* DLTiFGENQM VDsgaKFYRR YKnL.Arknt PFIRASGSDR  
VVASAEKFIN

*T. thermophilus* DLTPFGENQM IQlGIKFYnH YKSL.ARnaV PFVRCSGSDR  
VIASGr1FIE

*T. lanuginosa* NLTRFGEEQM MESGrQFYHR YREq.AReIV PFVRAAGSAR  
VIASAEfFnr

*M. thermophila* ELTRtGQQQM VNSGIKFYRR YRAL.ARksI PFVRTAGqDR  
VvhsAENftQ

Basidio DLvPFGAxQs sQAGqEaFtR YsxLvSxdnL PFVRASGSDR  
VVDSAtNWtA

Consensus DLTPFGEQQM VNSGIKFYRR YKAL-AR-IV PFVRASGSDR  
VIASAEKFIE

Fcp10 DLTPFGEQQM VNSGIKFYRR YKAL.ARkIV PFVRASGSDR  
VIASAEKFIE



151

200

*A. terreus* 9a1 GFQTARqDDh hAnphQPSPr VDVaIPEGsA YNNTLEHSLC  
 TAFes...St

*A. terreus* cbs GFQNARqGDP hAnphQPSPr VDVVIPEGtA YNNTLEHSIC  
 TAFEa...St

*A. niger* var. *awamori* GFQSTKLkDP rAqpgQSSPk IDVVISEAsS sNNTLDpGtC  
 TvFed...SE

*A. niger* NRRL3135 GFQSTKLkDP rAqpgQSSPk IDVVISEAsS sNNTLDpGtC  
 TvFed...SE

*A. fumigatus* 13073 GFQqAKLADP gAt.nRAAPa ISVIIPESet FNNTLDHGVC  
 TkFEa...SQ

*A. fumigatus* 32722 GFQqAKLADP gAt.nRAAPa ISVIIPESet FNNTLDHGVC  
 TkFEa...SQ

*A. fumigatus* 58128 GFQqAKLADP gAt.nRAAPa ISVIIPESet FNNTLDHGVC  
 TkFEa...SQ

*A. fumigatus* 26906 GFQqAKLADP gAt.nRAAPa ISVIIPESet FNNTLDHGVC  
 TkFEa...SQ

*A. fumigatus* 32239 GFQqANVADP gAt.nRAAPV ISVIIPESet YNNTLDHSVC  
 TnFEa...SE

*E. nidulans* GFRkAQLhDh g.s.gQATPV VNVIPEidG FNNTLDHStC  
 vSFEn...dE

*T. thermophilus* GFQSAKVlDP hSdKhDAPPt INVIIeEGpS YNNTLDtGsC  
 PvFed...Ss

*T. lanuginosa* GFQdAKdrDP rSnkdQAePV INVIISEEtG sNNTLDgltC  
 PAaEe...Ap

*M. thermophila* GFHSALLADR gStvrPTlPy dmVVIPETaG aNNTLHNDLC  
 TAFEegPySt

Basidio GFaxA..... ..sxntxxPx LxVILSExg. .NDTLDDNMC  
 .....PxAG

Consensus GFQSAKLADP -A---QASPV INVIIPEG-G YNNTLDHGLC  
 TAFE--P-SE

Fcp10 GFQSAKLADP GANPHQASPV INVIIPEGAG YNNTLDHGLC  
 TAFEE...SE

201

250

*A. terreus* 9a1 VGDDavANFT AVFAPAIaqR LEAdLPGVQL StDDVVNLMA  
 MCPFETVS1T

*A. terreus* cbs VGDAaADNFT AVFAPAIakR LEAdLPGVQL SADDVVNLMA  
 MCPFETVS1T

*A. niger* var. *awamori* LADtVEANFT AtFAPSIRqR LEndLSGVtL TDtEVtyLMD  
 MCSFDTIS1S

*A. niger* NRRL3135 LADtVEANFT AtFvPSIRqR LEndLSGVtL TDtEVtyLMD  
 MCSFDTIS1S

*A. fumigatus* 13073 LGDEVAANFT ALFAPdIRAR aEkhlPGVtL TDEDVVSLMD  
 MCSFDTVArT

*A. fumigatus* 32722 LGDEVAANFT ALFAPdIRAR aEkhlPGVtL TDEDVVSLMD  
 MCSFDTVArT

*A. fumigatus* 58128 LGDEVAANFT ALFAPdIRAR aEkhlPGVtL TDEDVVSLMD  
 MCSFDTVArT

*A. fumigatus* 26906 LGDEVAANFT ALFAPdIRAR aKkhLPGVtL TDEDVVSLMD  
 MCSFDTVArT

*A. fumigatus* 32239 LGDEVEANFT ALFAPAIRAR IEkhLPGVQL TDDDVVSLMD  
 MCSFDTVArT

*E. nidulans* rADEIEANFT AIMGPPIRkR LEndLPGIKL TNENViYlMD  
MCSFDTMaRT

*T. thermophilus* gGHDaQEKFk kqFAPAIleK IKDhLPGVDL AvsDVpyLMD  
LCPFETLArn

*T. lanuginosa* .DptqpAEFl qVFGPRVlkK ItkhMPGVNL TlEDVplFMD  
LCPFDTVGSd

*M. thermophila* IGDDaQDtYl StFAGPItAR VNAnLPGaNL TDADtVaLMD  
LCPFETVAsS

Basidio dSDpqxnXWl AVFAPPItAR LNAAApgANL TDxDaxNLxx  
LCPFETVS..

Consensus LGDDVEANFT AVFAPPiRAR LEA-LPGVNL TDEDVVNLMD  
MCPFDTVA-T

Fcp10 LGDDVEANFT AVFAPPiRAR LEAHLPGVNL TDEDVVNLMD  
MCPFDTVART

251

300

*A. terreus* 9a1 dD..Aht... ..LSPF CDLFta..tE WtQYNYLlSL  
dKYYGYGGGN

*A. terreus* cbs dD..Aht... ..LSPF CDLFta..aE WtQYNYLlSL  
dKYYGYGGGN

*A. niger* var. *awamori* Tv..DTK... ..LSPF CDLFTH..dE WiHYDYlQSL  
kKYYGHGAGN

*A. niger* NRRL3135 Tv..DTK... ..LSPF CDLFTH..dE WiNYDYlQSL  
kKYYGHGAGN

*A. fumigatus* 13073 SD..ASQ... ..LSPF CQLFTH..nE WkKYNYlQSL  
gKYYGYGAGN

*A. fumigatus* 32722 SD..ASQ... ..LSPF CQLFTH..nE WkKYNYLQSL  
gKYYGYGAGN

*A. fumigatus* 58128 SD..ASQ... ..LSPF CQLFTH..nE WkKYNYLQSL  
gKYYGYGAGN

*A. fumigatus* 26906 SD..ASQ... ..LSPF CQLFTH..nE WkKYNYLQSL  
gKYYGYGAGN

*A. fumigatus* 32239 AD..ASE... ..LSPF CAIFTH..nE WkKYDYLQSL  
gKYYGYGAGN

*E. nidulans* AH..GTE... ..LSPF CAIFTE..kE WlQYDYLQSL  
sKYYGYGAGS

*T. thermophilus* ht..DT.... ..LSPF CALsTQ..eE WqaYDYYQSL  
gKYYGnGGGN

*T. lanuginosa* PvlfPrQ... ..LSPF CHLFTa..dD WmaYDYYyTL  
dKYYSHGGGS

*M. thermophila* SsdpaTadag ggnggrpLSPF CrLFSE..sE WraYDYLQSV  
gKWYGYGPGN

Basidio .....xexxSxF CDLFexxpeE FxaFxYxgdL  
dKFYGTGyGQ

Consensus SD--ATQ--- -----LSPF CDLFTH---E W-QYDYLQSL -  
KYYGYGAGN

**Fcp10** SD..ATQ... ..LSPF CDLFTH..DE WlQYDYLQSL  
**GKYYGYGAGN**

301

350

*A. terreus* 9a1 PLGPvQGVGW aNELMARLTR A.PVHDHTCv NNTLDASPAT  
FPLNATLYAD

*A. terreus* cbs PLGPvQGVGW aNELIARLTR S.PVHDHTCv NNTLDANPAT  
FPLNATLYAD

*A. niger* var. *awamori* PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLDsNPAT  
FPLNSTLYAD

*A. niger* NRRL3135 PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLDSSPAT  
FPLNSTLYAD

*A. fumigatus* 13073 PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT  
FPLNATMYvD

*A. fumigatus* 32722 PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT  
FPLNATMYvD

*A. fumigatus* 58128 PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT  
FPLNATMYvD

*A. fumigatus* 26906 PLGPAQGIGF tNELIARLTR S.PVQDHTST NsTLvSNPAT  
FPLNATMYvD

*A. fumigatus* 32239 PLGPAQGIGF tNELIARLTN S.PVQDHTST NsTLdSDPAT  
FPLNATIYvD

*E. nidulans* PLGPAQGIGF tNELIARLTQ S.PVQDNTST NHTLDsNPAT  
FPLDrkLYAD

*T. thermophilus* PLGPAQGVGF vNELIARMTH S.PVQDYTTv NHTLDsNPAT  
FPLNATLYAD

*T. lanuginosa* AFGPSRGVGF vNELIARMTg NlPVKDHTTv NHTLDdNPET  
FPLDAvLYAD

*M. thermophila* PLGPTQGVGF vNELLARLA. GvPVRDgTST NRTLdGDPPrT  
FPLGrPLYAD

Basidio PLGPvQGVGY iNELLARLTx qa.VRDNTqT NRTLdSSPxT  
FPLNrTFYAD

Consensus PLGPAQGVGF -NELIARLTH S-PVQDHTST NHTLDsNPAT  
FPLNATLYAD

**Fcp10 PLGPAQGVGF VNELIARLTH S.PVQDHTST NHTLDSNPAT**  
**FPLNATLYAD**

351

400

*A. terreus* 9a1 FSHDSnLVSI FWALGLYNGT aPLSqTSVE. .SvsQTDGYA  
AAWTVPFAAR

*A. terreus* cbs FSHDSnLVSI FWALGLYNGT kPLSqTTVE. .ditrTDGYA  
AAWTVPFAAR

*A. niger* var. *awamori* FSHDNGIISI LFALGLYNGT kPLSTTTVE. .NitQTDGFS  
SAWTVPFASR

*A. niger* NRRL3135 FSHDNGIISI LFALGLYNGT kPLSTTTVE. .NitQTDGFS  
SAWTVPFASR

*A. fumigatus* 13073 FSHDNSMVSI FFALGLYNGT ePLSrTSVE. .SaKe1DGYS  
ASWvVPFGAR

*A. fumigatus* 32722 FSHDNSMVSI FFALGLYNGT gPLSrTSVE. .SaKe1DGYS  
ASWvVPFGAR

*A. fumigatus* 58128 FSHDNSMVSI FFALGLYNGT ePLSrTSVE. .SaKe1DGYS  
ASWvVPFGAR

*A. fumigatus* 26906 FSHDNSMVSI FFALGLYNGT ePLSrTSVE. .SaKe1DGYS  
ASWvVPFGAR

*A. fumigatus* 32239 FSHDNGMIPI FFAMGLYNGT ePLSqTSeE. .StKESNGYS  
ASWAVPFGAR

*E. nidulans* FSHDNSMISI FFAMGLYNGT qPLSmdSVE. .SiQEmDGYA  
ASWTVPFGAR

*T. thermophilus* FSHDNTMtSI FaALGLYNGT akLSTTeIK. .SiEETDGYS  
AAWTVPFGR

*T. lanuginosa* FSHDNTMtGI FsAMGLYNGT kPLSTSkIQP pTgAAADGYA  
ASWTVPFAAR

*M. thermophila* FSHDNdMMGV LgALGaYDgv pPLdkTA..R rdpEElGGYA  
ASWAVPFAAR

Basidio FSHDNqMVAI FsAMGLFNqS aPLdPSxpDP nrt.....Wv  
TSklVPFSAR

Consensus FSHDNTMVSI FFALGLYNGT -PLSTTSVEP -S-EETDGYA  
ASWTVPFAAR

**Fcp10** FSHDNTMVSI FFALGLYNGT KPLSTTSVE. .SIEETDGYA  
**ASWTVPFAAR**

401

450

*A. terreus* 9a1 AYVEMMQC.. ra.....EKEPL VRVLVNDVRM  
PLHGCPtDKL

*A. terreus* cbs AYIEMMQC.. ra.....EKQPL VRVLVNDVRM  
PLHGCAVDNL

*A. niger* var. *awamori* lYVEMMQC.. Qa.....EQEPL VRVLVNDRVV  
PLHGCPIDaL

*A. niger* NRRL3135 lYVEMMQC.. Qa.....EQEPL VRVLVNDRVV  
PLHGCPVDaL

*A. fumigatus* 13073 AYfEtMQC.. Ks.....EKEPL VRaLINDRVV  
PLHGCDVDKL

*A. fumigatus* 32722 AYfEtMQC.. Ks.....EKEPL VRaLINDRVV  
PLHGCDVDKL

*A. fumigatus* 58128 AYfEtMQC.. Ks.....EKESL VRaLINDRVV  
PLHGCDVDKL

*A. fumigatus* 26906 AYfEtMQC.. Ks.....EKEPL VRaLINDRVV  
PLHGCDVDKL

EP 1 092 764 A2

*A. fumigatus* 32239 AYfEtMQC.. Ks..... EKEPL VRaLINDRVV  
PLHGCAVDKL

*E. nidulans* AYfELMQC.. E..... KKEPL VRVLVNDRVV  
PLHGCAVDKF

*T. thermophilus* AYIEMMQC.. Dd..... sDEPV VRVLVNDRVV  
PLHGCEVDsL

*T. lanuginosa* AYVELLRC.. Etetsseeee EG...EDEPF VRVLVNDRVV  
PLHGCrVDRW

*M. thermophila* iYVEkMRC.. sggggggggg EGrqeKDEeM VRVLVNDRVM  
TLkGCGaDEr

Basidio mvVERLxCxx xgtxxxxxxx xxxxxxxxxxxx VRVLVNDaVq  
PLEfCGgDxd

Consensus AYVEMMQC-- E----- EG---EKEPL VRVLVNDRVV  
PLHGCGVDKL

Fcp10 AYVEMMQC.. EA..... EKEPL VRVLVNDRVV  
PLHGCGVDKL

451

482

*A. terreus* 9a1 GRCKrDAFVA GLSFAQAG.. GNWADCF~~~ ~~

*A. terreus* cbs GRCKrDDFVE GLSFARAG.. GNWAECF~~~ ~~

*A. niger* var. *awamori* GRCtrDsFVr GLSFARSG.. GDWAECsA~~ ~~

*A. niger* NRRL3135 GRCtrDsFVr GLSFARSG.. GDWAECFA~~ ~~

*A. fumigatus* 13073 GRCKlNDFVK GLSWARSG.. GNWGECFS~~ ~~

*A. fumigatus* 32722 GRCKlNDFVK GLSWARSG.. GNWGECFS~~ ~~

*A. fumigatus* 58128 GRCKlNDFVK GLSWARSG.. GNWGECFS~~ ~~

*A. fumigatus* 26906 GRCKlNDFVK GLSWARSG.. GNWGECFS~~ ~~

*A. fumigatus* 32239 GRCKlKDFVK GLSWARSG.. GNSEQSFS~~ ~~



<i>E. nidulans</i>	GRCtlDDWVE GLNFARSG.. GNWkLCFTl~ --
<i>T. thermophilus</i>	GRCKrDDFVr GLSFARqG.. GNWEGCYAas e-
<i>T. lanuginosa</i>	GRCRrDEWIK GLTFARqG.. GHWDrCF~~~ --
<i>M. thermophila</i>	GmCtlErFIE SMAFARGN.. GKWDlCFA~~ --
Basidio	GxCtlDAFVE SqxYAReDgq GDFEKCFAtp xx

Consensus GRCK-DDFVE GLSFARSG-- GNWEECFA-- --

**Fcp10 GRCKRDDFVE GLSFARSG.. GNWEECFA... ..**

**Figure 6**

CP-1

*Eco* RI M G V F V V L L S I A T L F G S T 17  
TATATGAATTCATGGGCGTGTTCGTGCTACTGTCCATTGCCACCTGTTCGGTTCCA  
1 -----+-----+-----+-----+-----+-----+ 60  
ATATACTTAAGTACCCGCACAAGCAGCACGATGACAGGTAACGGTGGAAACAAGCCAAGGT  
S G T A L G P R G N S H S C D T V D G G 37  
CATCCGGTACCGCCTTGGGTCCTCGTGGAATTCTCACTCTTGAGACTGTTGACGGTG  
61 -----+-----+-----+-----+-----+-----+ 120  
GTAGGCCATGGCGGAACCCAGGAGCACCATTAAAGAGTGAGAACACTGTGACAACCTGCCAC

CP-2

CP-3.10

Y Q C F P E I S H L W G Q Y S P F F S L 57  
GTTACCAATGTTTCCCAGAAATTTCTCACTTGTGGGGTCAATACTCTCCATTCTTCTCTT  
121 -----+-----+-----+-----+-----+-----+ 180

CAATGGTTACAAAGGGTCTTTAAAGAGTGAACACCCCAGTTATGAGAGGTAAGAAGAGAA

A D E S A I S P D V P K G C R V T F V Q 77

TGGCTGACGAATCTGCTATTTCTCCAGACGTTCCAAAGGGTTGTAGAGTTACTTTTCGTTT

181 -----+-----+-----+-----+-----+-----+ 240

ACCGACTGCTTAGACGATAAAGAGGTCTGCAAGGTTTCCCGACATCTCAATGAAAGCAAG

CP-4.10

CP-5.10

V L S R H G A R Y P T S S K S K K Y S A 97

AAGTTTTGTCTAGACACGGTGCTAGATACCCAACCTTCTTCTAAGTCTAAGAAGTACTCTG

241 -----+-----+-----+-----+-----+-----+ 300

TTCAAAACAGATCTGTGCCACGATCTATGGGTTGAAGAAGATTCAGATTCTTCATGAGAC

L I E A I Q K N A T A F K G K Y A F L K 117

CTTTGATTGAAGCTATTCAAAAGAACGCTACTGCTTTCAAGGGTAAGTACGCTTTCTTGA

301 -----+-----+-----+-----+-----+-----+ 360

GAAACTAACTTCGATAAGTTTTCTTGCGATGACGAAAGTCCCATTTCATGCGAAAGAACT

CP-6

CP-7.10

T Y N Y T L G A D D L T P F G E Q Q M V 137

AGACTTACAACCTACACTTTGGGTGCTGACGACTTGACTCCATTTCGGTGAACAACAAATGG

361 -----+-----+-----+-----+-----+-----+ 420

TCTGAATGTTGATGTGAAACCCACGACTGCTGAACTGAGGTAAGCCACTTGTGTTTACC

N S G I K F Y R R Y K A L A R K I V P F 157

TTAACTCTGGTATTAAGTTCTACAGAAGATACAAGGCTTTGGCTAGAAAGATTGTTCCAT

421 -----+-----+-----+-----+-----+ 480

AATTGAGACCATAATTCAAGATGTCTTCTATGTTCCGAAACCGATCTTTCTAACAAGGTA

CP-8.10

CP-9.10

V R A S G S D R V I A S A E K F I E G F 177

TCGTTAGAGCTTCTGGTTCTGACAGAGTTATTGCTTCTGCTGAAAAGTTCATTGAAGGTT

481 -----+-----+-----+-----+-----+ 540

AGCAATCTCGAAGACCAAGACTGTCTCAATAACGAAGACGACTTTTCAAGTAACTTCCAA

Q S A K L A D P G A N P H Q A S P V I N 197

TCCAATCTGCTAAGTTGGCTGACCCAGGTGCTAACCACACCAAGCTTCTCCAGTTATTA

541 -----+-----+-----+-----+-----+ 600

AGGTTAGACGATTCAACCGACTGGGTCCACGATTGGGTGTGGTTCGAAGAGGTCAATAAT

CP-10.10

CP-11.10

V I I P E G A G Y N N T L D H G L C T A 217

ACGTTATTATTCCAGAAGGTGCTGGTTACAACAACACTTTGGACCACGGTTTGTGTACTG

601 -----+-----+-----+-----+-----+ 660

TGCAATAATAAGGTCTTCCACGACCAATGTTGTTGTGAAACCTGGTGCCAAACACATGAC

F E E S E L G D D V E A N F T A V F A P 237

CTTTCGAAGAATCTGAATTGGGTGACGACGTTGAAGCTAACTTCACTGCTGTTTTCGCTC

661 -----+-----+-----+-----+-----+ 720

GAAAGCTTCTTAGACTTAACCCACTGCTGCAACTTCGATTGAAGTGACGACAAAAGCGAG

CP-12.10

P I R A R L E A H L P G V N L T D E D V 257

CACCTATTAGAGCTAGATTGGAAGCTCACTTGCCAGGTGTTAACTGACTGACGAAGACG

721 -----+-----+-----+-----+-----+-----+ 780

GTGGATAATCTCGATCTAACCTTCGAGTGAACGGTCCACAATTGAACTGACTGCTTCTGC

CP-13.10

V N L M D M C P F D T V A R T S D A T Q 277

TTGTTAACTTGATGGACATGTGTCCATTTCGACACTGTTGCTAGAACTTCTGACGCTACTC

781 -----+-----+-----+-----+-----+-----+ 840

AACAATTGAACTACCTGTACACAGGTAAGCTGTGACAACGATCTTGAAGACTGCGATGAG

L S P F C D L F T H D E W I Q Y D Y L Q 297

AATTGTCTCCATTCTGTGACTTGTTCACTCACGACGAATGGATTCAATACGACTACTTGC

841 -----+-----+-----+-----+-----+-----+ 900

TTAACAGAGGTAAGACACTGAACAAGTGAGTGCTTGCTTACCTAAGTTATGCTGATGAACG

CP-14.10CP-15.10

S L G K Y Y G Y G A G N P L G P A Q G V 317

AATCTTTGGGTAAGTACTACGGTTACGGTGCTGGTAACCCATTGGGTCCAGCTCAAGGTG

901 -----+-----+-----+-----+-----+-----+ 960

TTAGAAACCCATTCATGATGCCAATGCCACGACCATTGGGTAACCCAGGTCGAGTTCCAC

G F V N E L I A R L T H S P V Q D H T S 337

TTGGTTTCGTTAACGAATTGATTGCTAGATTGACTCACTCTCCAGTTCAAGACCACACTT

961 -----+-----+-----+-----+-----+-----+  
1020

AACCAAAGCAATTGCTTAACTAACGATCTAACTGAGTGAGAGGTCAAGTTCTGGTGTGAA

CP-16.10

CP-17.10

T N H T L D S N P A T F P L N A T L Y A 357

CTACTAACCACACTTTGGACTCTAACCAGCTACTTTCCCATGAACGCTACTTTGTACG

1021 -----+-----+-----+-----+-----+-----+  
1080

GATGATTGGTGTGAAACCTGAGATTGGGTCGATGAAAGGGTAACTTGGGATGAAACATGC

D F S H D N T M V S I F F A L G L Y N G 377

CTGACTTCTCTCAGACAACACTATGGTTTCTATTTTCTTCGCTTTGGGTTGTACAACG

1081 -----+-----+-----+-----+-----+-----+  
1140

GACTGAAGAGAGTGCTGTTGTGATACCAAAGATAAAAGAAGCGAAACCCAAACATGTTGC

CP-18.10

CP-19.10

T K P L S T T S V E S I E E T D G Y A A 397

GTACTAAGCCATTGTCTACTACTTCTGTTGAATCTATTGAAGAACTGACGGTTACGCTG

1141 -----+-----+-----+-----+-----+-----+  
1200

CATGATTCCGTAACAGATGATGAAGACAACTTAGATAACTTCTTTGACTGCCAATGCGAC

S W T V P F A A R A Y V E M M Q C E A E 417

CTTCTTGGA CTGTTCCATTGCTGCTAGAGCTTACGTTGAAATGATGCAATGTGAAGCTG  
 1201 -----+-----+-----+-----+-----+-----+  
 1260  
 GAAGAACCTGACAAGGTAAGCCACGATCTCGAATGCAACTTTACTACGTTACACTTCGAC  
 CP-20.10  
 CP-21.10  
 K E P L V R V L V N D R V V P L H G C G 437  
 AAAAGGAACCATTTGGTTAGAGTTTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTG  
 1261 -----+-----+-----+-----+-----+-----+  
 1320  
 TTTTCCTTGGTAAACCAATCTCAAAACCAATTGCTGTCTCAACAAGGTAACGTGCCAACAC  
 V D K L G R C K R D D F V E G L S F A R 457  
 GTGTTGACAAGTTGGGTAGATGTAAGAGAGACGACTTCGTTGAAGGTTTGTCTTTCGCTA  
 1321 -----+-----+-----+-----+-----+-----+  
 1380  
 CACAACCTGTTCAACCCATCTACATTCTCTCTGCTGAAGCAACTTCCAAACAGAAAGCGAT  
 CP-22.10  
 S G G N W E E C F A \* Eco RI 467  
 GATCTGGTGGTAACTGGGAAGAATGTTTCGCTTAAGAATTCATATA  
 1381 -----+-----+-----+-----+-----+----- 1426  
 CTAGACCACCATTGACCCTTCTTACAAAGCGAATTCTTAAGTATAT

Figure 7

1

50

*P. involutus* (phyA1) ~~~~~~ ~FPipeseqR nWSPYSPYFP LAEyKA....  
 pPaGCQInqV

*P. involutus* (phyA2) ~~~~~~ ~FsipeseqR nWSPYSPYFP LAEyKA....  
 pPaGCeInqV

*T. pubescens* ~~~~~~ ~LDvtRDVqQ sWSmYSPYFP aAtyvA....  
 pPaSCQInqV

*A. pediades* ~~~~~~ ~pffpPQIqD sWAaYTPYYP VqAyTP....  
 pPKDCKITqV

*P. lycii* ~~~~~~ ~LPipAQnTs nWGPYdPFFP VEpyAA....  
 pPEGCTVTqV

*A. terreus* 9a1 KhSDCNSVDh GYQCfPELSH kWGLYAPYFS LqDESFPF1D  
 VPEDCHITFV

*A. terreus* cbs NhSDCtSVDr GYQCfPELSH kWGLYAPYFS LqDESFPF1D  
 VPDDCHITFV

*A. niger* var. *awamori* NqSTCDTVDq GYQCfSEtSH LWGQYAPFFS LANESAISPD  
 VPaGCRVTFa

*A. niger* T213 NqSSCDTVDq GYQCfSEtSH LWGQYAPFFS LANESvISPD  
 VPaGCRVTFa

*A. niger* NRRL3135 NqSSCDTVDq GYQCfSEtSH LWGQYAPFFS LANESvISPE  
 VPaGCRVTFa

*A. fumigatus* ATCC13073 GSkSCDTVD1 GYQCSPAtSH LWGQYSPFFS LEDElSVSSK  
 LPKDCRITLV

*A. fumigatus* ATCC32722 GSkSCDTVD1 GYQCSPAtSH LWGQYSPFFS LEDElSVSSK  
 LPKDCRITLV

*A. fumigatus* ATCC58128 GSkSCDTVD1 GYQCSPAtSH LWGQYSPFFS LEDElSVSSK  
 LPKDCRITLV

<i>A. fumigatus</i> ATCC26906	GSkSCDTVDl GYQCSPAtSH LWGQYSPFFS LEDElSVSSK LPKDCRITLV
<i>A. fumigatus</i> ATCC32239	GSkACDTVEl GYQCSPGtSH LWGQYSPFFS LEDElSVSSD LPKDCRVTFV
<i>E. nidulans</i>	QNHSCNTaDg GYQCfPNVSH VWGQYSPYFS IEQESAISeD VPhGCeVTFV
<i>T. thermophilus</i>	DSHSCNTVEg GYQCrPEISH sWGQYSPFFS LADQSEISPD VPQNCKITFV
<i>T. lanuginosa</i>	----- ~~~~nVDIAR hWGQYSPFFS LAEvSEISPA VPKGCRVeFV
<i>M. thermophila</i>	ESRPCDTpDl GFQCgTAISH FWGQYSPYFS VPSElDaS.. IPDDCeVTFa
<b>Consensus Seq. 11</b>	<b>NSHSCDTVD- GYQC-PEISH LWGQYSPFFS LADESAISPD</b> <b>VPKGCRVTFV</b>
100	51
<i>P. involutus</i> (phyA1)	NIIqRHGARF PTSGaTtRik AgLtKLQgvq nftDAKFnFI KSFKYdLGns
<i>P. involutus</i> (phyA2)	NIIqRHGARF PTSGaAtRik AgLsKLQsvq nftDPKFDFI KSftYdLGts
<i>T. pubescens</i>	HIIqRHGARF PTSGaAKRiq TaVAKLKaaS nytDPfLLAFV tnYtYSLGqD
<i>A. pediades</i>	NIIqRHGARF PTSGaGtRiq AaVKKLQsak TytDPRLDFL tnYtYTLGhD
<i>P. lycii</i>	NLIqRHGARW PTSGarsRqv AaVAKIQmar PftDPKYEFL NdFvYkFGvA



<i>A. terreus</i> 9a1	QVLARHGARs PThSKTKaYA AtIAaIQKSA TaFpGKYAFL QSYNYSLDSE
<i>A. terreus</i> cbs	QVLARHGARs PTdSKTKaYA AtIAaIQKNA TaLpGKYAFL KSYNYSMGSE
<i>A. niger</i> var. <i>awamori</i>	QVLSRHGARY PTeSKGKKYS ALIEeIQQNv TtFDGKYAFL KTYNYSLGAD
<i>A. niger</i> T213	QVLSRHGARY PTeSKGKKYS ALIEeIQQNv TtFDGKYAFL KTYNYSLGAD
<i>A. niger</i> NRRL3135	QVLSRHGARY PTdSKGKKYS ALIEeIQQNA TtFDGKYAFL KTYNYSLGAD
<i>A. fumigatus</i> ATCC13073	QVLSRHGARY PTSSKSKKYk kLVtaIQaNA TdFKGKFAFL KTYNYTLGAD
<i>A. fumigatus</i> ATCC32722	QVLSRHGARY PTSSKSKKYk kLVtaIQaNA TdFKGKFAFL KTYNYTLGAD
<i>A. fumigatus</i> ATCC58128	QVLSRHGARY PTSSKSKKYk kLVtaIQaNA TdFKGKFAFL KTYNYTLGAD
<i>A. fumigatus</i> ATCC26906	QVLSRHGARY PTSSKSKKYk kLVtaIQaNA TdFKGKFAFL KTYNYTLGAD
<i>A. fumigatus</i> ATCC32239	QVLSRHGARY PTASKSKKYk kLVtaIQKNA TeFKGKFAFL ETNYNYTLGAD
<i>E. nidulans</i>	QVLSRHGARY PTeSKSKaYS GLIEaIQKNA TsFwGQYAFL ESYNYTLGAD
<i>T. thermophilus</i>	QLLSRHGARY PTSSKTELYS qLIIsRIQKtA TayKGyYAFL KdYrYqLGAN
<i>T. lanuginosa</i>	QVLSRHGARY PTAhKSEvYA ELLQRIQDtA TeFKGDFAFL RdYaYhLGAD
<i>M. thermophila</i>	QVLSRHGARA PtlkRAasYv DLIDRIHhGA isYgPgYEFL RTYDYTLGAD

Consensus Seq. 11  
 KTYNYTLGAD

QVLSRHGARY PTSSKSKKYS ALIERIQKNA T-FKGKYAFL

101

150

<i>P. involutus</i> (phyA1)	DLvPFGAAQs fDAGqEaFaR YskLvSKNnL PFIRAdGSDR VVDsAtNWtA
<i>P. involutus</i> (phyA2)	DLvPFGAAQs fDAGLEvFaR YskLvSsDnL PFIRsdGSDR VVDtAtNWtA
<i>T. pubescens</i>	sLveLGAtQs sEAGqEaFtR YsSLvSaDeL PFVRASGSDR VVATANNWtA
<i>A. pediades</i>	DLvPFGAlQs sQAGeEtFQR YsfLvSKEnL PFVRASSsNR VVDsAtNWtE
<i>P. lycii</i>	DLlPFGANQs hQTGtDMYtR YsTLfEgGdV PFVRAAGdQR VVDSSStNWtA
<i>A. terreus</i> 9a1	ELTPFGGrNQL rDlGaQFYeR YNAL.TRHIn PFVRATDAsR VhESAeKFVE
<i>A. terreus</i> cbs	NLTPFGGrNQL qDlGaQFYRR YDTL.TRHIn PFVRAADsSR VhESAeKFVE
<i>A. niger</i> var. <i>awamori</i>	DLTPFGEQEL VNsgIKFYQR YESL.TRNII PFIRSSGSsR VIASGEKFIE
<i>A. niger</i> T213	DLTPFGEQEL VNsgIKFYQR YESL.TRNII PFIRSSGSsR VIASGEKFIE
<i>A. niger</i> NRRL3135	DLTPFGEQEL VNsgIKFYQR YESL.TRNIV PFIRSSGSsR VIASGKKFIE
<i>A. fumigatus</i> ATCC13073	DLTPFGEQQL VNsgIKFYQR YKAL.ARSVV PFIRASGSDR VIASGEKFIE
<i>A. fumigatus</i> ATCC32722	DLTPFGEQQL VNsgIKFYQR YKAL.ARSVV PFIRASGSDR VIASGEKFIE

*A. fumigatus* ATCC58128 DLTPFGEQQL VNSGIKFYQR YKAL.ARSVV PFIRASGSDR  
VIASGEKFIE

*A. fumigatus* ATCC26906 DLTAfGEQQL VNSGIKFYQR YKAL.ARSVV PFIRASGSDR  
VIASGEKFIE

*A. fumigatus* ATCC32239 DLTPFGEQQM VNSGIKFYQK YKAL.AgSVV PFIRSSGSDR  
VIASGEKFIE

*E. nidulans* DLTiFGENQM VDSGaKFYRR YKnL.ARKnt PFIRASGSDR  
VVASAEKFIE

*T. thermophilus* DLTPFGENQM IQlGIKFYnH YKSL.ARNvV PFVRCGSDR  
VIASGr1FIE

*T. lanuginosa* NLTRFGEEQM MESGrQFYHR YREq.AREIV PFVRAAGSAR  
VIASAEfFnr

*M. thermophila* ELTRtGQQQM VNSGIKFYRR YRAL.ARKsI PFVRTAGqDR  
VvhSAENftQ

**Consensus Seq. 11** DLTPFGENQM VNSGIKFYRR YKAL-ARNIV PFVRASGSDR  
**VIASAEKFIE**

151

200

*P. involutus* (phyA1) GFaSA..... ..shNtvqPk LNLILPQ..T gNDTLEDNMC  
PAaGD.....

*P. involutus* (phyA2) GFaSA..... ..srNaiqPk LDLILPQ..T gNDTLEDNMC  
PAaGE.....

*T. pubescens* GFa1A..... ..ssNsiTPV LSvIIE..A gNDTLDDNMC  
PAaGD.....

*A. pediades* GFsAA..... ..shHv1NPI LfVILSE..S LNDTLDDAMC  
PnaGs.....

<i>P. lycii</i> PnevD.....	GFgdA..... ..sgEtv1Pt LQVVLQE..E gNcTLcNNMC
<i>A. terreus</i> 9a1 TAFES...ST	GFQTARqDDh hAnpHQPSPr VDVaIPEGSA YNNTLEHSLC
<i>A. terreus</i> cbs TAFEA...ST	GFQNARqGDP hAnpHQPSPr VDVVIPEGTA YNNTLEHSIC
<i>A. niger</i> var. <i>awamori</i> TvFED...Se	GFQSTKLkDP rAqpgQSSPk IDVWISEASS sNNTLDpGtC
<i>A. niger</i> T213 TvFED...Se	GFQSTKLkDP rAqpgQSSPk IDVWISEASS sNNTLDpGtC
<i>A. niger</i> NRRL3135 TvFED...Se	GFQSTKLkDP rAqpgQSSPk IDVWISEASS sNNTLDpGtC
<i>A. fumigatus</i> ATCC13073 TkFEA...Sq	GFQqAKLADP gAt.NRAAPa ISVIIPESeT FNNTLDHGVC
<i>A. fumigatus</i> ATCC32722 TkFEA...Sq	GFQqAKLADP gAt.NRAAPa ISVIIPESeT FNNTLDHGVC
<i>A. fumigatus</i> ATCC58128 TkFEA...Sq	GFQqAKLADP gAt.NRAAPa ISVIIPESeT FNNTLDHGVC
<i>A. fumigatus</i> ATCC26906 TkFEA...Sq	GFQqAKLADP gAt.NRAAPa ISVIIPESeT FNNTLDHGVC
<i>A. fumigatus</i> ATCC32239 TnFEA...Se	GFQqANVADP gAt.NRAAPV ISVIIPESeT YNNTLDHSVC
<i>E. nidulans</i> vSFEN...de	GFRkAQLhDh g.s.gQATPV VNVIIPeIdG FNNTLDHStC
<i>T. thermophilus</i> PvFED...SS	GFQSAKVlDP hSdKHDApPt INVIIeEGPS YNNTLDtGsC
<i>T. lanuginosa</i> PAaEE...AP	GFQdAKdrDP rSnkDQAePV INVIISEETG sNNTLDgltC

*M. thermophila* GFHSAILADR gStvRPTlPy dmVVIPETAG aNNTLHNDLC  
TAFEEgpyST

**Consensus Seq. 11** GFQSAKLADP -A--HQASPV INVIIPEGSG YNNTLDHGLC  
**TAFED---ST**

201

250

*P. involutus* (phyA1) .SDpqvnaWl AVafPSItAR LNAaaPSVNL TDtDafNLVs  
LCAFlTVSK.

*P. involutus* (phyA2) .SDpqvDaWl AsafPSVtAQ LNAaaPGaNL TDADafNLVs  
LCPFmTVSK.

*T. pubescens* .SDpqvnQWl AqFAPPMtAR LNagaPGaNL TDtDtyNLLt  
LCPFETVAt.

*A. pediades* .SDpqtGiWT SIYGTPIanR LNqqaPGaNI TAADVsnLIp  
LCAFETivK.

*P. lycii* .GDESt.tWl GVfAPhItAR LNAaaPSaNL SDsDaLtLMD  
MCPFDTLsS.

*A. terreus* 9a1 VGDDAvANFT AVFAPAIaQr LEAdLPGVQL StDDVVNLMA  
MCPFETVSlt

*A. terreus* cbs VGDAADNFT AVFAPAIakR LEAdLPGVQL SADDVVNLMA  
MCPFETVSlt

*A. niger* var. *awamori* LADtveANFT AtFAPSIRqR LEndLSGVtL TDtEVtyLMD  
MCSFDTIStS

*A. niger* T213 LADtveANFT AtFAPSIRqR LEndLSGVtL TDtEVtyLMD  
MCSFDTIStS

*A. niger* NRRL3135 LADtveANFT AtFvPSIRqR LEndLSGVtL TDtEVtyLMD  
MCSFDTIStS

<i>A. fumigatus</i> ATCC13073	LGDEvAANFT ALFAPdIRAR aEkhLPGVtL TDEDVVSLMD MCSFDTVART
<i>A. fumigatus</i> ATCC32722	LGDEvAANFT ALFAPdIRAR aEkhLPGVtL TDEDVVSLMD MCSFDTVART
<i>A. fumigatus</i> ATCC58128	LGDEvAANFT ALFAPdIRAR aEkhLPGVtL TDEDVVSLMD MCSFDTVART
<i>A. fumigatus</i> ATCC26906	LGDEvAANFT ALFAPdIRAR aKkhLPGVtL TDEDVVSLMD MCSFDTVART
<i>A. fumigatus</i> ATCC32239	LGDEvEANFT ALFAPAIRAR IEkhLPGVQL TDDDVVSLMD MCSFDTVART
<i>E. nidulans</i>	rADEiEANFT AIMGPPIRkR LEndLPGIKL TNENViYLMD MCSFDTMART
<i>T. thermophilus</i>	gGHDAQEKFA kqFAPAILEK IKDhLPGVDL AvsDVpyLMD LCPFETLArN
<i>T. lanuginosa</i>	.DptqpAEFl qVFGPRVlkK ItkhMPGVNL TLEDVplFMD LCPFDTVGSd
<i>M. thermophila</i>	IGDDAQDtYl StFAGPItAR VNAnLPGaNL TDADtVaLMD LCPFETVAsS
<b>Consensus Seq. 11</b>	<b>LGDDAEANFT AVFAPPiRAR LEA-LPGVNL TDEDVVNLMD MCPFDTVART</b>
	251
300	
<i>P. involutus</i> (phyA1)	..... . . . . ekkSdF CtLFegiPGs FeaFAYggdL dKFYGTGyGQ
<i>P. involutus</i> (phyA2)	..... . . . . eqkSdF CtLFegiPGs FeaFAYagdL dKFYGTGyGQ

<i>T. pubescens</i>	.....errSeF CDIYeelqAE .daFAYnadL
dKFYGTGyGQ	
<i>A. pediades</i>	.....etpSPF CNLF..TPEE FaQFEYFgdL
dKFYGTGyGQ	
<i>P. lycii</i>	.....gnaSPF CDLF..TAAE YvsYEEYydL
dKYYGTGPGN	
<i>A. terreus</i> 9a1	dD..Aht... ..LSPF CDLF..TatE WtQYNYLlSL
dKYYGYGGGN	
<i>A. terreus</i> cbs	dD..Aht... ..LSPF CDLF..TAAE WtQYNYLlSL
dKYYGYGGGN	
<i>A. niger</i> var. <i>awamori</i>	Tv..DTK... ..LSPF CDLF..ThDE WiHYDYlQSL
kKYYGHGAGN	
<i>A. niger</i> T213	Tv..DTK... ..LSPF CDLF..ThDE WiHYDYLRSL
kKYYGHGAGN	
<i>A. niger</i> NRRL3135	Tv..DTK... ..LSPF CDLF..ThDE WiNYDYlQSL
kKYYGHGAGN	
<i>A. fumigatus</i> ATCC13073	SD..ASQ... ..LSPF CQLF..ThNE WkKYNYLQSL
gKYYGYGAGN	
<i>A. fumigatus</i> ATCC32722	SD..ASQ... ..LSPF CQLF..ThNE WkKYNYLQSL
gKYYGYGAGN	
<i>A. fumigatus</i> ATCC58128	SD..ASQ... ..LSPF CQLF..ThNE WkKYNYLQSL
gKYYGYGAGN	
<i>A. fumigatus</i> ATCC26906	SD..ASQ... ..LSPF CQLF..ThNE WkKYNYLQSL
gKYYGYGAGN	
<i>A. fumigatus</i> ATCC32239	AD..ASE... ..LSPF CAIF..ThNE WkKYDYlQSL
gKYYGYGAGN	
<i>E. nidulans</i>	AH..GTE... ..LSPF CAIF..TEKE WlQYDYlQSL
sKYYGYGAGS	

<i>T. thermophilus</i>	ht..DT.... .....LSPF CALs..TqEE WqaYDYYQSL gKYYGnGGCN
<i>T. lanuginosa</i>	PvlfPrQ... .....LSPF CHLF..TADD WmaYDYYyTL dKYYSHGGGS
<i>M. thermophila</i>	SsdpATadag ggnggrpLSPF CrLF..SEsE WraYDYLQSV gKWYGYGPGN
<b>Consensus Seq. 11</b>	<b>SD--ATQ--- -----LSPF CDLF--TADE W-QYDYLQSL -</b> <b>KYYGYGAGN</b>
	301
350	
<i>P. involutus</i> (phyA1)	eLGPvQGVGY vNELIARLTN S.AVRDNTqT NRTLDA SPvT FPLNkTFYAD
<i>P. involutus</i> (phyA2)	ALGPvQGVGY iNELLARLTN S.AVNDNTqT NRTLDAApDT FPLNkTMYAD
<i>T. pubescens</i>	PLGPvQGVGY iNELIARLTa q.nVsDHTqT NsTLDSSPET FPLNrTLYAD
<i>A. pediades</i>	PLGPvQGVGY iNELLARLTE m.PVRDNTqT NRTLDS SPlT FPLDrSIYAD
<i>P. lycii</i>	ALGPvQGVGY vNELLARLTg q.AVRDETqT NRTLDSDPAT FPLNrTFYAD
<i>A. terreus</i> 9a1	PLGPvQGVGW aNELMARLTR A.PVHDHTCv NNTLDAS PAT FPLNATLYAD
<i>A. terreus</i> cbs	PLGPvQGVGW aNELIARLTR S.PVHDHTCv NNTLDANPAT FPLNATLYAD
<i>A. niger</i> var. <i>awamori</i>	PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLDSNPAT FPLNSTLYAD



<i>A. niger</i> T213	PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLDSNPAT FPLNSTLYAD
<i>A. niger</i> NRRL3135	PLGPTQGVGY aNELIARLTH S.PVHDDTSS NHTLDSSPAT FPLNSTLYAD
<i>A. fumigatus</i> ATCC13073	PLGPAQGIGF tNELIARLTR S.PVQDHTST NstLvSNPAT FPLNATMYvD
<i>A. fumigatus</i> ATCC32722	PLGPAQGIGF tNELIARLTR S.PVQDHTST NstLvSNPAT FPLNATMYvD
<i>A. fumigatus</i> ATCC58128	PLGPAQGIGF tNELIARLTR S.PVQDHTST NstLvSNPAT FPLNATMYvD
<i>A. fumigatus</i> ATCC26906	PLGPAQGIGF tNELIARLTR S.PVQDHTST NstLvSNPAT FPLNATMYvD
<i>A. fumigatus</i> ATCC32239	PLGPAQGIGF tNELIARLTN S.PVQDHTST NstLDSDPAT FPLNATIYvD
<i>E. nidulans</i>	PLGPAQGIGF tNELIARLTQ S.PVQDNTST NHTLDSNPAT FPLDrkLYAD
<i>T. thermophilus</i>	PLGPAQGVGF vNELIARMTH S.PVQDYTTv NHTLDSNPAT FPLNATLYAD
<i>T. lanuginosa</i>	AFGPSRGVGF vNELIARMTg NlPVKDHTTv NHTLDdNPET FPLDAvLYAD
<i>M. thermophila</i>	PLGPTQGVGF vNELLARLA. GvPVRDgTST NRTLGDPrT FPLGrPLYAD

**Consensus Seq. 11**      **PLGPAQGVGF -NELIARLTH S-PVQDHTST NHTLDSNPAT**  
**FPLNATLYAD**

<i>P. involutus</i> (phyA1)	FSHDNlMVAV FsAMGLFrqP aPLSTsvPNP wrt.....Wr TSSlVPFSGR
<i>P. involutus</i> (phyA2)	FSHDNlMVAV FsAMGLFrqS aPLSTSTpDP nrt.....Wl TSSvVPFSAR
<i>T. pubescens</i>	FSHDNqMVAI FsAMGLFNqS aPLdPTTpDP art.....Fl vkkivPFASAR
<i>A. pediades</i>	LSHDNqMIAI FsAMGLFNqS sPLdPSfpNP krt.....Wv TSRltPFASAR
<i>P. lycii</i>	FSHDNTMVPI FaALGLFNAT a.LdPlkpDe nrl.....Wv DSklVPFSGH
<i>A. terreus</i> 9a1	FSHDSnLVSI FWALGLYNGT aPLSqTSVES Vs..QTDGYA AAWTVPFASAR
<i>A. terreus</i> cbs	FSHDSnLVSI FWALGLYNGT KPLSqTTVE d It..rTDGYA AAWTVPFASAR
<i>A. niger</i> var. <i>awamori</i>	FSHDNGIISI LFALGLYNGT KPLSTTTVEN It..QTDGFS SAWTVPFASR
<i>A. niger</i> T213	FSHDNGIISI LFALGLYNGT KPLSTTTVEN It..QTDGFS SAWTVPFASR
<i>A. niger</i> NRRL3135	FSHDNGIISI LFALGLYNGT KPLSTTTVEN It..QTDGFS SAWTVPFASR
<i>A. fumigatus</i> ATCC13073	FSHDNSMVSI FFALGLYNGT EPLSrTSVES ak..EldGYS ASWvVPFGAR
<i>A. fumigatus</i> ATCC32722	FSHDNSMVSI FFALGLYNGT gPLSrTSVES ak..EldGYS ASWvVPFGAR
<i>A. fumigatus</i> ATCC58128	FSHDNSMVSI FFALGLYNGT EPLSrTSVES ak..EldGYS ASWvVPFGAR
<i>A. fumigatus</i> ATCC26906	FSHDNSMVSI FFALGLYNGT EPLSrTSVES ak..EldGYS ASWvVPFGAR

<i>A. fumigatus</i> ATCC32239	FSHDNGMIPI FFAMGLYNGT EPLSqtSeES tk..ESNGYS ASWAVPFGAR
<i>E. nidulans</i>	FSHDNSMISI FFAMGLYNGT QPLSmdSVES Iq..EmDGYA ASWTVPFGAR
<i>T. thermophilus</i>	FSHDNTMtSI FaALGLYNGT akLSTTeIKS Ie..ETDGYS AAWTVPFGGR
<i>T. lanuginosa</i>	FSHDNTMtGI FsAMGLYNGT KPLSTSkIQP ptgaAADGYA ASWTVPFAAR
<i>M. thermophila</i>	FSHDNdMMGV LgALGaYDGv pPLdkTArrd ..peElGGYA ASWAVPFAAR
<b>Consensus Seq. 11</b>	<b>FSHDNTMVSI FFALGLYNGT KPLSTTSVES I---ETDGYA ASWTVPFAAR</b>
	401
450	
<i>P. involutus</i> (phyA1)	mvVErLsC.. fGt..... Tk VRVLVQDQVq PLEfCGgDRn
<i>P. involutus</i> (phyA2)	maVErLsC.. AGt..... Tk VRVLVQDQVq PLEfCGgDQd
<i>T. pubescens</i>	mvVErLDC.. GGa..... Qs VRLLVNDaVq PLafCGaDts
<i>A. pediades</i>	mvTErLlCQr DGTGsGGpsr imrNgnvQTF VRILVNDaLq PLkfCGgDmd
<i>P. lycii</i>	mtVEkLaC.. .....sgKea VRVLVNDaVq PLEfCGg.vd
<i>A. terreus</i> 9a1	AYVEMMQCrA ..... ..EK...EPL VRVLVNDRVM PLHGCPtDKL

EP 1 092 764 A2

<i>A. terreus</i> cbs	AYIEMMQCrA .....	..EK...QPL VRVLVNDVRVM
PLHGCAVDNL		
<i>A. niger</i> var. <i>awamori</i>	lYVEMMQCQA .....	..EQ...EPL VRVLVNDRVV
PLHGCPIDaL		
<i>A. niger</i> T213	lYVEMMQCQA .....	..EQ...EPL VRVLVNDRVV
PLHGCPIDaL		
<i>A. niger</i> NRRL3135	lYVEMMQCQA .....	..EQ...EPL VRVLVNDRVV
PLHGCPVDaL		
<i>A. fumigatus</i> ATCC13073	AYfEtMQCKS .....	..EK...EPL VRaLINDRVV
PLHGCDVDKL		
<i>A. fumigatus</i> ATCC32722	AYfEtMQCKS .....	..EK...EPL VRaLINDRVV
PLHGCDVDKL		
<i>A. fumigatus</i> ATCC58128	AYfEtMQCKS .....	..EK...ESL VRaLINDRVV
PLHGCDVDKL		
<i>A. fumigatus</i> ATCC26906	AYfEtMQCKS .....	..EK...EPL VRaLINDRVV
PLHGCDVDKL		
<i>A. fumigatus</i> ATCC32239	AYfEtMQCKS .....	..EK...EPL VRaLINDRVV
PLHGCAVDKL		
<i>E. nidulans</i>	AYfELMQCE. ....	..KK...EPL VRVLVNDRVV
PLHGCAVDKF		
<i>T. thermophilus</i>	AYIEMMQCDD .....	..sD...EPV VRVLVNDRVV
PLHGCEVDsL		
<i>T. lanuginosa</i>	AYVELLRcET ETsSeEEeEG ..	..ED...EPF VRVLVNDRVV
PLHGCrVDRW		
<i>M. thermophila</i>	iYVEkMRCsG GGgGgGGgEG ..	..rQekdEeM VRVLVNDRVV
TLkGCGaDEr		
<b>Consensus Seq. 11</b>	<b>AYVEMMQCEA GG-G-GG-EG --</b>	<b>..EK---EPL VRVLVNDRVV</b>
<b>PLHGCGVDKL</b>		

	451	482
<i>P. involutus</i> (phyA1)	GlCtLAKFVE SqTFARSDga GDFEKCFAts a~	
<i>P. involutus</i> (phyA2)	GlCaLDKFVE SqAYARSGga GDFEKCLAtt v~	
<i>T. pubescens</i>	GvCtLDAFVE SqAYARNDge GDFEKCFAt~ ~~	
<i>A. pediades</i>	S1CtLEAFVE SqkYAReDgq GDFEKCfD~~ ~~	
<i>P. lycii</i>	GvCELSAFVE SqTYAReNgq GDFAKCgfv se	
<i>A. terreus</i> 9a1	GRCKrDAFVA GLSFAQAG.. GNWADCF~~~ ~~	
<i>A. terreus</i> cbs	GRCKrDDFVE GLSFARAG.. GNWAE CF~~~ ~~	
<i>A. niger</i> var. <i>awamori</i>	GRCtrDsFVr GLSFARSG.. GDWAECSA~~ ~~	
<i>A. niger</i> T213	GRCtrDsFVr GLSFARSG.. GDWAE CFA~~ ~~	
<i>A. niger</i> NRRL3135	GRCtrDsFVr GLSFARSG.. GDWAE CFA~~ ~~	
<i>A. fumigatus</i> ATCC13073	GRCKLNDFVK GLSWARSG.. GNWGE CFS~~ ~~	
<i>A. fumigatus</i> ATCC32722	GRCKLNDFVK GLSWARSG.. GNWGE CFS~~ ~~	
<i>A. fumigatus</i> ATCC58128	GRCKLNDFVK GLSWARSG.. GNWGE CFS~~ ~~	
<i>A. fumigatus</i> ATCC26906	GRCKLNDFVK GLSWARSG.. GNWGE CFS~~ ~~	
<i>A. fumigatus</i> ATCC32239	GRCKLKDFVK GLSWARSG.. GNSEQSFS~~ ~~	
<i>E. nidulans</i>	GRCTLDDWVE GLNFARSG.. GNWktCFT1~ ~~	
<i>T. thermophilus</i>	GRCKrDDFVr GLSFARqG.. GNWEGCYAas e~	
<i>T. lanuginosa</i>	GRCRrDEWIK GLTFARqG.. GHWDrCF~~~ ~~	
<i>M. thermophila</i>	GmCtLErFIE SMAFARGN.. GKWDlCFA~~ ~~	
<b>Consensus Seq. 11</b>	<b>GRCKLDDFVE GLSFARSG-- GNWAE CFA-- --</b>	

**Figure 8**

M G V F V V L L S I A T L F G S T S G T  
 20  
 ATGGGCGTGTTTCGTCTGCTACTGTCCATTGCCACCTTGTTTCGGTTCACATCCGGTACC  
 1 ---+-----+-----+-----+-----+-----+-----  
 60  
 TACCCGCACAAGCAGCACGATGACAGGTAACGGTGGAACAAGCCAAGGTGTAGGCCATGG  
 A L G P R G N S H S C D T V D G G Y Q C  
 40  
 GCCTTGGGTCTCTCGTGGTAATTCTCACTCTTGTGACACTGTTGACGGTGGTTACCAATGT  
 61 ---+-----+-----+-----+-----+-----+-----  
 120  
 CGGAACCCAGGAGCACCATTAAGAGTGAGAACACTGTGACAACCTGCCACCAATGGTTACA  
 F P E I S H L W G T Y S P Y F S L A D E  
 60  
 TTCCCAGAAATTTCTCACTTGTGGGGTACCTACTCTCCATACTTCTCTTTGGCAGACGAA  
 121 ---+-----+-----+-----+-----+-----+-----  
 180  
 AAGGGTCTTTAAAGAGTGAACACCCCATGGATGAGAGGTATGAAGAGAAACCGTCTGCTT  
 S A I S P D V P D D C R V T F V Q V L S  
 80  
 TCTGCTATTTCTCCAGACGTTCCAGACGACTGTAGAGTTACTTTTCGTTCAAGTTTGTCT  
 187 ---+-----+-----+-----+-----+-----+-----  
 240

AGACGATAAAGAGGTCTGCAAGGTCTGCTGACATCTCAATGAAAGCAAGTTCAAAACAGA

R H G A R Y P T S S A S K A Y S A L I E

100

AGACACGGTGCTAGATACCCAACCTCTTCTGCGTCTAAGGCTTACTCTGCTTTGATTGAA

241 ---+-----+-----+-----+-----+-----+-----+-----

300

TCTGTGCCACGATCTATGGGTGAAGAAGACGCAGATTCCGAATGAGACGAACTAACTT

A I Q K N A T A F K G K Y A F L K T Y N

120

GCTATTCAAAGAACGCTACTGCTTTCAAGGGTAAGTACGCTTTCTTGAAGACTTACAAC

301 ---+-----+-----+-----+-----+-----+-----+-----

360

CGATAAGTTTTCTTGCGATGACGAAAGTTCCCATTCATGCGAAAGAACTTCTGAATGTTG

Y T L G A D D L T P F G E N Q M V N S G

140

TACACTTTGGGTGCTGACGACTTGACTCCATTCGGTGAAAACCAAATGGTTAACTCTGGT

361 ---+-----+-----+-----+-----+-----+-----+-----

420

ATGTGAAACCCACGACTGCTGAACTGAGGTAAGCCACTTTTGGTTTACCAATTGAGACCA

I K F Y R R Y K A L A R K I V P F I R A

160

ATTAAGTTCTACAGAAGATACAAGGCTTTGGCTAGAAAGATTGTTCCATTCATTAGAGCT

421 ---+-----+-----+-----+-----+-----+-----+-----

480

TAATTCAAGATGTCTTCTATGTTCCGAAACCGATCTTTCTAACAAGGTAAGTAATCTCGA

S G S D R V I A S A E K F I E G F Q S' A

180

TCTGGTTC TGACAGAGTTATTGCTTCTGCTGAAAAGTTCATTGAAGGTTTCCAATCTGCT

481 ---+-----+-----+-----+-----+-----+-----

540

AGACCAAGACTGTCTCAATAACGAAGACGACTTTTCAAGTAACTTCCAAAGGTTAGACGA

K L A D P G S Q P H Q A S P V I N V I I

200

AAGTTGGCTGACCCAGGTCTCAACCACACCAAGCTTCTCCAGTTATTAACGTGATCATT

541 ---+-----+-----+-----+-----+-----+-----

600

TTCAACCGACTGGGTCCAAGAGTTGGTGTGGTTCGAAGAGGTCAATAATTGCACTAGTAA

P E G S G Y N N T L D H G T C T A F E D

220

CCAGAAGGATCCGGTTACAACAACACTTTGGACCACGGTACTTGTACTGCTTTTGAAGAC

601 ---+-----+-----+-----+-----+-----+-----

660

GGTCTTCCTAGGCCAATGTTGTTGTGAAACCTGGTGCCATGAACATGACGAAGCTTCTG

S E L G D D V E A N F T A L F A P A I R

240

TCTGAATTAGGTGACGACGTTGAAGCTAACTTCACTGCTTTGTTTCGCTCCAGCTATTAGA

661 ---+-----+-----+-----+-----+-----+-----

720



AGACTTAATCCACTGCTGCAACTTCGATTGAAGTGACGAAACAAGCGAGGTCGATAATCT

260

A R L E A D L P G V T L T D E D V V Y L

GCTAGATTGGAAGCTGACTTGCCAGGTGTTACTTTGACTGACGAAGACGTTGTTTACTTG

780

721 ---+-----+-----+-----+-----+-----+-----

CGATCTAACCTTCGACTGAACGGTCCACAATGAAACTGACTGCTTCTGCAACAAATGAAC

280

M D M C P F D T V A R T S D A T E L S P

ATGGACATGTGTCCATTCGACACTGTCGCTAGAACTTCTGACGCTACTGAATTGTCTCCA

840

781 ---+-----+-----+-----+-----+-----+-----

TACCTGTACACAGGTAAGCTGTGACAGCGATCTTGAAGACTGCGATGACTTAACAGAGGT

300

F C A L F T H D E W I Q Y D Y L Q S L G

TTCTGTGCTTTGTTCACTCACGACGAATGGATCCAATACGACTACTTGCAAAGCTTGGGT

900

841 ---+-----+-----+-----+-----+-----+-----

AAGACACGAAACAAGTGAGTGCTGCTTACCTAGGTTATGCTGATGAACGTTTCGAACCCA

320

K Y Y G Y G A G N P L G P A Q G V G F A

AAGTACTACGGTTACGGTGCTGGTAACCCATTGGGTCCAGCTCAAGGTGTTGTTTCGCT

960

901 ---+-----+-----+-----+-----+-----+-----

TTCATGATGCCAATGCCACGACCATTGGGTAACCCAGGTCGAGTTCCACAACCAAAGCGA

N E L I A R L T H S P V Q D H T S T N H

340

AACGAATTGATTGCTAGATTGACTCACTCTCCAGTTCAAGACCACACTTCTACTAACCAC

961 ---+-----+-----+-----+-----+-----+-----

1020

TTGCTTAACTAACGATCTAACTGAGTGAGAGGTCAAGTTCTGGTGTGAAGATGATTGGTG

T L D S N P A T F P L N A T L Y A D F S

360

ACTTTGGACTCTAACCCAGCTACTTTCCCATTGAACGCTACTTTGTACGCTGACTTCTCT

1021 ---+-----+-----+-----+-----+-----+-----

1080

TGAAACCTGAGATTGGGTCGATGAAAGGGTAACCTGCGATGAAACATGCGACTGAAGAGA

H D N T M I S I F F A L G L Y N G T K P

380

CACGACAACACTATGATATCTATTTTCTTCGCTTTGGGTTTGTACAACGGTACCAAGCCA

1081 ---+-----+-----+-----+-----+-----+-----

1140

GTGCTGTTGTGATACTATAGATAAAAGAAGCGAAACCCAAACATGTTGCCATGGTTCGGT

L S T T S V E S I E E T D G Y S A S W T

400

TTGTCTACTACTTCTGTTGAATCTATTGAAGAACTGACGGTTACTCTGCTTCTTGGACT

1141 ---+-----+-----+-----+-----+-----+-----

1200

AACAGATGATGAAGACAACTTAGATAAATTCTTTGACTGCCAATGAGACGAAGAACCTGA

V P F A A R A Y V E M M Q C Q A E K E P

420

GTTCCATTGCTGCTAGAGCTTACGTTGAAATGATGCAATGTCAAGCTGAAAAGGAACCA

1201 ---+-----+-----+-----+-----+-----+-----

1260

CAAGGTAAGCGACGATCTCGAATGCAACTTTACTACGTTACAGTTCGACTTTTCCTTGGT

L V R V L V N D R V V P L H G C A V D K

440

TTGGTTAGAGTTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTGCTGTTGACAAG

1261 ---+-----+-----+-----+-----+-----+-----

1320

AACCAATCTCAAAACCAATTGCTGTCTCAACAAGGTAACGTGCCAACACGACAACTGTTT

L G R C K R D D F V E G L S F A R S G G

460

TTGGGTAGATGTAAGAGAGACGACTTCGTTGAAGGTTTGTCTTTCGCTAGATCTGGTGGT

1321 ---+-----+-----+-----+-----+-----+-----

1380

AACCCATCTACATTCTCTCTGCTGAAGCAACTTCCAAACAGAAAGCGATCTAGACCACCA

N W A E C F A \* 467

AACTGGGCTGAATGTTTCGCTTAA

1381 ---+-----+-----+ 1410

TTGACCCGACTTACAAAGCGAATT

Figure 9

M G V F V V L L S I A T L F G S T S G T  
 20  
 ATGGGCGTGTTTCGTCGTGCTACTGTCCATTGCCACCTTGTTTCGGTTCCACATCCGGTACC  
 1 -----+-----+-----+-----+-----+-----+  
 60  
 TACCCGCACAAGCAGCACGATGACAGGTAACGGTGGAACAAGCCAAGGTGTAGGCCATGG  
 A L G P R G N S H S C D T V D G G Y Q C  
 40  
 GCCTTGGGTCTCTCGTGGTAACTCTCACTCTTGTGACACTGTTGACGGTGGTTACCAATGT  
 61 -----+-----+-----+-----+-----+-----+  
 120  
 CGGAACCCAGGAGCACCATTGAGAGTGAGAACACTGTGACAACCTGCCACCAATGGTTACA  
 A F P E I S H L W G T Y S P F F S L A D E  
 60  
 TTCCCAGAAATTTCTCACTTGTGGGGTACATACTCTCCATTCTTCTCTTTGGCTGACGAA  
 121 -----+-----+-----+-----+-----+-----+  
 180  
 AAGGGTCTTTAAAGAGTGAACACCCCATGTATGAGAGGTAAGAAGAGAAACCGACTGCTT  
 S A I S P D V P K G C R V T F V Q V L S  
 80  
 TCTGCTATTTCTCCAGACGTTCCAAAGGGTGTAGAGTTACTTTCGTTCAAGTTTGTCT  
 181 -----+-----+-----+-----+-----+-----+  
 240

AGACGATAAAGAGGTCTGCAAGGTTTCCCAACATCTCAATGAAAGCAAGTTCAAAACAGA

R H G A R Y P T S S A S K A Y S A L I E

100

AGACACGGTGCTAGATACCCAACCTTCTTCTGCGTCTAAGGCGTACTCTGCTTTGATTGAA

241 -----+-----+-----+-----+-----+-----+

300

TCTGTGCCACGATCTATGGGTTGAAGAAGACGCAGATTCCGCATGAGACGAACTAACTT

A I Q K N A T A F K G K Y A F L K T Y N

120

GCTATTCAAAAGAACGCTACTGCTTTCAAGGGTAAGTACGCTTCTTGAAGACTTACAAC

301 -----+-----+-----+-----+-----+-----+

360

CGATAAGTTTTCTTGCGATGACGAAAGTTCCCATTCATGCGAAAGAACTTCTGAATGTTG

Y T L G A D D L T P F G E Q Q M V N S G

140

TACACTTTGGGTGCTGACGACTTGACTCCATTCCGGTGAACAACAAATGGTTAACTCTGGT

361 -----+-----+-----+-----+-----+-----+

420

ATGTGAAACCCACGACTGCTGAACTGAGGTAAGCCACTTGTTGTTTACCAATTGAGACCA

I K F Y R R Y K A L A R K I V P F I R A

160

ATTAAGTTCTACAGAAGATACAAGGCTTTGGCTAGAAAGATTGTTCCATTTCATTAGAGCT

421 -----+-----+-----+-----+-----+-----+

480

TAATTCAAGATGTCTTCTATGTTCCGAAACCGATCTTTCTAACAAGGTAAGTAATCTCGA

S G S D R V I A S A E K F I E G F Q S A

180

TCTGGTTCTGACAGAGTTATTGCTTCTGCTGAAAAGTTCATTGAAGGTTCCAATCTGCT

481 -----+-----+-----+-----+-----+-----+

540

AGACCAAGACTGTCTCAATAACGAAGACGACTTTTCAAGTAACTTCCAAAGGTTAGACGA

K L A D P G A N P H Q A S P V I N V I I

200

AAGTTGGCTGACCCAGGTGCTAACCCACACCAAGCTTCTCCAGTTATTAACGTTATTATT

541 -----+-----+-----+-----+-----+-----+

600

TTCAACCGACTGGGTCCACGATTGGGTGTGGTTCGAAGAGGTCAATAATTGCAATAATAA

P E G A G Y N N T L D H G L C T A F E E

220

CCAGAAGGTGCTGGTTACAACAACACTTTGGACCACGGTTTGTGTACTGCTTTCGAAGAA

601 -----+-----+-----+-----+-----+-----+

660

GGTCTTCCACGACCAATGTTGTTGTGAAACCTGGTGCCAAACACATGACGAAAGCTTCTT

S E L G D D V E A N F T A V F A P P I R

240

TCTGAATTGGGTGACGACGTTGAAGCTAACTTCACTGCTGTTTTCGCTCCACCAATTAGA

661 -----+-----+-----+-----+-----+-----+

720

AGACTTAACCCACTGCTGCAACTTCGATTGAAGTGACGACAAAAGCGAGGTGGTTAATCT

A R L E A H L P G V N L T D E D V V N L

260

GCTAGATTGGAAGCTCACTTGCCAGGTGTTAACTTGACTGACGAAGACGTTGTTAACTTG

721 -----+-----+-----+-----+-----+-----+

780

CGATCTAACCTTCGAGTGAACGGTCCACAATTGAACTGACTGCTTCTGCAACAATTGAAC

M D M C P F D T V A R T S D A T Q L S P

280

ATGGACATGTGTCCATTCGACACTGTTGCTAGAACTTCTGACGCTACTCAATTGTCTCCA

781 -----+-----+-----+-----+-----+-----+

840

TACCTGTACACAGGTAAGCTGTGACAACGATCTTGAAGACTGCGATGAGTTAACAGAGGT

F C D L F T H D E W I Q Y D Y L Q S L G

300

TTCTGTGACTTGTTCACTCACGACGAATGGATTCAATACGACTACTTGCAATCTTTGGGT

841 -----+-----+-----+-----+-----+-----+

900

AAGACACTGAACAAGTGAGTGCTGCTTACCTAAGTTATGCTGATGAACGTTAGAAACCCA

K Y Y G Y G A G N P L G P A Q G V G F V

320

AAGTACTACGGTTACGGTGCTGGTAACCCATTGGGTCCAGCTCAAGGTGTTGGTTTCGTT

901 -----+-----+-----+-----+-----+-----+

960

TTCATGATGCCAATGCCACGACCATTGGGTAACCCAGGTCGAGTTCCACAACCAAAGCAA  
 N E L I A R L T H S P V Q D H T S T N H  
 340  
 AACGAATTGATTGCTAGATTGACTCACTCTCCAGTTCAAGACCACACTTCTACTAACCAC  
 961 -----+-----+-----+-----+-----+-----+  
 1020  
 TTGCTTAACTAACGATCTAACTGAGTGAGAGGTCAAGTTCTGGTGTGAAGATGATTGGTG  
 T L D S N P A T F P L N A T L Y A D F S  
 360  
 ACTTTGGACTCTAACCCAGCTACTTTCCCATGGAACGCTACTTTGTACGCTGACTTCTCT  
 1021 -----+-----+-----+-----+-----+-----+  
 1080  
 TGAAACCTGAGATTGGGTCGATGAAAGGGTAACTTGCGATGAAACATGCGACTGAAGAGA  
 H D N T M V S I F F A L G L Y N G T K P  
 380  
 CACGACAACACTATGGTTTCTATTTTCTTCGCTTTGGGTTTGTACAACGGTACTAAGCCA  
 1081 -----+-----+-----+-----+-----+-----+  
 1140  
 GTGCTGTTGTGATACCAAAGATAAAAGAAGCGAAACCCAAACATGTTGCCATGATTCGGT  
 L S T T S V E S I E E T D G Y S A S W T  
 400  
 TTGTCTACTACTTCTGTTGAATCTATTGAAGAACTGACGGTTACTCTGCTTCTTGACT  
 1141 -----+-----+-----+-----+-----+-----+  
 1200



AACAGATGATGAAGACAACTTAGATAACTTCTTTGACTGCCAATGAGACGAAGAACCTGA

V P F A A R A Y V E M M Q C E A E K E P

420

GTTCCATTTCGCTGCTAGAGCTTACGTTGAAATGATGCAATGTGAAGCTGAAAAGGAACCA

1201 -----+-----+-----+-----+-----+-----+

1260

CAAGGTAAGCGACGATCTCGAATGCAACTTTACTACGTTACACTTCGACTTTTCCTTGGT

L V R V L V N D R V V P L H G C G V D K

440

TTGGTTAGAGTTTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTGGTGTGACAAG

1261 -----+-----+-----+-----+-----+-----+

1320

AACCAATCTCAAAACCAATTGCTGTCTCAACAAGGTAACGTGCCAACACCACAACCTGTTTC

L G R C K R D D F V E G L S F A R S G G

460

TTGGGTAGATGTAAGAGAGACGACTTCGTTGAAGGTTTGTCTTTCGCTAGATCTGGTGGT

1321 -----+-----+-----+-----+-----+-----+

1380

AACCCATCTACATTCTCTCTGCTGAAGCAACTTCCAAACAGAAAGCGATCTAGACCACCA

N W E E C F A \* 467

AACTGGGAAGAATGTTTCGCTTAA

1381 -----+-----+----- 1404

TTGACCCTTCTTACAAAGCGAATT

**Figure 10**

```

M G V F V V L L S I A T L F G S T S G T 20
ATGGGGGTTTTCGTCGTTCTATTATCTATCGCGACTCTGTTCCGGCAGCACATCGGGCACT
1 -----+-----+-----+-----+-----+-----+ 60
TACCCCCAAAAGCAGCAAGATAATAGATAGCGCTGAGACAAGCCGTCGTGTAGCCCGTGA
A L G P R G N H S K S C D T V D L G Y Q 40
GCGCTGGGCCCCCGTGGAATCACTCCAAGTCCTGCGATACGGTAGACCTAGGGTACCAG
61 -----+-----+-----+-----+-----+-----+ 120
CGCGACCCGGGGGCACCTTTAGTGAGGTTTCAGGACGCTATGCCATCTGGATCCCATGGTC
C S P A T S H L W G T Y S P Y F S L E D 60
TGCTCCCCCTGCGACTTCTCATCTATGGGGCACGTACTCGCCATaCTTTTCGCTCGAGGAC
121 -----+-----+-----+-----+-----+-----+ 180
ACGAGGGGACGCTGAAGAGTAGATACCCCGtgCATGAGCGGTAtGAAAAGCGAGCTCCTG
E L S V S S K L P K D C R I T L V Q V L 80
GAGCTGTCCGTGTCGAGTAAGCTTCCCAAGGATTGCCGGATCACCTTGGTACAGGTGCTA
181 -----+-----+-----+-----+-----+-----+ 240
CTCGACAGGCACAGCTCATTCGAAGGGTTCCTAACGGCCTAGTGGAACCATGTCCACGAT
S R H G A R Y P T S S K S K K Y K K L I 100
TCGCGCCATGGAGCGCGGTACCCAACCAGCTCCAAGAGCAAAAAGTATAAGAAGCTTaTt
241 -----+-----+-----+-----+-----+-----+ 300

```

AGCGCGGTACCTCGCGCCATGGGTTGGTCGAGGTTCTCGTTTTTCATATTCTTCGAAtAa

T A I Q A N A T D F K G K Y A F L K T Y 120

ACGGCGATCCAGGCCAATGCCACCGACTTCAAGGGCAAGTAcGCCTTTTTGAAGACGTAC

301 -----+-----+-----+-----+-----+-----+ 360

TGCCGCTAGGTCCGGTTACGGTGGCTGAAGTTCCCGTTCAtgCGGAAAACTTCTGCATG

N Y T L G A D D L T P F G E Q Q L V N S 140

AACTATACTCTGGGTGCGGATGACCTCACTCCCTTTGGGGAGCAGCAGCTGGTGAACTCG

361 -----+-----+-----+-----+-----+-----+ 420

TTGATATGAGACCCACGCCTACTGGAGTGAGGGAAACCCCTCGTCGTCGACCACTTGAGC

G I K F Y Q R Y K A L A R S V V P F I R 160

GGCATCAAGTTCTACCAGAGGTACAAGGCTCTGGCGCGCAGTGTGGTGCCGTTTATTCGC

421 -----+-----+-----+-----+-----+-----+ 480

CCGTAGTTCAAGATGGTCTCCATGTTCCGAGACCGCGCGTCACACCACGGCAAATAAGCG

A S G S D R V I A S G E K F I E G F Q Q 180

GCCTCAGGCTCGGACCGGGTTATTGCTTCGGGAGAGAAGTTCATCGAGGGGTTCCAGCAG

481 -----+-----+-----+-----+-----+-----+ 540

CGGAGTCCGAGCCTGGCCCAATAACGAAGCCCTCTCTTCAAGTAGCTCCCCAAGGTCGTC

A K L A D P G A T N R A A P A I S V I I 200

GCGAAGCTGGCTGATCCTGGCGCGACGAACCGCGCCGCTCCGGCGATTAGTGTGATTATT

541 -----+-----+-----+-----+-----+-----+ 600

CGCTTCGACCGACTAGGACCGCGCTGCTTGGCGCGGCGAGGCCGCTAATCACACTAATAA

P E S E T F N N T L D H G V C T K F E A 220

CCGAGAGCGAGACGTTCAACAATACGCTGGACCACGGTGTGTGCACGAAGTTTGAGGCG

601 -----+-----+-----+-----+-----+-----+ 660

GGCCTCTCGCTCTGCAAGTTGTTATGCGACCTGGTGCCACACACGTGCTTCAAACGCCG

S Q L G D E V A A N F T A L F A P D I R 240

AGTCAGCTGGGAGATGAGGTTGCGGCCAATTTCACTGCGCTCTTTGCACCCGACATCCGA

661 -----+-----+-----+-----+-----+-----+ 720

TCAGTCGACCCTCTACTCCAACGCCGGTTAAAGTGACGCGAGAAACGTGGGCTGTAGGCT

A R L E K H L P G V T L T D E D V V S L 260

GCTCGCctCGAGAAGCATCTTCCTGGCGTGACGCTGACAGACGAGGACGTTGTCAGTCTA

721 -----+-----+-----+-----+-----+-----+ 780

CGAGCGgaGCTCTTCGTAGAAGGACCGCACTGCGACTGTCTGCTCCTGCAACAGTCAGAT

M D M C P F D T V A R T S D A S Q L S P 280

ATGGACATGTGTcCGTTTGATACGGTAGCGCGCACCAGCGACGCAAGTCAGCTGTCACCG

781 -----+-----+-----+-----+-----+-----+ 840

TACCTGTACACAgGCAAACCTATGCCATCGCGCGTGGTCGCTGCGTTCAGTCGACAGTGGC

F C Q L F T H N E W K K Y D Y L Q S L G 300

TTCTGTCAAACCTTCACTCACAATGAGTGGAAGAAGTACgACTACCTTCAGTCCTTGGGC

841 -----+-----+-----+-----+-----+-----+ 900

AAGACAGTTGAGAAAGTGAGTGTTACTCACCTTCTTCATGcTGATGGAAGTCAGGAACCCG

K Y Y G Y G A G N P L G P A Q G I G F T 320

AAGTACTACGGCTACGGCGCAGGCAACCCTCTGGGACCGGCTCAGGGGATAGGGTTCACC

901 -----+-----+-----+-----+-----+-----+ 960

TTCATGATGCCGATGCCGCGTCCGTTGGGAGACCCTGGCCGAGTCCCCTATCCCAAGTGG

N E L I A R L T R S P V Q D H T S T N S

340

AACGAGCTGATTGCCCCGGTTGACgCGTTGCCAGTGCAGGACCACACCAGCACTAACTCG

961 -----+-----+-----+-----+-----+-----+ 1020

TTGCTCGACTAACGGGCCAACTGcGCAAGCGGTCACGTCCTGGTGTGGTCGTGATTGAGC

T L V S N P A T F P L N A T M Y V D F S

360

ACTCTAGTCTCCAACCCGGCCACCTTCCCGTTGAACGCTACCATGTACGTCGACTTTTCA

1021 -----+-----+-----+-----+-----+-----+ 1080

TGAGATCAGAGGTTGGGCCGGTGGAAGGGCAACTTGCGATGGTACATGCAGCTGAAAAGT

H D N S M V S I F F A L G L Y N G T E P

380

CACGACAACAGCATGGTTTCCATCTTCTTTGCATTGGGCCTGTACAACGGCACTGAACCC

1081 -----+-----+-----+-----+-----+-----+ 1140

GTGCTGTTGTCGTACCAAAGGTAGAAGAAACGTAACCCGGACATGTTGCCGTGACTTGGG

L S R T S V E S A K E L D G Y S A S W V  
 400  
 TTGTCCCGGACCTCGGTGGAAAGCGCCAAGGAATTGGATGGGTATTCTGCATCCTGGGTG  
 1141 -----+-----+-----+-----+-----+  
 1200  
 AACAGGGCCTGGAGCCACCTTTCGCGGTTCCCTAACCTACCCATAAGACGTAGGACCCAC  
 V P F G A R A Y F E T M Q C K S E K E P  
 420  
 GTGCCTTTCGGCGCGCGAGCCTACTTCGAGACGATGCAATGCAAGTCGGAAAAGGAGCCT  
 1201 -----+-----+-----+-----+-----+  
 1260  
 CACGGAAAGCCGCGCGCTCGGATGAAGCTCTGCTACGTTACGTTACGCCTTTTCCTCGGA  
 L V R A L I N D R V V P L H G C D V D K  
 440  
 CTTGTTCGCGCTTTGATTAATGACCGGGTTGTGCCACTGCATGGCTGCGATGTGGACAAG  
 1261 -----+-----+-----+-----+-----+  
 1320  
 GAACAAGCGCGAAACTAATTACTGGCCCAACACGGTGACGTACCGACGCTACACCTGTTC  
 L G R C K L N D F V K G L S W A R S G G  
 460  
 CTGGGGCGATGCAAGCTGAATGACTTTGTCAAGGGATTGAGTTGGGCCAGATCTGGGGGC  
 1321 -----+-----+-----+-----+-----+  
 1380  
 GACCCCGCTACGTTCGACTTACTGAAACAGTTCCCTAACTCAACCCGGTCTAGACCCCGG

N W G E C F S \* 467

AACTGGGGAGAGTGCTTTAGTGA

1381 -----+-----+----- 1404

TTGACCCCTCTCACGAAATCAACT

Figure 11

CP-1

Eco RI M G V F V V L L S I A T L F G S T

**TATATGAATTCATGGGCGTGTTCGTGCTACTGTCCATTGCCACCTGTTTCGGTTCCA**

1 -----+-----+-----+-----+-----+-----+ 60

ATATACTTAAGTACCCGCACAAGCAGCACGATGACAGGTAACGGTGGAACAAGCCAAGGT

S G T A L G P R G N S H S C D T V D G G

**CATCCGGTACCGCCTTGGGTCCTCGTGGTAATTCTCACTCTTGTGACACTGTTGACGGTG**

61 -----+-----+-----+-----+-----+-----+ 120

GTAGGCCATGGCGGAACCCAGGAGCACCATTAAAGAGTGAGAACACTGTGACAACTGCCAC

CP-2

CP-3

Y Q C F P E I S H L W G Q Y S P Y F S L

GTTACCAATGTTTCCCAGAAATTTCTCACTTGTGGGGTCAATACTCTCCATACTTCTCTT

121 -----+-----+-----+-----+-----+-----+ 180

**CAATGGTTACAAAGGGTCTTTAAAGAGTGAACACCCCAGTTATGAGAGGTATGAAGAGAA**

E D E S A I S P D V P D D C R V T F V Q

**TGGAAGACGAATCTGCTATTTCTCCAGACGTTCCAGACGACTGTAGAGTTACTTTCGTTT**

181 -----+-----+-----+-----+-----+-----+ 240

ACCTTCTGCTTAGACGATAAAGAGGTCTGCAAGGTCTGCTGACATCTCAATGAAAGCAAG

CP-4.7

CP-5.7

V L S R H G A R Y P T D S K G K K Y S A

AAGTTTTGTCTAGACACGGTGCTAGATACCCAAGTgacTCTAAGggtAAGaagTACTCTG  
 241 -----+-----+-----+-----+-----+-----+ 300  
 TTCAAAACAGATCTGTGCCACGATCTATGGGTTGActgAGATTCCcaTTcTtcATGAGAC  
  
 L I E A I Q K N A T A F K G K Y A F L K  
 CTTTGATTGAAGCTATTCAAAAGAACGCTACTGCTTTCAAGGGTAAGTACGCTTTCTTGA  
 301 -----+-----+-----+-----+-----+-----+ 360  
 GAAACTAACTTCGATAAGTTTTCTTGCGATGACGAAAGTTCCCATTTCATGCGAAAGAACT

CP-6

CP-7

T Y N Y T L G A D D L T P F G E N Q M V  
 AGACTTACAACCTACACTTTGGGTGCTGACGACTTGACTCCATTTCGGTGAAAACCAAATGG  
 361 -----+-----+-----+-----+-----+-----+ 420  
 TCTGAATGTTGATGTGAAACCCACGACTGCTGAACTGAGGTAAGCCACTTTTGGTTTACC  
  
 N S G I K F Y R R Y K A L A R K I V P F  
 TTAACCTCTGGTATTAAGTTCTACAGAAGATACAAGGCTTTGGCTAGAAAGATTGTTCCAT  
 421 -----+-----+-----+-----+-----+-----+ 480  
 AATTGAGACCATAATTCAAGATGTCTTCTATGTTCCGAAACCGATCTTTCTAACAAGGTA

CP-8.7

CP-9

I R A S G S S R V I A S A E K F I E G F  
 TCATTAGAGCTTCTGGTTCTtctAGAGTTATTGCTTCTGCTGAAAAGTTCATTGAAGGTT  
 481 -----+-----+-----+-----+-----+-----+ 540  
 AGTAATCTCGAAGACCAAGAagaTCTCAATAACGAAGACGACTTTTCAAGTAACCTTCAA

Q S A K L A D P G S Q P H Q A S P V I D  
 TCCAATCTGCTAAGTTGGCTGACCCAGGTTCTCAACCACACCAAGCTTCTCCAGTTATTG  
 541 -----+-----+-----+-----+-----+-----+ 600  
 AGGTTAGACGATTCAACCGACTGGGTCCAAGAGTTGGTGTGGTTCGAAGAGGTCAATAAC

CP-10.7CP-11.7

V I I S E A S S Y N N T L D P G T C T A



ACGTTATTATTtctGAcgetTCTtctTACAACAACACTTTGGACccaGGTACTTGTACTG  
601 -----+-----+-----+-----+-----+-----+ 660  
TGCAATAATAAagaCTgccgaAGGgagaATGTTGTTGTGAAACCTGggtCCATGAACATGAC

F E D S E L A D T V E A N F T A L F A P  
 CTTTCGAAGACTCTGAATTGgctGACactGTTGAAGCTAACTTCACTGCTTTGTTTCGCTC

661 -----+-----+-----+-----+-----+-----+-----+ 720

GAAAGCTTCTGAGACTTAACcgaCTGtgaCAACTTCGATTGAAGTGACGAAACAAGCGAG

CP-12.7

A I R A R L E A D L P G V T L T D T E V  
 CAGCTATTAGAGCTAGATTGGAAGCTGACTTGCCAGGTGTTACTTTGACTGACactgaaG

721 -----+-----+-----+-----+-----+-----+-----+ 780

GTCGATAATCTCGATCTAACCTTCGACTGAACGGTCCACAATGAAACTGACTGtgacttC

CP-13.7

T Y L M D M C S F E T V A R T S D A T E  
 TTactTACTTGATGGACATGTGTtctTTCGAAACTGTTGCTAGAACTTCTGACGCTACTG

781 -----+-----+-----+-----+-----+-----+-----+ 840

AATgaATGAACTACCTGTACACAagaAAGCTTTGACAACGATCTTGAAGACTGCGATGAC

L S P F C A L F T H D E W R H Y D Y L Q  
 AATTGTCTCCATTCTGTGCTTTGTTCACTCACGACGAATGGAGAcacTACGACTACTTGC

841 -----+-----+-----+-----+-----+-----+-----+ 900

TTAACAGAGGTAAGACACGAAACAAGTGAGTGCTGCTTACCTCTgtgATGCTGATGAACG

CP-14.7

CP-15.7

S L K K Y Y G H G A G N P L G P T Q G V  
 AATCTTTGaagAAGTACTACGGTcacGGTGCTGGTAACCCATTGGGTCCAactCAAGGTG

901 -----+-----+-----+-----+-----+-----+-----+ 960

TTAGAAACttcTTCATGATGCCAgtgCCACGACCATTGGGTAACCCAGGTtgaGTTCCAC

G F A N E L I A R L T R S P V Q D H T S  
 TTGGTTTCGCTAACGAATTGATTGCTAGATTGACTAGATCTCCAGTTCAAGACCACACTT

961 -----+-----+-----+-----+-----+-----+-----+  
 1020

AACCAAAGCGATTGCTTAACTAACGATCTAACTGATCTAGAGGTCAAGTTCTGGTGTGAA

CP-16

CP-17.7

T N H T L D S N P A T F P L N A T L Y A  
 CTACTAACCACACTTTGGACTCTA~~ACC~~CAGCTACTTTCCCATTTGAACGCTACTTTGTACG  
 1021 -----+-----+-----+-----+-----+-----+  
 1080  
 GATGATTGGTGTGAAACCTGAGATTGGGTCGATGAAAGGGTAACTTGCATGAAACATGC  
 D F S H D N G I I S I F F A L G L Y N G  
 CTGACTTCTCTCACGACAACggtattATTTCTATTTTCTTCGCTTTGGGTTTGTACAACG  
 1081 -----+-----+-----+-----+-----+-----+  
 1140  
 GACTGAAGAGAGTGCTGTTGccataaTAAAGATAAAAGAAGCGAAACCCAAACATGTTGC  
 CP-18.7  
 CP-19.7  
 T A P L S T T S V E S I E E T D G Y S S  
 GTACTGCTCCATTGTCTACTACTTCTGTTGAATCTATTGAAGAACTGACGGTTACTCTt  
 1141 -----+-----+-----+-----+-----+-----+  
 1200  
 CATGACGAGGTAACAGATGATGAAGACAACCTTAGATAACTTCTTTGACTGCCAATGAGAA  
 A W T V P F A S R A Y V E M M Q C Q A E  
 ctgctTGGACTGTTCCATTTCgcttctAGAGCTTACGTTGAAATGATGCAATGTCAAGCTG  
 1201 -----+-----+-----+-----+-----+-----+  
 1260  
 gacgaACCTGACAAGGTAAGcgaagaTCTCGAATGCAACTTTACTACGTTACAGTTTCGAC  
 CP-20  
 CP-21  
 K E P L V R V L V N D R V V P L H G C A  
 AAAAGGAACCATTTGGTTAGAGTTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTG  
 1261 -----+-----+-----+-----+-----+-----+  
 1320  
 TTTTCCTTGGTAACCAATCTCAAAACCAATTGCTGTCTCAACAAGGTAACGTGCCAACAC

V D K L G R C K R D D F V E G L S F A R  
**CTGTTGACAAGTTGGGTAGATGTAAGAGAGACGACTTCGTTGAAGGTTTGTCTTTCGCTA**  
1321 -----+-----+-----+-----+-----+-----+-----+  
1380  
GACAACTGTTCAACCCATCTACATTCTCTCTGCTGAAGCAACTTCC**AAACAGAAAGCGAT**  
S G G N W A E C F A \* *Eco* RI CP-22  
**GATCTGGTGGTAACTGGGCTGAATGTTTCGCTTAAGAATTCATATA**  
1381 -----+-----+-----+-----+-----+----- 1426  
**CTAGACCACCATTGACCCGACTTACAAAGCGAATTCTTAAGTATAT**

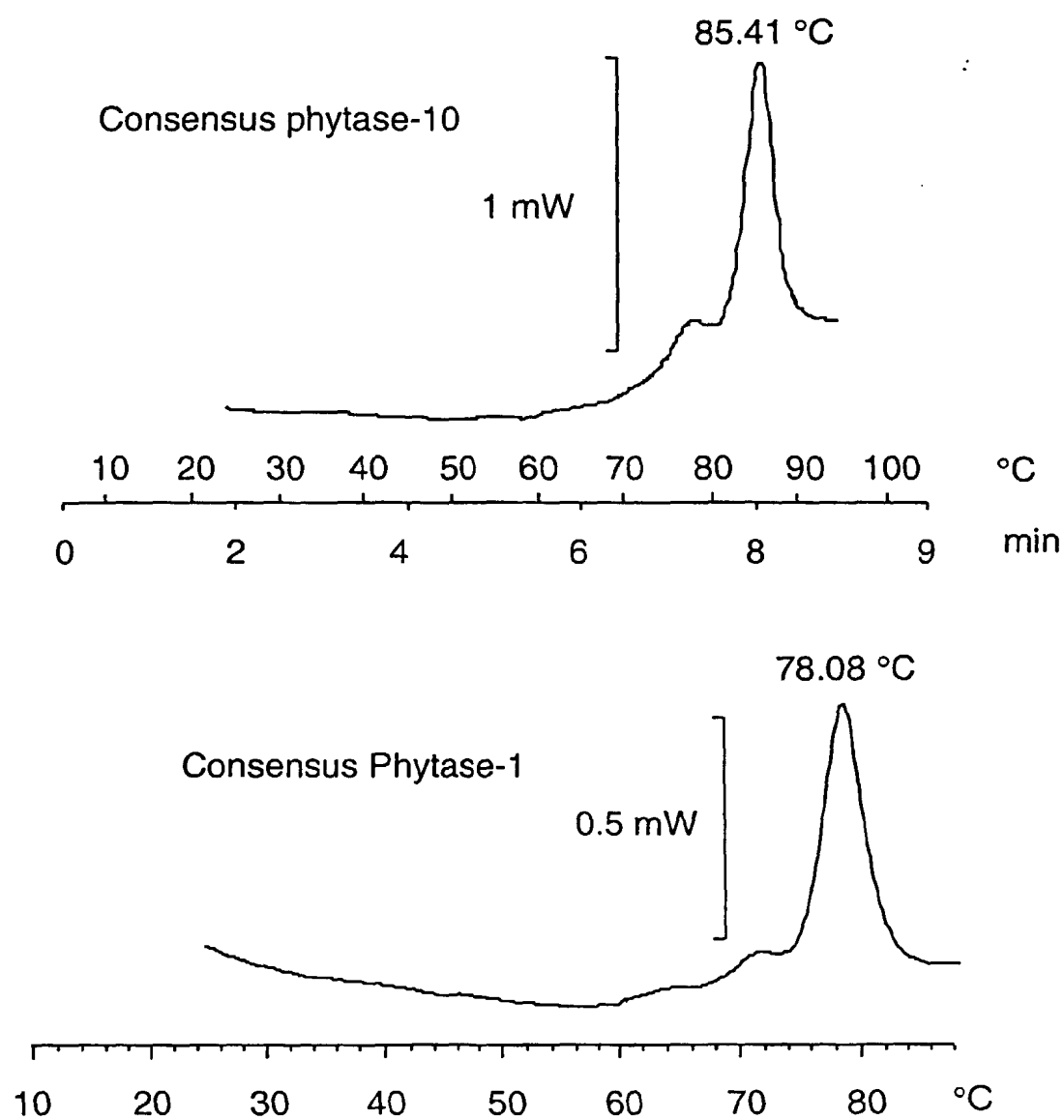
Figure 12

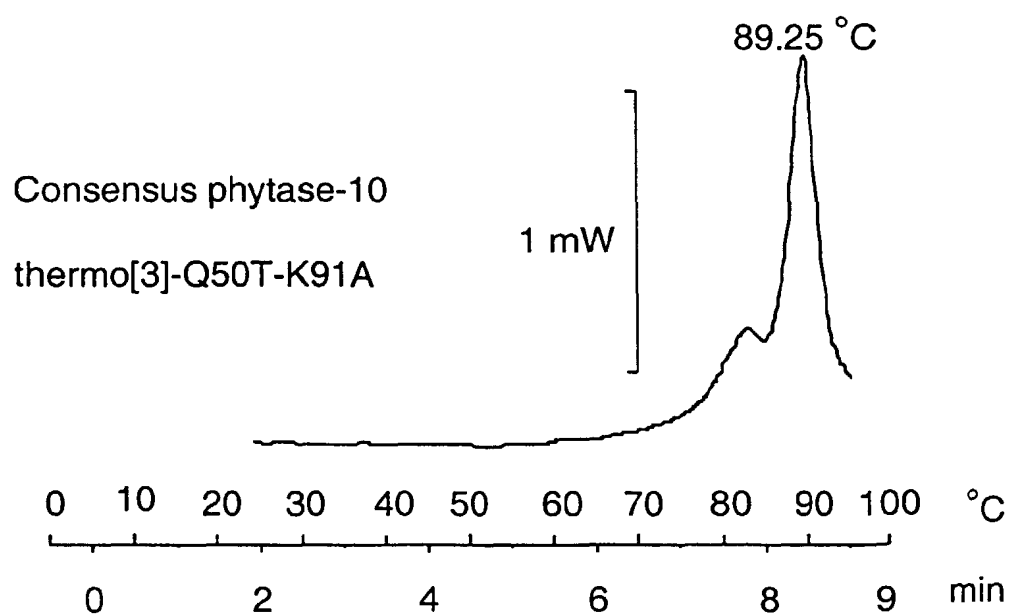
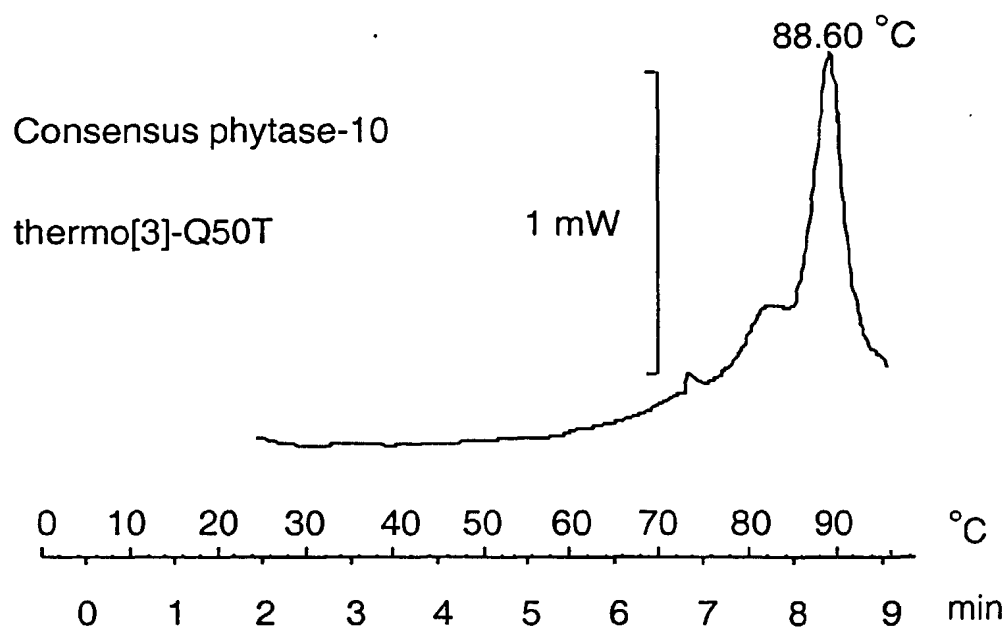
Figure 13

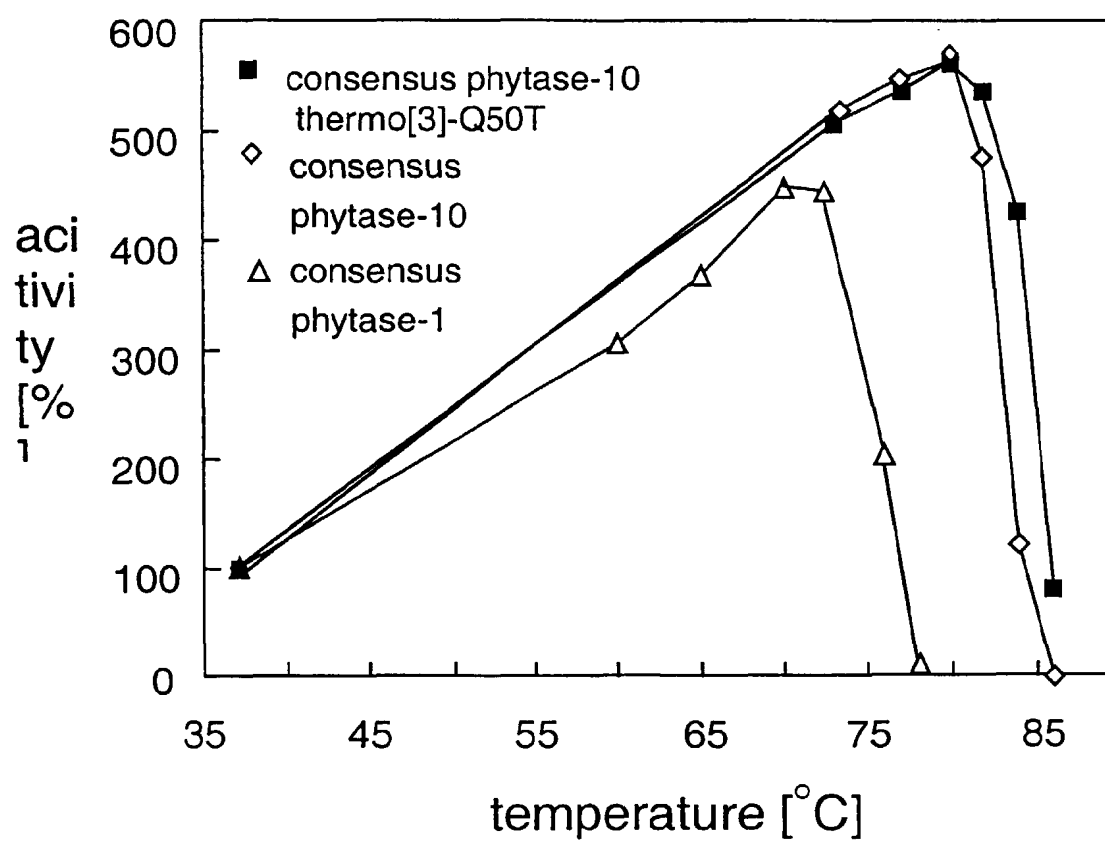
Figure 14

Figure 15

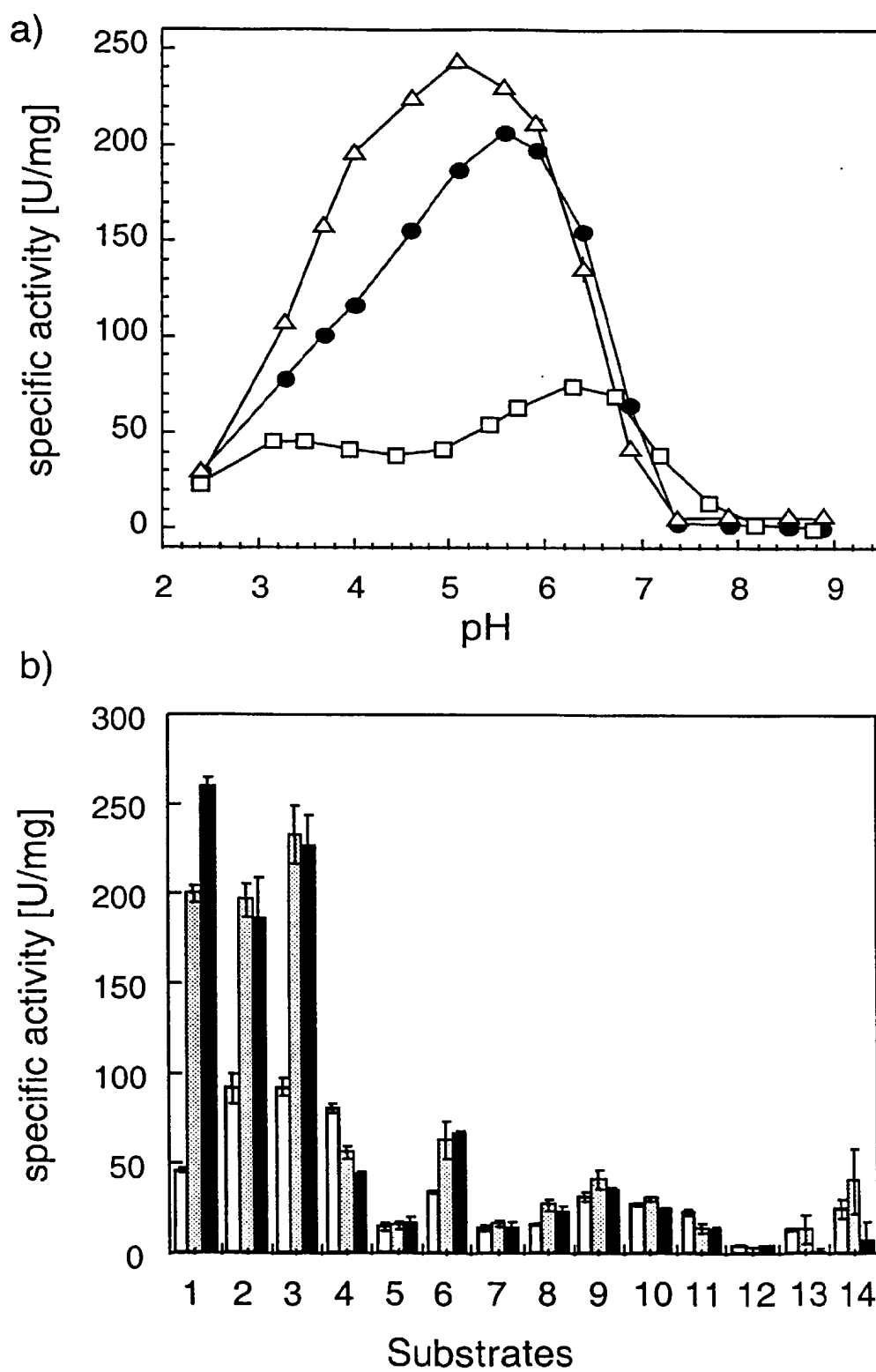




Figure 16

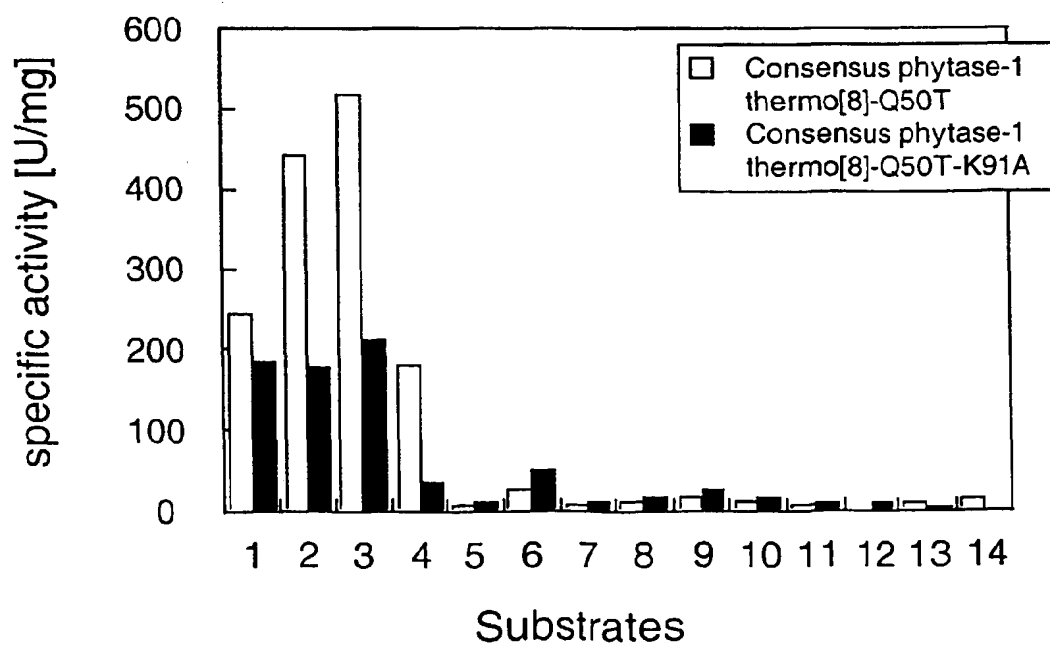
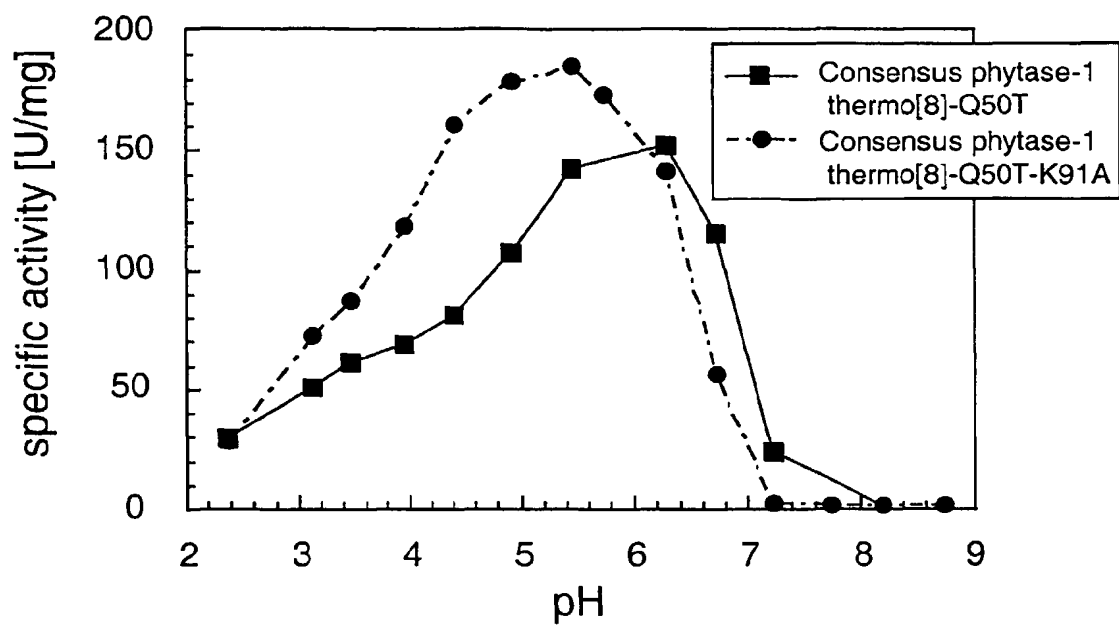


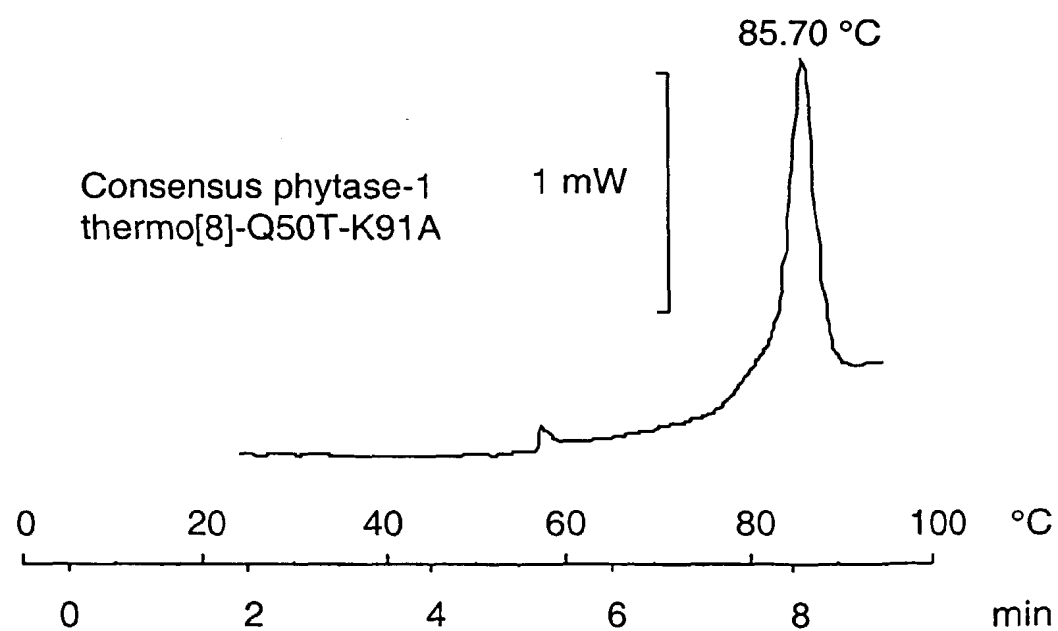
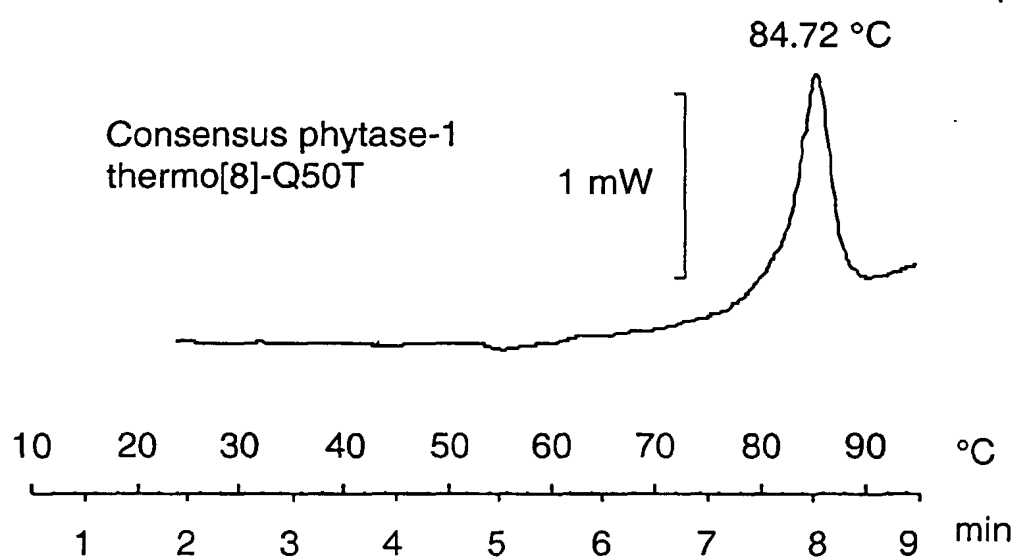
Figure 17

Figure 18

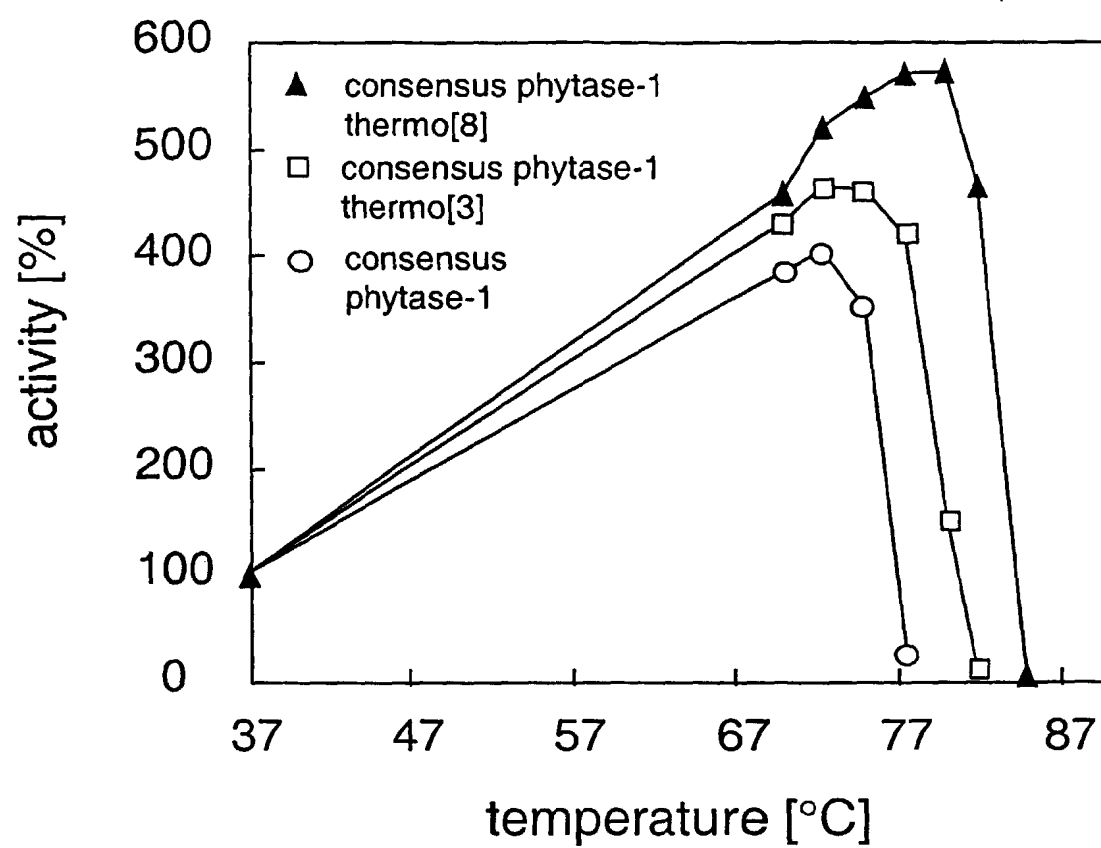


Figure 19

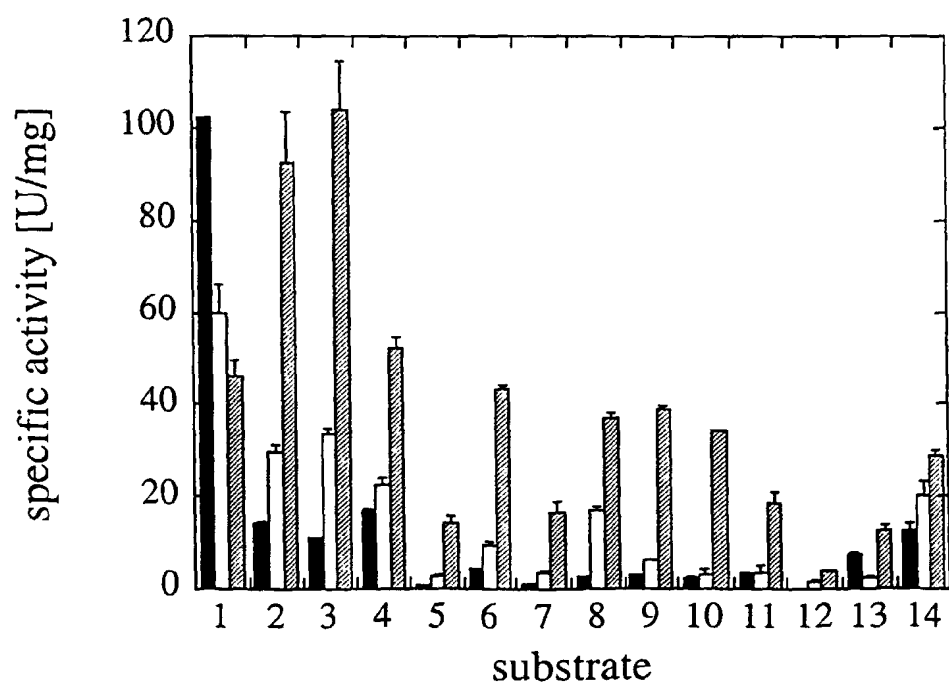
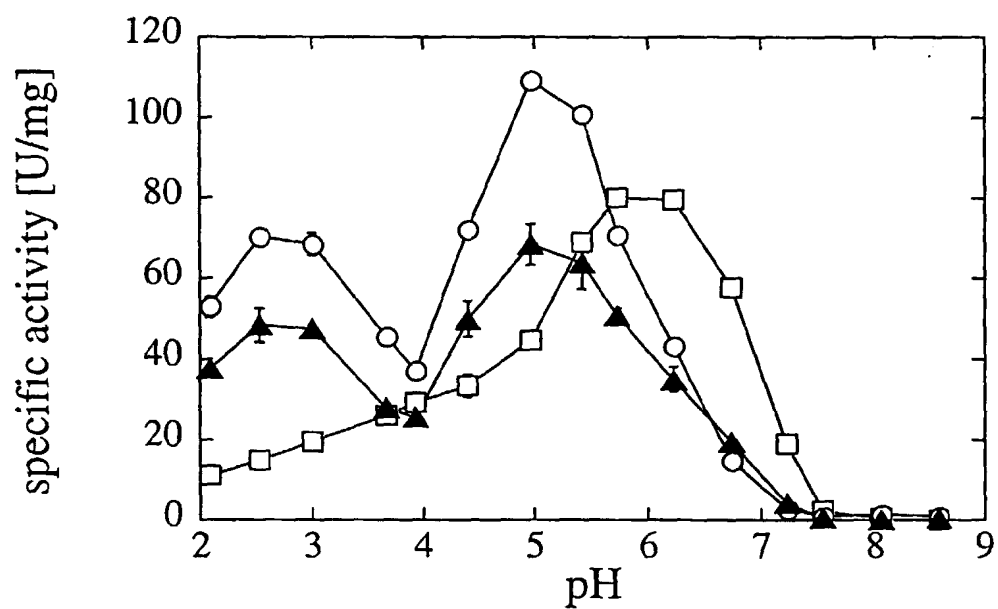


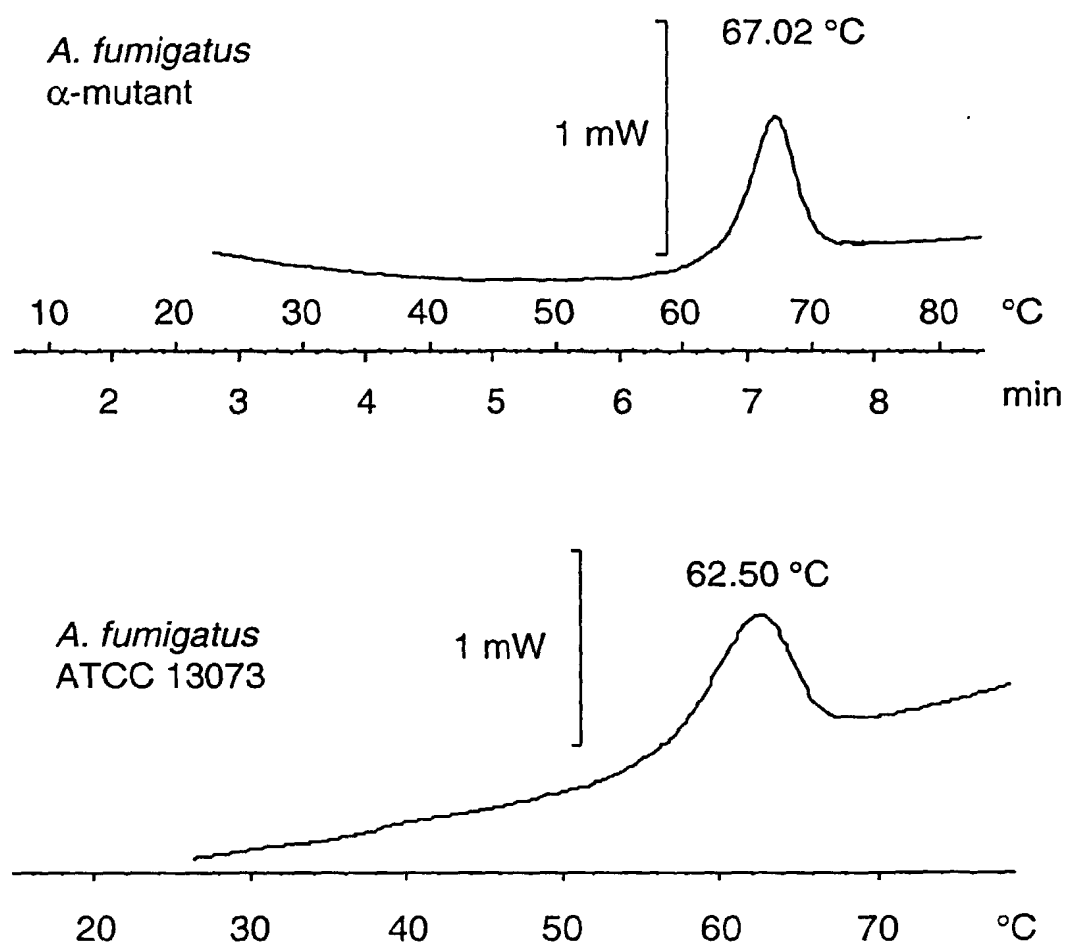
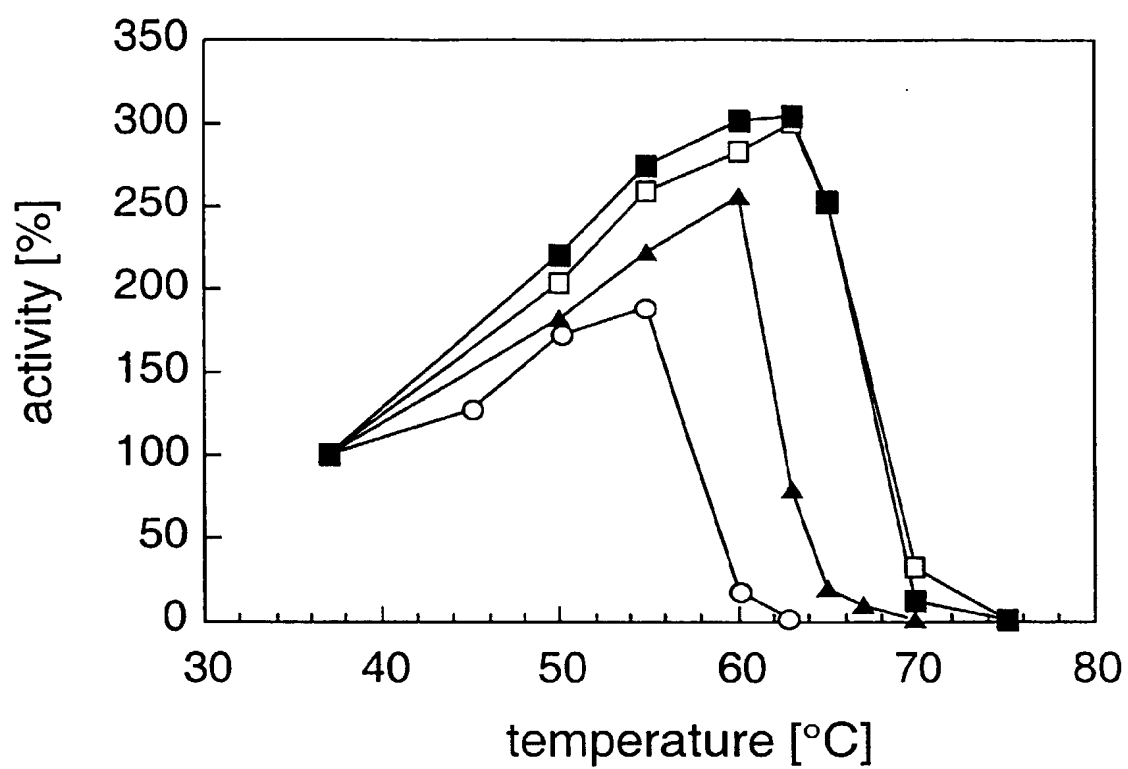
Figure 20

Figure 21



**Figure 22**

1 MGVFVLLSI ATLFGSTSGT ALGPRGNSHS CDTV~~D~~GGYQC FPEISS~~N~~W~~S~~P  
 51 YSPY~~F~~SLADE SAISPDV~~P~~KG CRVTFVQVL~~Q~~ RHGARE~~F~~PTSG AATR~~I~~SALIE  
 101 AIQKNATAFK GKYAFLKTYN YTLGADDL~~V~~P FGAN~~Q~~SS~~Q~~AG IKFYRRYKAL  
 151 ARKIVPFIRA SGSDRVID~~S~~A TNWIEGFQSA KLADPGANPH QASPVINVII  
 201 PEGAGYNNTL DHGLCTAFEE SELGDDVEAN FTAVFAPP~~I~~R ARLEAHLPGV  
 251 NLTDEDVVNL MDMCPFD~~T~~VA RTSDATE~~L~~SP FCDLFTHDEW IQYDYL~~G~~D~~L~~D  
 301 KYYGT~~I~~GAGNP LGPAQGVGFV NELIARLTHS PVQDHTSTNH TLDSNPATFP  
 351 LNATLYADES HDNTMV~~A~~IFF ALGLYNGTKP LSTTSVESIE ETDGYSASW~~L~~  
 401 VPF~~S~~ARM~~Y~~VE MMQCEAEKEP LVRVLVND~~R~~V VPLHGCGVDK LGRCKRDDFV  
 451 EGLSFARSGG NWE~~E~~CFA